

**SYNTHESIS OF ALLYL ALCOHOLS BY
PALLADIUM-CATALYZED 1,3-SUBSTITUTION
REACTIONS OF ALKENYL EPOXIDES WITH
ORGANOBORONS**

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ABBREVIATIONS

Ac	Acetate
aq.	Aqueous
BINAP	2,2'-Bis(diphenylphosphino)-1,1'-binaphthyl
Bn	Benzyl
Bu	n-Butyl
<i>t</i> -Bu	tert-Butyl
Cy	Cyclohexane
dba	Dibenzylideneacetone
DCM	Dichloromethane
DIBAL-H	Diisobutylaluminium hydride
DMAP	4-Dimethylaminopyridine
DME	1,2-Dimethoxyethane
DMF	<i>N,N</i> -Dimethylformamide
DPEPhos	Bis-[2-(diphenylphosphino)phenyl]ether
dppb	1,4-bis(diphenylphosphino)butane
dppe	1,2-bis(diphenylphosphino)ethane
dppm	Bis(diphenylphosphino)methane
dppp	1,3-bis(diphenylphosphino)propane
d.r.	Diastereomeric ratio
Et	Ethyl
eqv.	Equivalent
Et ₂ O	Diethyl ether
h	hour
<i>m</i> -CPBA	meta-chloroperoxybenzoic acid
M	Molar
Me	Methyl
mg	Miligrams
min.	Minute
mL	Mililiter
mmol	Milimoles
N.D.	Not determined

neop	Neopentyl glycol ester
NMR	Nuclear magnetic resonance
O.N.	Overnight
Ph	Phenyl
Pr	Propyl
rt	Room Temperature
t	Time
<i>i</i> -Pr	iso-Propyl
SIPr	1,3-bis(2,6-di- <i>i</i> -propylphenyl) imidazol-2-ylidene
TBDMS	tert-Butyldimethylsilyl
THF	Tetrahydrofuran
Xanthphos	4,5-Bis(diphenylphosphino)-9,9-dimethylxanthene
Xphos	2-Dicyclohexylphosphino-2',4',6' triisopropylbiphenyl

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ABSTRACT

SYNTHESIS OF ALLYL ALCOHOLS BY PALLADIUM-CATALYZED 1,3-SUBSTITUTION REACTIONS OF ALKENYL EPOXIDES WITH ORGANOBORONS

1,3-Substitution reactions of allylic compounds having a good leaving group is a prominent method in Organic Chemistry for the synthesis of new allylic reagents with an exchanged functional group. These reactions usually require the use of metal catalysts and one of the most challenging aspects for these applications is the regio- and stereo-selectivity of the process for a wide range of substrate types. Other compounds such as vinyl epoxides are also acceptable for substitution reactions.

An important advantage of using these reagents is that opening of the oxirane ring during the substitution process lead to the generation of a hydroxyl group and as a result affords allyl alcohols which are important intermediaries in organic syntheses.

An example of regio-selective metal-catalyzed reactions of vinyl epoxides having a terminal alkenyl group with environmentally benign organoborons was reported in the literature. However, no such success could be achieved with vinyl epoxides with an internal alkenyl group. Therefore, within the context of this method internal vinyl epoxides were successfully subjected to 1,3-substitution reactions with organoborons which yielded arylated allyl alcohols in both a regio- and stereo-selective manner. The method is applicable under quite mild conditions where a palladium-AsPh₃ combination is used to activate the process.

ÖZET

ALKENİL EPOKSİTLERİN ORGANOBORLAR İLE PALADYUM KATALİZLİ 1,3-SÜBSTİTÜSYON TEPKİMELERİ ARACILIĞI İLE ALLİL ALKOLLERİN SENTEZİ

Kolay bir terkeden gruba sahip allilik bileşiklerin 1,3-sübstütüsyon tepkimeleri farklı bir fonksiyonel gruba sahip yeni allilik bileşiklerin sentezi organik kimyada önemli bir tepkime tipidir. Bu tip tepkimeler çoğunlukla bir metal katalizörü ile gerçekleştirilmektedir. Bu amaçla uygulanan yöntemlerin uygulamasında en önemli zorluk tepkimelerin yeterli mertebede regio ve stereo seçimli olarak geniş bir sübstrat yelpazesini içerecek şekilde gerçekleştirmektir.

Sübstütüsyon tepkimelerine uygun bir allilik bileşikte vinil epoksitlerdir. Bu yapıların kullanımında önemli bir avantaj epoksit oksijenin tepkime sonunda yalnızca bir bağının kopması sonucu hidroksil grubuna dönüşerek organik sentezlerde önemli bir ara yapı olan allil alkollerin oluşmasıdır.

Literatürde terminal alkenil gruba sahip vinil epoksitlerin çevre dostu organoborlar ile metal katalizli tepkimelerinin regio- ve stereo seçimli olarak gerçekleştiği rapor edilmiştir. Ancak aynı başarı iç alkenil gruba sahip yapılar için sağlanamamıştır. Bu nedenle bu tez çalışması kapsamında iç alkenil gruplu vinil epoksitlerin organoboronlar ile regio ve stereo seçimli olarak 1,3-sübstütüsyon tarzında arillenmesi başarılmıştır. Yöntem bir paladyum katalizörü ve $AsPh_3$ ligandının kullanıldığı ılıman koşullar gerektirmektedir.

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CHAPTER 1

INTRODUCTION

The studies about transition-metal-catalyzed reactions with various organometallic reagents have been an increasingly attractive method in recent years. The alkenyl or aryl addition to unsaturated compounds which have at least one leaving group is a well-known procedure. These compounds give generally S_N2 type reaction with a nucleophile in the absence of transition metals. However, the transition metals can change this orientation and the reactions may occur in the S_N2' manner.

The hard nucleophiles are used generally with copper- or nickel-catalyzed reactions over unsaturated compounds in order to shift the selectivity toward S_N2' type reaction. However, the usage of these nucleophiles such as grignard reagents, organolithium, and organozinc compounds have some restrictions, since they are highly air and moisture sensitive.

On the other hand, the organoboron compounds are highly stable to moisture and easy-to-handle without special precautions. These advantages have increased the popularity of organoborons day by day. In 1979, the palladium catalyzed cross-coupling reaction with derivatives of organoboronic acids known as the Suzuki reaction was published by Suzuki and Miyaura (Miyaura, Yamada, and Suzuki 1979). In a short interval, a variety of organoboron compounds began to be used on different types of coupling and substitution reactions over transition metals. Generally, the 1,3- selective substitution of propargylic compounds is performed to produce allenyl compounds (Moriya, Miyaura and A. 1994). And also in 2011, the palladium catalyzed S_N2'' -selective reaction with boronic acids was first performed on 2-en-4-yne carbonate structures (Üçüncü et al. 2011).

Inspired of these information, in contrast to large number of reports about allylic alkylations, there are only a few allylic arylation reactions over the alkenyl compounds in the literature. In these works, the hard reaction conditions were required because the air sensitive compounds like grignard reagents and organozinc compounds were used as the nucleophile. Therefore, we performed palladium(0)- catalyzed 1,3-selective allylic

arylation reaction on vinyl oxiranes to provide arylated allyl alcohols under mild conditions.

CHAPTER 2

LITERATURE WORKS

2.1. Transition- Metal-Catalyzed Allylic Arylation Reaction of Allylic Compounds

The alkenyl compounds play an important role in organic synthesis. 1,3 Substitution reaction is a useful method for the formation of allylic reagents with a variety of functional groups. In literature, there are vast number of studies on highly efficient allylic alkylation reactions on vinyl compounds. But there are a few works about the allylic arylation reaction on alkenyl compounds and the regio selectivity of these reactions were not effectiveness.

In 1994, Bäckvall reported highly regio-selective Cu-catalyzed S_N2' -type arylation with Grignard reagents. In the absence of the catalyst the selectivity shifted in the favor of S_N2 pathway, the ratio of $S_N2:S_N2'$ being 91:9 in this case (Figure 2.1).(Bäckvall, Persson, and Bombrun 1994)

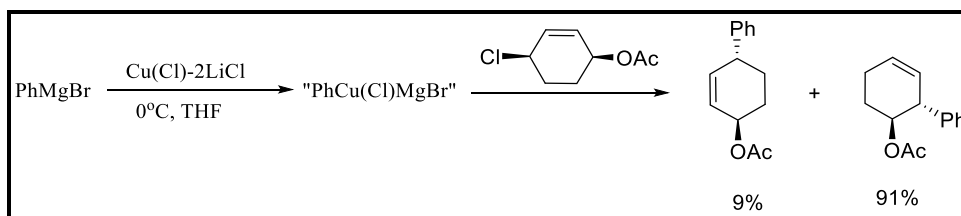


Figure 2.1. Cu(I)-catalyzed arylation reaction of an alkenyl chloride with a Grignard Reagent (Source: Bäckvall, Persson, and Bombrun 1994).

After this invention on cyclic alkenyl compounds, Kacprzynski reported the copper catalyzed arylation on silyl substituted alkenyl phosphorous esters with arylzinc reagents. To achieve the enantio- and regio-selective process, it was assisted from the use of N-heterocyclic carbene (NHC) ligands (Figure 2.2).(Kacprzynski et al. 2007)

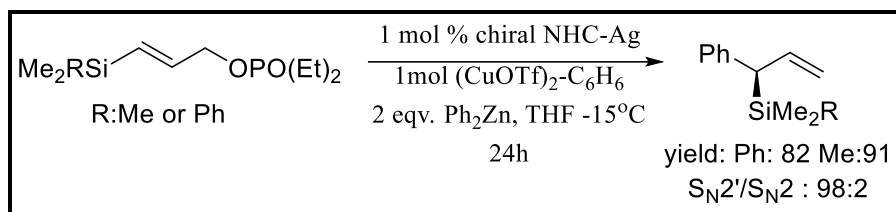


Figure 2.2. Copper-catalyzed arylation reaction of silyl substituted alkenyl phosphono esters (Source: Kacprzynski et al. 2007).

An asymmetric method on allylic arylation of non-cyclic allyl halide compounds were introduced by Selim (Selim, Yamada, and Tomioka 2008). This work shows that the copper-catalyzed stereo-selective arylation reaction can be applied to the non-cyclic allyl compounds with moderate regioselectivities. Interestingly, however, all the improvements of the regio-selectivity always resulted in a reduced enantio-selectivity (Figure 2.3).

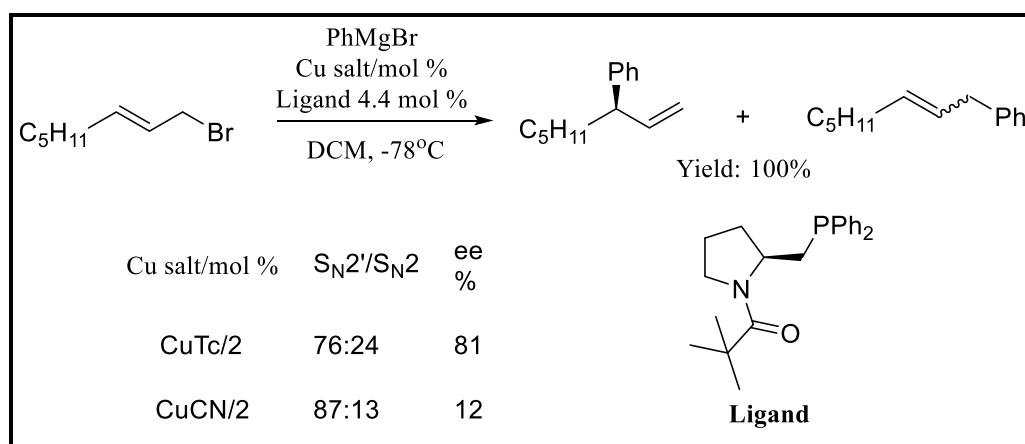


Figure 2.3. Copper catalyzed reaction of a non-cyclic allyl bromide with phenyl grignard reagent (Source: Selim, Yamada, and Tomioka 2008).

Selim also published another paper about stereo- and regio-selective reactions of phenyl substituted allyl compounds after 1 year. They achieved the asymmetric arylation with NHC based copper catalyst at high regio- and enantio-selectivities (Figure 2.4) (Selim et al. 2009).

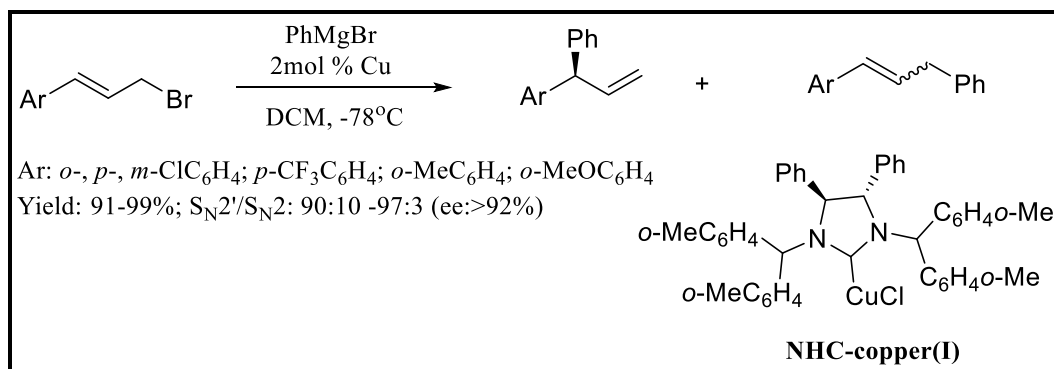


Figure 2.4. NHC-Copper(I) catalyzed reaction of phenyl substituted alkenyl compound (Source: Selim et al. 2009).

Enantio-selective iridium-catalyzed allylic arylation reaction was applied on the aryl substituted allyl compounds having a carbonate group as a leaving group by Polet (Polet et al. 2009). Though this method afforded the molecules in high ee percentages, the regio-selectivities were exceedingly poor (Figure 2.5).

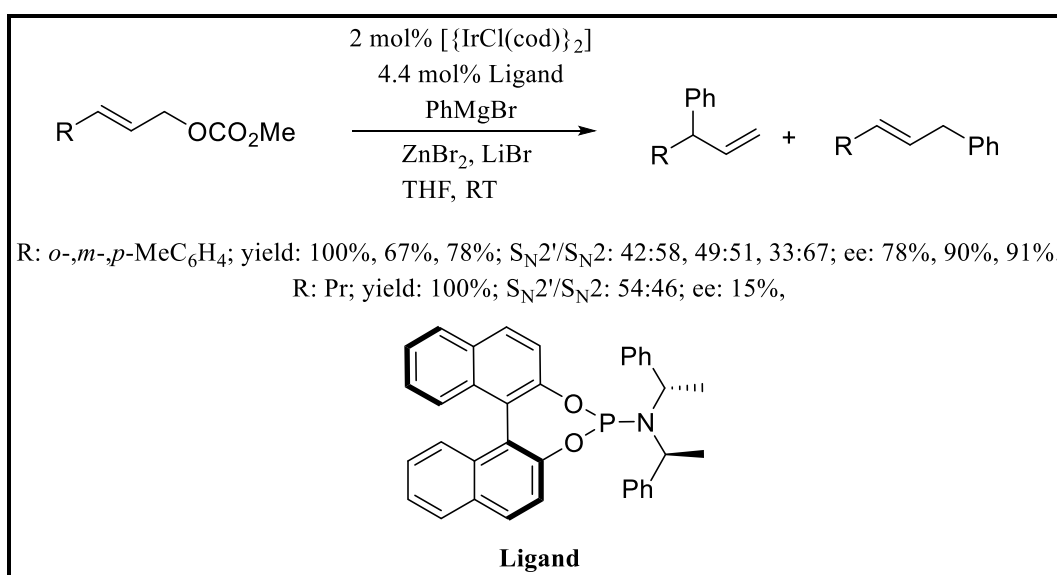


Figure 2.5. Iridium-catalyzed allylic arylation (Source: Polet et al. 2009).

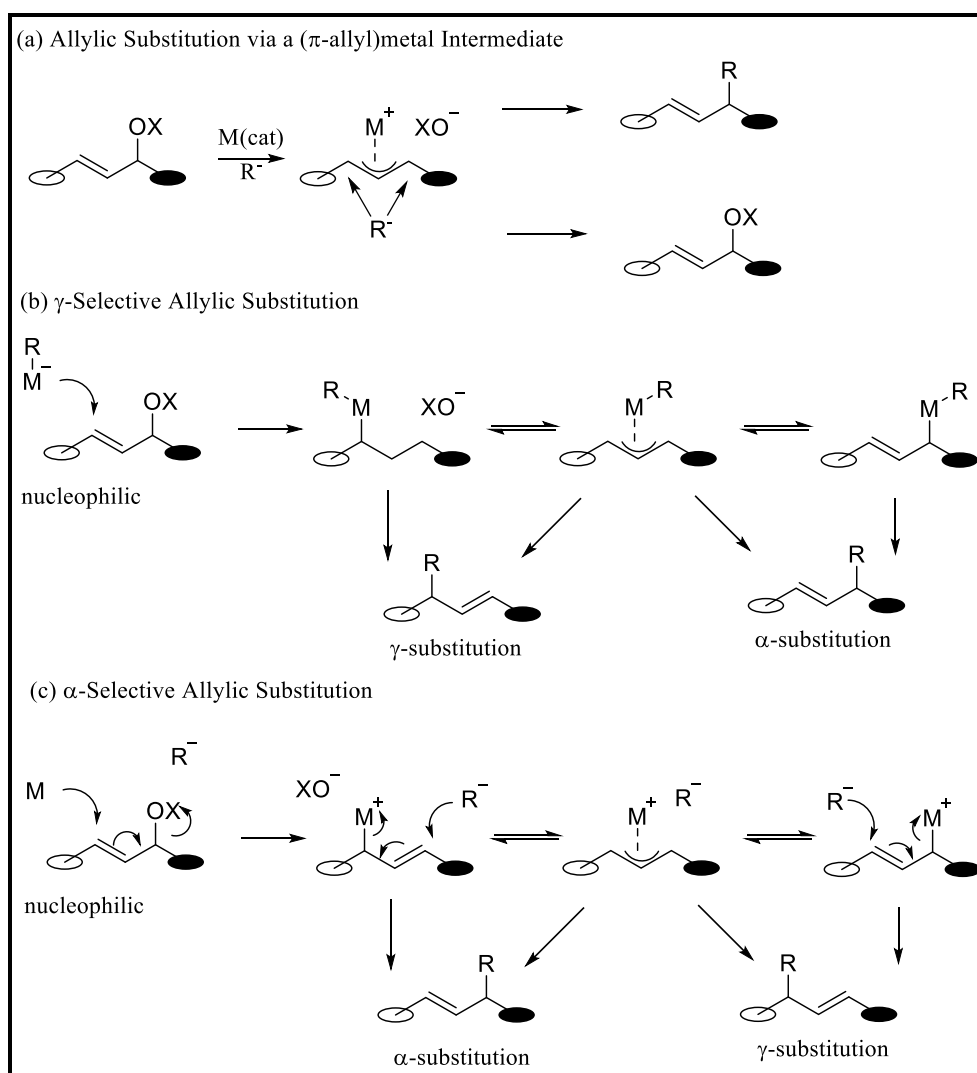


Figure 2.6. Metal-Mediated Allylic Substitution
(Source: Ohmiya, Yokokawa, and Sawamura 2010)

Before the Ohmiya paper, there is three type of metal-mediated allylic substitutions mechanisms (Figure 2.6). First one is the allylic substitution involving a (π -allyl) metal intermediate. However, in this mechanism, the regioselectivity is highly dependent on the electronic and/or steric substituent effects of compounds (Trost and Crawley 2003). In second example mechanism, the formal S_N2' attack of the organometallic nucleophile which are generally monoalkyl heterocuprate reagents produces a (σ -allyl) metal species via oxidative addition the result of the reductive elimination, the C-C bond at the γ -position was formed. But in some examples of these type mechanism, the significant S_N2 product was formed (Yoshikai, Zhang, and Nakamura 2008). In the last example, (σ -allyl) metal complexes was formed after strongly nucleophilic translation metal complex attacked the allylic substrates. The

complex underwent a second allylic displacement with a carbon nucleophile. After reductive elimination of metal complexes, the α or γ selective substitution products were occurred.

Hirohisa et al. reported Pd(II)-catalyzed arylation of allylic acetates with arylboronic acids in 2010 (Ohmiya et al. 2010). Their method was highly γ -selective (Figure 2.7), (Figure 2.8). They proposed a plausible mechanism to account for this high selectivity as is outlined in Figure 2.8., The reaction starts with the formation of cationic mono(acetoxo)-palladium(II) complex **A** which is the product of the 1,10-phenanthroline-ligated Pd(OAc)₂ and AgSbF₆. The complex **A** involves with transmetalation with arylboronic acid to afford the complex **B**. In next step, **B** interact with the allylic acetate which is shown with **C**. Then, this interaction undergoes regio-selective C-C double bond insertion into the aryl-Pd bond (carbopalladation) in order to form a metallacyclic alkylpalladium (II) complex **D**. Finally, β -acetoxy elimination occur and the phenyl substituted product is formed. They also noted that the attempts to use a pinacolato ester instead of boronic acid resulted in no reaction.

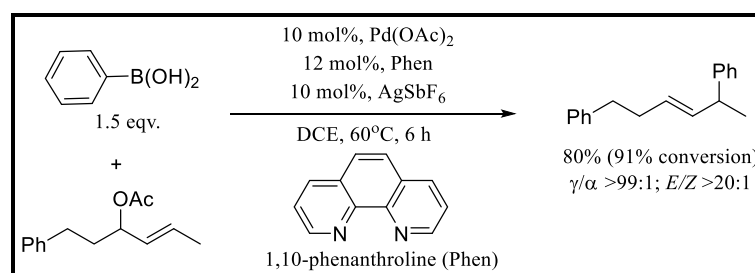


Figure 2.7. Palladium-catalyzed reaction of allylic acetates with arylboronic acids (Source: Ohmiya et al. 2010)

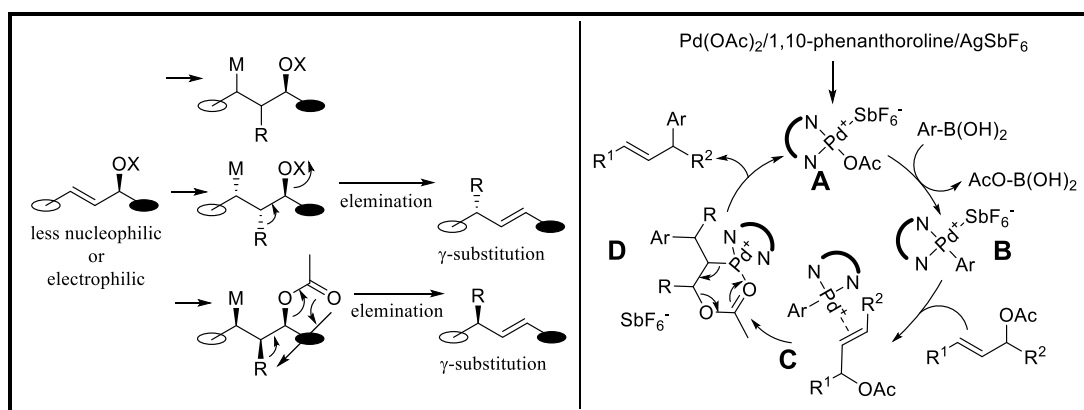


Figure 2.8. Scenarios for new type of γ -selective allylic substitution & proposed mechanism for the reaction (Source: Ohmiya et al. 2010)

In 2010, Whittaker report the copper-catalyzed 1,3-selective substitution of allylic chlorides with arylboronic esters (Whittaker, Rucker, and Lalic 2010). The use of arylboronic acid pinacol ester in the copper(I)-catalyzed reaction provided higher regioselectivity as compared to those used hard nucleophiles, such as phenylmagnesium halides and arylzinc compounds. This reaction was applied with NHC-ligand and the regioselectivity was well up to 99% for 18 examples (Figure 2.9).

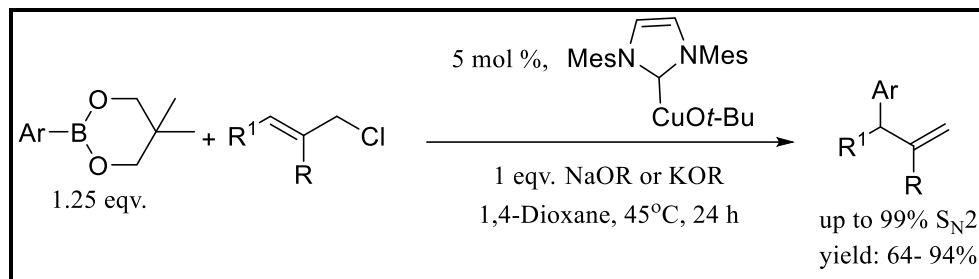


Figure 2.9. Copper catalyzed 1,3-selective substitution of allylic chlorides with arylboronic esters (Source: Whittaker, Rucker, and Lalic 2010).

Allyl phosphono esters could be arylated with arylboronic esters over CuCl/asymmetric NHC ligand with excellent stereo- and regio-selectivities (Figure 2.10) (Takeda et al. 2014).

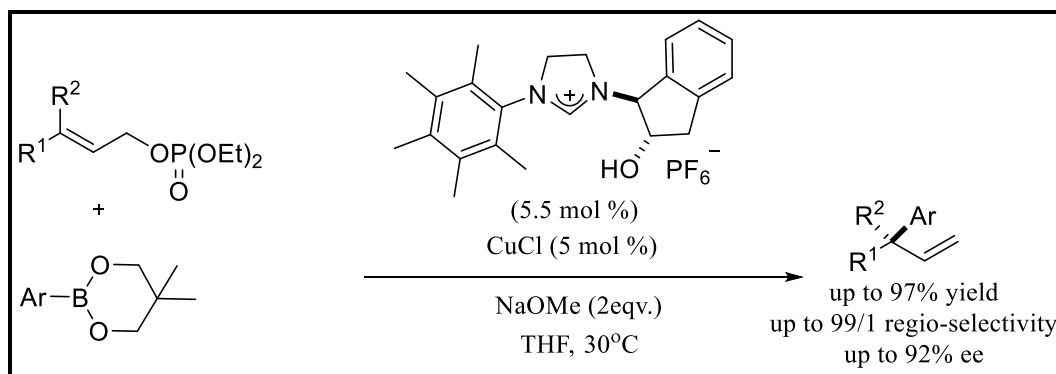


Figure 2.10. Copper-catalyzed asymmetric allylic substitution of allyl phosphates with aryl boronates (Source: Takeda et al. 2014).

2.2. Metal-Catalyzed Reactions of Vinyl Oxiranes with Grignard Reagents

In 2005, Ueki reported the reaction of *gem*-difluorinated vinyl oxiranes with monoalkylcopper reagents (Ueki et al. 2005). In this report, they tried catalytic reaction on the fluorinated vinyl oxiranes but only 43% of reactant was consumed and the regioselectivity of reaction was poor. On the other hand, the non-catalytic trial on this substrate provided good regioselectivity for the S_N2-product and yield (Figure 2.11). When they used the non-fluorinated compound, the regioselectivity of reaction changed to S_N2'-product.

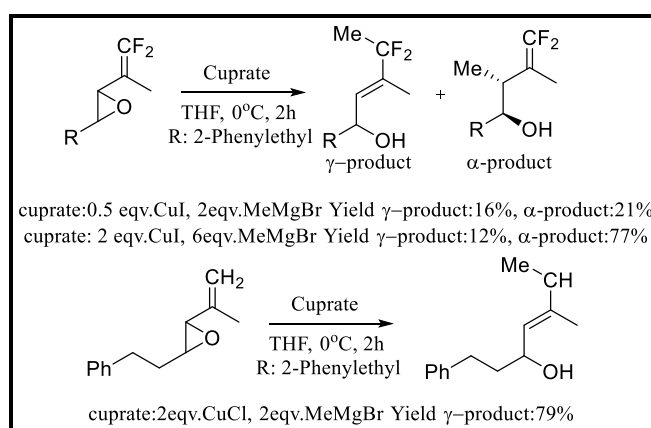


Figure 2.11. The Reaction of alkenyl oxiranes with Grignard reagents (Source: Ueki et al. 2005)

In 2007, the copper catalyzed kinetic resolution of 1,3-cyclohexadiene monoepoxide with Grignard reagents leading to good yields, excellent regioselectivity and good enantiomeric excesses was published by Millet and Alexakis (Millet and Alexakis 2007). But this chiral ferrocenyl-based diphosphine ligand performance provided only 61% ee and moderate regioselectivity for *i*-PrMgCl as used for derivative of secondary Grignard reagents (Figure 2.12).

After one year, Millet and Alexakis published another method for copper catalyzed the kinetic resolution of 1,3-cyclohexadiene monoepoxide with Grignard reagents (Millet and Alexakis 2008). This method shows that SimplePhos ligand is able to perform the kinetic resolution of racemic cyclic vinyloxirane with good regio- and stereo-selectivities and from low to good yields. And also this reaction condition increased the

stereo-selectivity of secondary grignard reagents like *i*-PrMgCl on the S_N2' product (Figure 2.13).

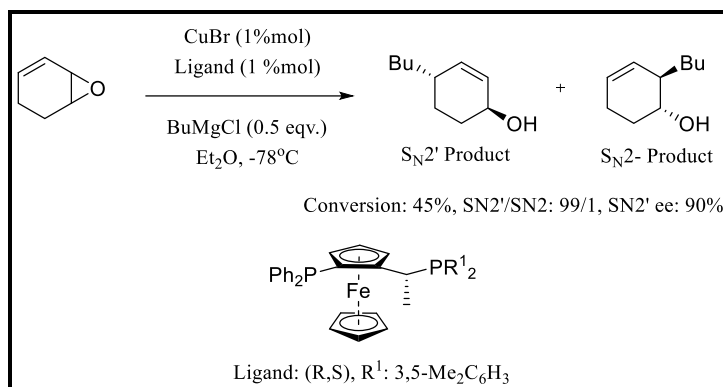


Figure 2.12. Copper catalyzed kinetic resolution of 1,3-Cyclohexadiene monoepoxide with grignard reagents (Source: Millet and Alexakis 2007)

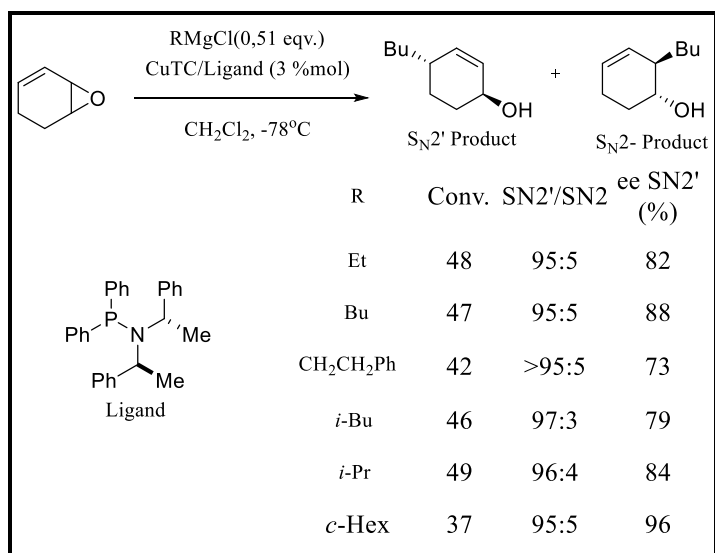


Figure 2.13. SimplePhos as efficient ligand for the copper-catalyzed kinetic resolution of cyclic vinyloxiranes with grignard reagents (Source: Millet and Alexakis 2008)

In 2012, Dieter published the copper catalyzed reaction of silylated-vinyloxirane (Dieter, Huang, and Guo 2012). They showed that the copper catalyzed reactions with alkyl grignard reagents were S_N2' product which were *E* isomers. These reactions also are highly stereo-selective (Figure 2.14).

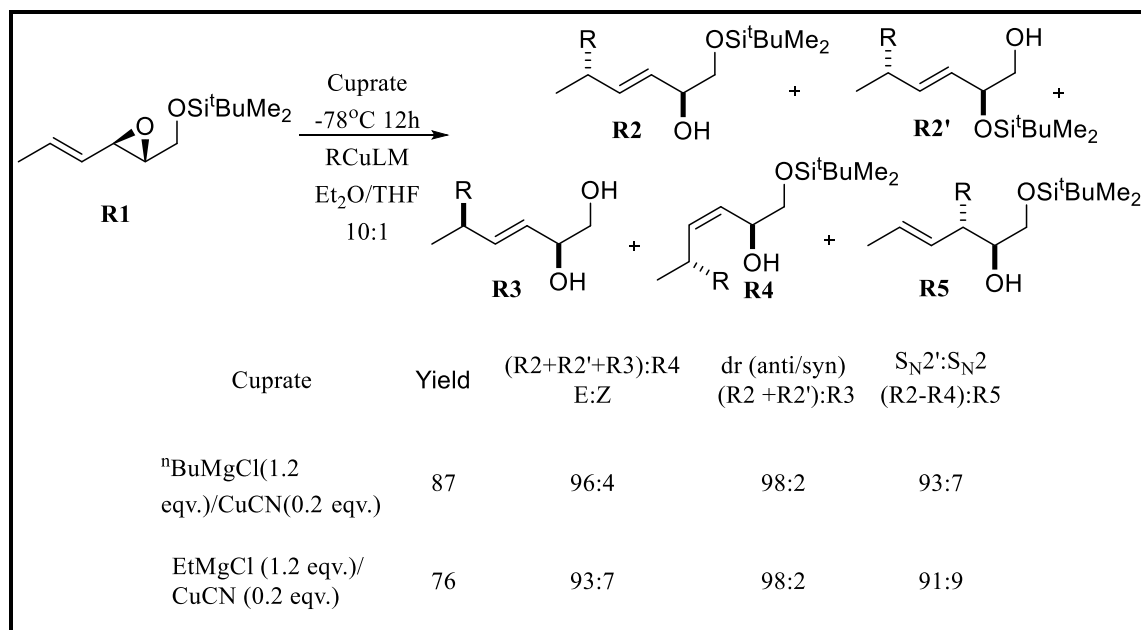


Figure 2.14. Copper-mediated reactions of trans-1-(tert-Butyldimethylsilyloxy)-2,3-epoxy-4-hexene (Source: Dieter, Huang, and Guo 2012)

In 2010, the iron-catalyzed regio selective reaction of γ,δ -epoxy- α,β -unsaturated esters with grignard reagents was published by Hata (Hata et al. 2010). Result of the reaction, only one regio isomer was occurred at good yield with alkyl grignard reagent and at moderate yield with phenyl grignard and methoxy substituted phenyl grignard reagent (Figure 2.15).

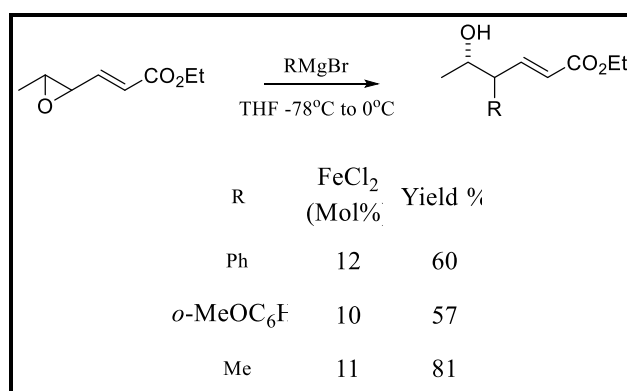


Figure 2.15 The iron-catalyzed reaction of vinyl oxirane (Source: Hata et al. 2010)

2.3. Metal-Catalyzed Reactions of Vinyl Oxiranes with Organoborons

The regio- and stereo-selective substitution reactions of vinyl oxiranes can be a good strategy for the synthesis of allyl alcohols.

In 2009, Tortosa reported that 1,4-diol compounds can be obtained via the copper-catalyzed successive borylation and oxidation reactions of vinyl oxiranes (Figure 2.16) (Tortosa 2011).

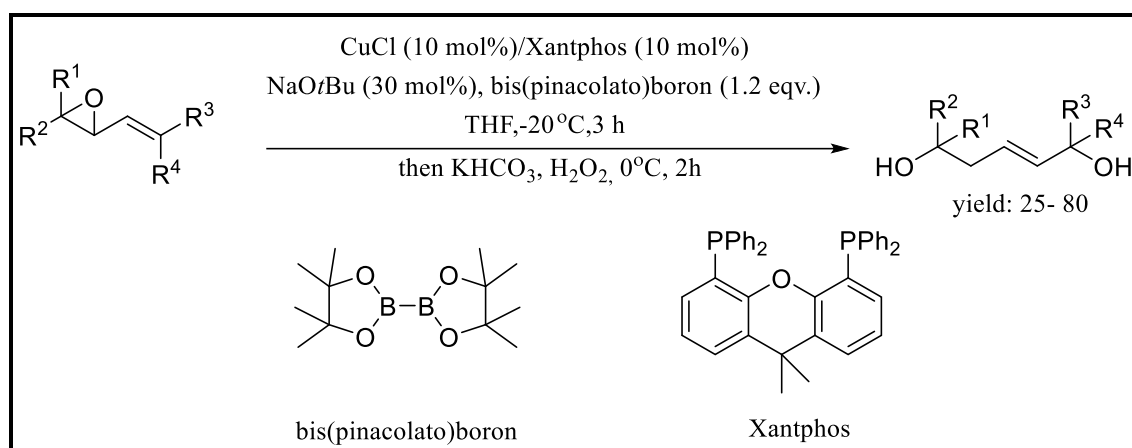


Figure 2.16. A Cu-catalyzed borylation and oxidation sequence leading to 1,4-diol compounds (Source: Tortosa 2011).

A Nickel-catalyzed borylative ring opening reaction of vinyl epoxides was published by Crotti *et al.* Accordingly, a regio-selective borylative ring opening takes place catalytically (Figure 2.17) and this follows by addition onto an aldehyde to yield a diol compound.

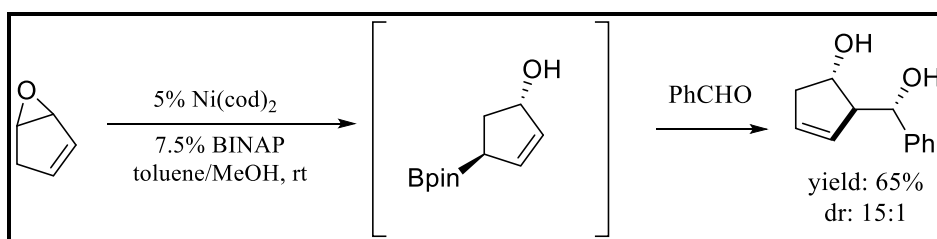


Figure 2.17. Nickel-catalyzed borylative ring opening reaction of vinyl epoxides (Source: Crotti *et al.* 2009)

The Pd-catalyzed reaction of vinyl oxiranes with alkenylborons is reported in 1982 by Miyaura. In their studies 3,4-epoxy-1-butane was the only vinyl epoxide

compound that was tested and no sufficient regio-selectivity could be provided with this substrate (Figure 2.18).

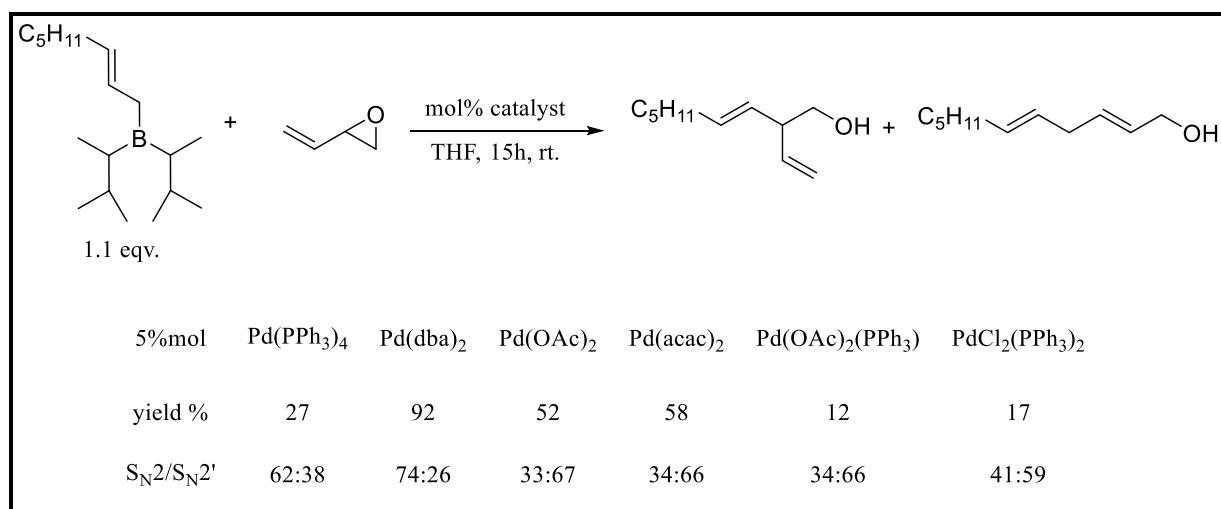


Figure 2.18. Pd-catalyzed reaction of (*E*)-hexenylbis(1,2-dimethylpropyl) borane with 3,4-epoxy-1-butene (Miyaura, Tanebe and Suginome 1982).

A Pd-pincer complex-catalyzed method on alkenylation and arylation of terminal vinyl epoxides with organoborons was developed by Kjellgren. The linear allyl alcohols were produced with high regio-selectivity (Figure 2.19). But, when this method was applied on a cyclic structure epoxide, the regio-selectivity was decreased to 1,3-/1,2-ratio of 2:1 (Kjellgren et al. 2005).

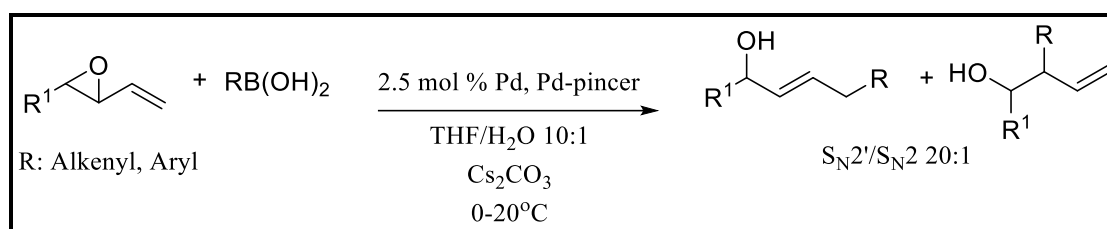


Figure 2.19. Pincer complex catalyzed cross-coupling reaction of vinyl epoxides with boronic acids (Source: Kjellgren et al. 2005).

CHAPTER 3

EXPERIMENTAL STUDY

3.4. General

Tetrahydrofuran (THF), diethyl ether (Et₂O) and 1,4 -dioxane solvents were distilled from benzophenone-ketyl under argon prior to use. Dichloromethane (DCM) and toluene were dried using molecular sieve 4A. Dimethylformamide (DMF) was purified by reflux over calcium hydride (5 w/v) under inert gas overnight and stored on molecular sieve 4A. Dimethoxyethane (DME) was dried by storing over the metallic sodium wire. Acetonitrile stirred with P₂O₅ (5% w/v) for 24h and then it was refluxed under inert gas overnight, then it was distilled.

The palladium complexes of Pd₂(dba)₃·CHCl₃ (Ukai, et al. 1974), Pd₂(dba)₃ (Zalesskiy and Ananikov 2012), and [PdCl(C₃H₅)₂] (Dent, Long and Wilkinson 1964) were synthesized in the laboratory.

3.5. Synthesis of Substrates

3.5.1. Synthesis of Vinyl Oxiranes

The syntheses of all compounds were performed under argon gas and the column chromatography purification of the synthesized compounds was applied on the silica gel 60 (35–70 μm).

Triethyl phosphonoacetate (4.8mL, 24 mmol) was added drop by drop to the solution of NaH (525 mg, 22 mmol) in 50 mL THF and the mixture was stirred for 1 hour at room temperature. After that, the reaction mixture was cooled to –78°C and *trans*-aldehyde (~20 mmol) was added slowly and stirred for 1 hour. The reaction was terminated by the addition of saturated NH₄Cl and extracted with Et₂O. The organic layer

was dried over MgSO_4 , filtered, and concentrated under reduced pressure to obtain **S2**. The residue was used in following step without any further purification.

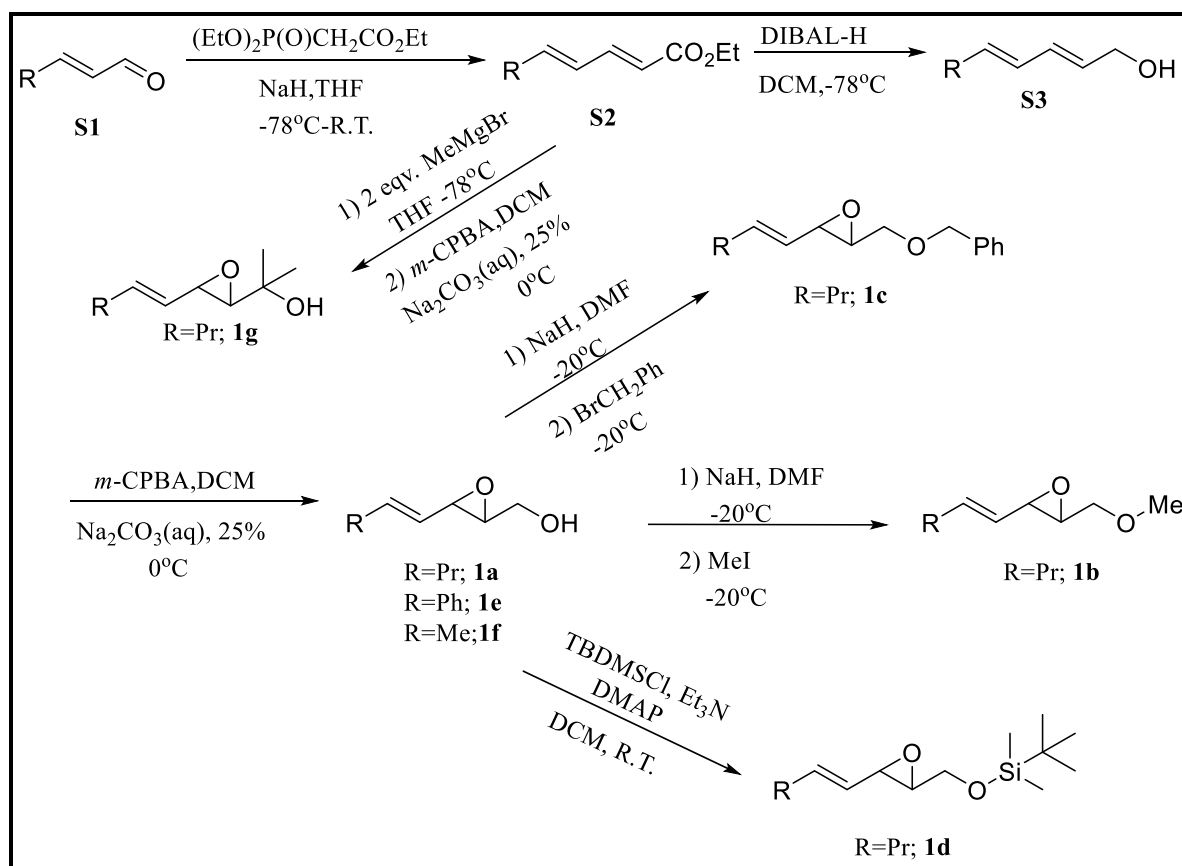


Figure 3.1. Synthesis of Alkenyl Oxiranes

At $-78\text{ }^\circ\text{C}$, the DIBAL-H ($\sim 30\text{ mmol}$, 1.0 M in DCM) solution was added dropwise to the solution of **S2** in DCM (80 mL). The reaction mixture was stirred for 3 hours and then the reaction was neutralized with 1 M HCl(aq) solution and extracted with DCM. The combined extracts were dried over MgSO_4 , filtered, and concentrated under reduced pressure. Purification by column chromatography on silica gel 60 (hexane/ ethyl acetate as the eluent) gave the diene compounds **S3** (yield: R: Pr, 1.64 g , 65% ; R, Ph, 1.92 g , 60% , R, Me, 1.08 g , 55%).

To a mixture of **S3** ($\sim 3\text{ mmol}$) in DCM (50 mL) and 16 mL solution of Na_2CO_3 (25%) was added 5.1 mmol (880 mg) of *m*-CPBA dropwise at $0\text{ }^\circ\text{C}$ under Argon. The mixture was controlled with TLC until the reactant was consumed completely. At the end of the process, the mixture was extracted with DCM, dried over anhydrous MgSO_4 , filtered, and concentrated under reduced pressure. The crude mixture was chromatographed on NEt_3 -treated silica gel column (hexane/ ethyl acetate as the eluent)

which afforded the vinyl oxiranes (yield: **1a**, 234,2 mg, 55%; **1e**, 306,2 mg, 58%, **1f**, 110 mg 30%).

1 mL of 2 M solution of vinyl epoxide alcohol (**1a** or **1b**) in DMF was added on the suspension of NaH (2.2 mmol) in DMF (1 mL) at -20 °C. The mixture was stirred until all NaH solids were dissolved before the addition of methyl iodide (2.4 mmol) or benzyl bromide (2.4 mmol). The mixture was stirred for 4 h at the same temperature, and then the reaction was terminated by the addition of MeOH (5 mL) and brine (5 mL) and extracted with DCM. The combined extracts were dried over MgSO₄, filtered, and concentrated under reduced pressure. The crude mixture was subjected to column chromatography over NEt₃-treated silica gel column (eluent: hexane-EtOAc) to produce compound **1b** or **1c** or **1f** (yield, **1b**: 278 mg, 90% ,**1e**: 200 mg, 96%, **1c**: 334 mg, 77%).

A mixture of **1a** (285 mg, 2 mmol), Et₃N (0.36 mL, 2.6 mmol), *t*-butyldimethylsilyl chloride (400 mg, 2.6 mmol), and 4-dimethylaminopyridine (DMAP) (30 mg, 0.24 mmol) in DCM (30 mL) was stirred at room temperature for 1 h and then, the mixture was extracted with water and the water phase washed with DCM. The organic solution was dried with MgSO₄, filtered, and evaporated. The residue was chromatographed over NEt₃-treated silica gel column to afford silylated vinyl oxirane compound **1d** (hexane-EtOAc as an eluent, yield, **1d**: 500 mg, 97%).

At -78°C, the 3.5 mL of 3 M MeMgBr in THF was added drop by drop to 5 mmol of **S2** solution in 15 mL THF. After the addition, the reaction mixture was heated slowly to rt and then stirred for 30 min. The saturated NH₄Cl(aq) solution (10 mL) was added in the mixture and then extracted with Et₂O. The organic layer was dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was used in following step without any further purification. To the mixture of dienol obtained (~5 mmol) in DCM (70 mL) and 25 mL solution of Na₂CO₃ (25%) was added 7 mmol (1.2 g) of *m*-CPBA dropwise at 0 °C under Argon. The mixture was controlled with TLC until the reactant was consumed completely. At the end of the process, the mixture was extracted with DCM, dried over anhydrous MgSO₄, filtered, and concentrated under reduced pressure. The crude mixture was chromatographed on NEt₃-treated silica gel column (hexane/ ethyl acetate as the eluent) which afforded the vinyl oxirane (yield: 1g, 485 mg ,57%) (Kus and Artok 2015).

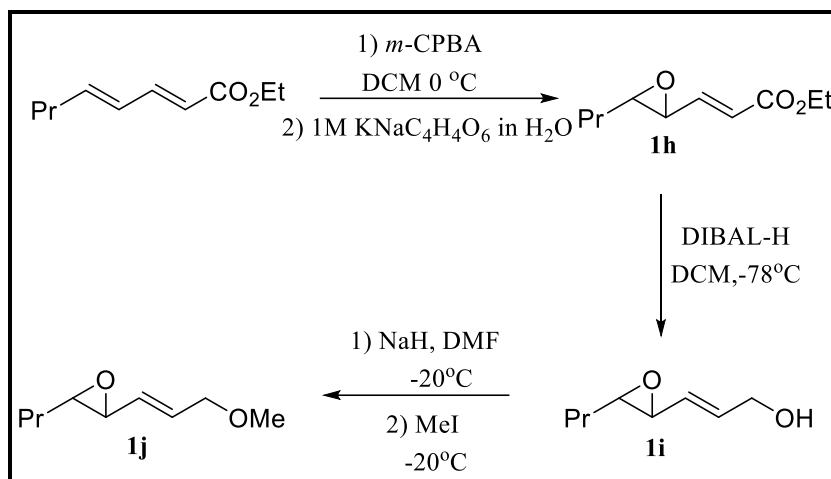


Figure 3.2 Epoxidation of ethyl (2*E*,4*E*)-octa-2,4-dienoate to synthesis **1h**

To the DCM solution (15 mL) of ethyl-(2*E*,4*E*)-octa-2,4-dienoate (840 mg, 5mmol) was added *m*-CPBA (1.7 g 6 mmol) in one portion at 0 °C and stirred for 2 h. The crude solution was extracted with a saturated solution of Na₂CO₃(aq) and DCM, successively, and dried with MgSO₄, filtered, and evaporated under reduced pressure. The substrate was purified using silica gel column (hexane-EtOAc as an eluent, yield, **1h**, 698 mg, 76%) (Dieter, Huang, and Guo 2012).

To a solution of **1j** (920 mg, 5 mmol) in DCM (9.0 mL), was added DIBAL-H (11.0 mL of a 1.0 M solution in DCM, 5.2 mmol) dropwise at -78 °C and stirred for 30 min. Then, MeOH (4.5 mL) was added and the mixture was warmed to ambient temperature. Then, an aqueous sodium potassium tartrate (1.0 M) solution (Rochelle's salt, 15 mL) was added and the mixture was stirred at rt for 1 h. The aqueous phase was extracted with EtOAc. The combined organic layers were dried over anhydrous Na₂SO₄, filtered, and concentrated to provide allylic alcohol **1i**. The substrate **1i** was purified using silica gel column (hexane-EtOAc as the eluent, yield, **1i**, 575 mg, 80%) (Tortosa 2011). The above-mentioned methylation process was applied for synthesis **1j** with the level of 90% yield.

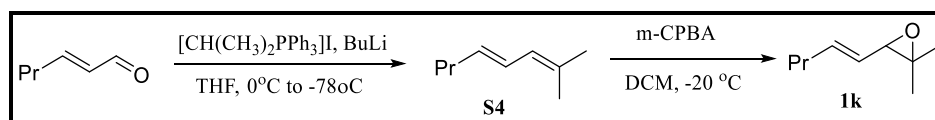


Figure 3.3 Synthesis of (*E*)-2,2-dimethyl-3-(pent-1-en-1-yl) oxirane **1k**

A hexane solution of BuLi (4.8 mL, 12 mmol, 2.5 M) was added dropwise to a solution of isopropyl(triphenyl)-phosphonium iodide (4.32 g, 10 mmol) in THF (30 mL) at 0 °C and stirred for further 1 h. The *trans*-2-hexenal (1.1 mL, 10 mmol) was added slowly in the mixture at -78 °C. The reaction mixture was stirred until α,β unsaturated aldehyde was consumed as judged by TLC. After that, the reaction was neutralized and extracted with saturated NH₄Cl solution and Et₂O, successively. The combined extracts were dried over MgSO₄, filtered, and separated from the solvent by reduced pressure at 0 °C. After purification over the silica gel column the pure dialkyl substituted diene compound **S4** was obtained (hexane-EtOAc as the eluent, yield, 920 mg, 74%).

To the suspension of *m*-CPBA (7.5 mmol) in 30 mL DCM was added drop by drop in to 0.1 M the diene compound (**S4**) solution in 50 mL DCM at -20 °C. After the solution was stirred for 2 h, the extraction process was applied with water and DCM. After that the organic layer was dried with MgSO₄, and concentrated under reduced pressure at 25 °C. Using silica gel column (hexane-EtOAc as the eluent), the pure vinyl oxirane **1k** was obtained. The regio-selectivity of the oxirane ((*E*)-2,2-dimethyl-3-(pent-1-en-1-yl)oxirane/2-(2-methylprop-1-en-1-yl)-3-propyloxirane) was determined to be 9.6:1 (yield, **1k**: 365 mg 52%) (Kuş et al., 2015).

1,3-Cyclohexadiene (1mL, 10 mmol), 50mL DCM, and 16 mL 25% Na₂CO₃(aq) was added into a flask in sequence. After this mixture was cooled to 0 °C, 2 g *m*-CPBA was added and the reaction was controlled with help of TLC. After reactant was consumed, the solution was extracted with DCM, dried with MgSO₄, and filtered over a short silica gel column. The solvent was removed under reduced pressure at 0 °C and the cyclic epoxide **11** was recovered in pure form (yield, 430 mg, 45%). (Ramesh, et al. 1992).

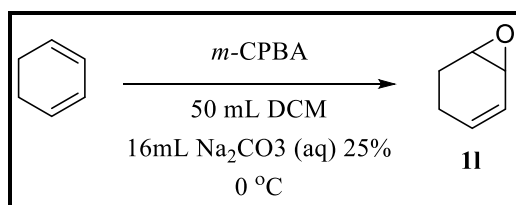


Figure 3.4 The synthesis of 3,4-epoxycyclohexene **11**.

To a solution of hex-1-yne (2.05 g, 25.0 mmol) in hexanes (15 mL) at 0 °C was added diisobutylaluminum hydride (1 M in DCM, 25 mL, 25.0 mmol). The solution was warmed to 40 °C and stirred for 16 h. Volatile components were then evaporated, and the

residue was dissolved in THF (50 mL) and cooled to 0 °C. CuCl (2.25 g, 27.5 mmol) was then added in portions over 10 min, and the suspension allowed to warm to room temperature and stir for 4 h. The black suspension was then poured onto a stirred mixture of H₂SO₄ (5%, 100 mL) and Et₂O (100 mL), the phases were separated, and the aqueous layer further extracted with Et₂O. The substrate (5E,7E)-dodeca-5,7-diene was purified using silica gel column (hexane as the eluent, yield 1,6 g, 79%)(Farthing and Koc 1998). After that, the epoxidation process was applied and the substrate **1m** was obtained.

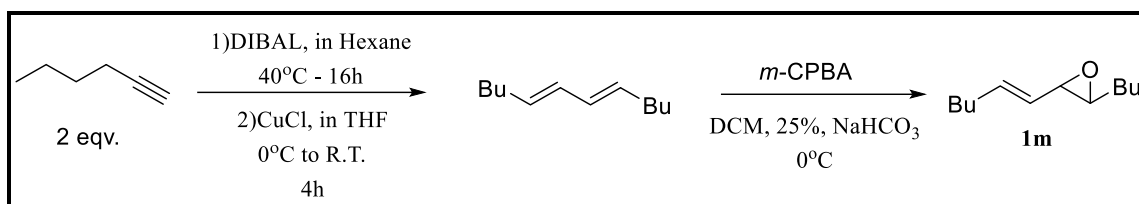


Figure 3.5 The synthesis of (E)-2-butyl-3-(hex-1-en-1-yl)oxirane

3.5.2. Characterization of Alkenyl Oxiranes

The synthesized reactants were analyzed by GC-MS and the NMR spectra were recorded on a 400 MHz spectrometer.

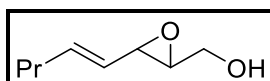


Figure 3.6. (E)-(3-(pent-1-en-1-yl) oxiran-2-yl) methanol

1a: ¹H NMR (400 MHz, CDCl₃) δ: 5,96 (td, *J* = 6,7, 15,5 Hz, 1H), 5,20 (tdd, *J* = 1,4, 8,3, 15,5 Hz, 1H), 3,94 (d, *J* = 12,5 Hz, 1H), 3,73–3,63 (m, 1H), 3,38 (dd, *J* = 2,3, 8,2 Hz, 1H), 3,08 (td, *J* = 2,3, 3,9 Hz, 1H), 2,06 (q, *J* = 6,7 Hz, 2H), 1,42 (sex, *J* = 7,4 Hz, 2H), 0,91 (t, *J* = 7,4 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ: 137,7, 126,4, 61,3, 59,9, 55,9, 34,4, 22,0, 13,6. MS (EI, *m/z*): 111(22), 99(24), 83(78), 69(98), 55(100).

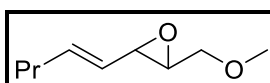


Figure 3.7. (E)-2-(methoxymethyl)-3-(pent-1-en-1-yl) oxirane

1b: ^1H NMR (400 MHz, CDCl_3) δ : 5,94 (td, $J = 6,7, 15,5$ Hz, 1H), 5,18 (dd, $J = 8,2, 15,7$ Hz, 1H), 3,66 (dd, $J = 3,1, 11,3$ Hz, 1H), 3,44–3,41 (m, 1H), 3,40 (s, 3H), 3,24 (dd, $J = 2,0, 8,2$ Hz, 1H), 3,05 (td, $J = 2,7, 5,5$ Hz, 1H), 2,05 (q, $J = 7,0$ Hz, 1H), 1,41 (sxt, $J = 7,4$ Hz, 2H), 0,91 (t, $J = 7,4$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ : 137,5, 126,7, 72,4, 59,2, 58,4, 56,1, 34,4, 22,0, 13,6. MS (EI, m/z): 123(4), 113(34), 83(24), 81(38), 71(40), 58(80), 45(100).

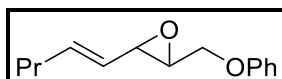


Figure 3.8. (*E*)-2-(pent-1-en-1-yl)-3-(phenoxy)methyl oxirane

1c: ^1H NMR (400 MHz, CDCl_3) $\delta = 7,40 - 7,26$ (m, 5H), 5,93 (td, $J = 6,9, 15,5$ Hz, 1H), 5,17 (dd, $J = 8,4, 15,5$ Hz, 1H), 4,60 (d, $J_{AB} = 11,7$, Hz 1H), 4,56 (dd, $J_{AB} = 11,7$ Hz, 1H), 3,75 (dd, $J_{AB} = 11,3$ Hz, $J_{AX} = 5,5$ Hz, 1H), 3,51 (dd, $J_{AB} = 11,3$ Hz, $J_{BX} = 3,1$ Hz, 1H), 3,25 (dd, $J = 2,3, 8,2$ Hz, 1H), 3,10 (td, $J = 2,6, 5,4$ Hz, 1H), 2,04 (q, $J = 6,7$ Hz, 2H), 1,41 (sxt, $J = 7,4$ Hz, 2H), 0,90 (t, $J = 7,2$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 137,8, 137,6, 128,4, 127,8, 127,8, 126,7, 73,3, 70,0, 58,7, 56,2, 34,4, 22,0, 13,7$. MS (EI, m/z) 232 (1^+ , M^+), 126 (11), 111(2), 107 (8), 91 (100), 83 (41), 68 (5), 65 (12), 55 (25), 41 (15).

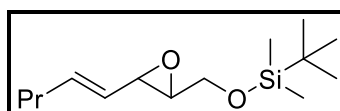


Figure 3.9. (*E*)-*tert*-butyldimethyl((3-(pent-1-en-1-yl) oxiran-2-yl) methoxy) silane

1d: ^1H NMR (400 MHz, CDCl_3) $\delta = 5,92$ (td, $J = 6,7, 15,5$ Hz, 1H), 5,19 (tdd, $J = 1,6, 8,3, 15,6$ Hz, 1H), 3,85 (dd, $J_{AB} = 11,9$ Hz, $J_{AX} = 4,5$ Hz, 1H), 3,70 (dd, $J_{AB} = 11,9$ Hz, $J_{BX} = 3,3$ Hz, 1H), 2,99 (ddd, $J = 1,6, 2,6, 3,6$ Hz, 1H), 2,08 – 2,00 (m, $J = 1,4, 8,0$ Hz, 2H), 1,41 (sxt, $J = 7,4$ Hz, 2H), 0,89 (s, 9H), 0,90 (t, $J = 7,4$ Hz, 3H), 0,07 (d, $J = 3,1$ Hz, 6H). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 137,1, 126,9, 63,2, 60,3, 56,3, 34,4, 25,9, 22,1, 13,6, -5,3$. MS (EI, m/z) 256 (1^+ , M^+), 199 (24), 185 (2), 169 (8), 157 (9), 143 (23), 129 (13), 117 (14), 113 (22), 107 (16), 101 (33), 89 (18), 83 (22), 79 (15), 75 (99), 59 (38), 55 (100), 41 (33).

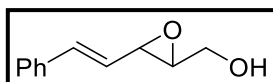


Figure 3.10. (*E*)-3-(3-styryloxiran-2-yl) methanol

1e: ^1H NMR (400 MHz, CDCl_3) δ = 7,42 – 7,36 (m, 2H), 7,35 – 7,30 (m, 2H), 7,29 – 7,26 (m, 1H), 6,81 (d, J = 16,0 Hz, 1H), 5,94 (dd, J = 8,0, 15,8 Hz, 1H), 4,06 – 3,97 (m, 1H), 3,81 – 3,70 (m, 1H), 3,60 (dd, J = 2,2, 8,0 Hz, 1H), 3,21 (td, J = 2,3, 3,9 Hz, 1H). ^{13}C NMR (101 MHz, CDCl_3) δ = 135,9, 135,0, 128,7, 128,2, 126,5, 125,7, 61,1, 60,4, 55,9. MS (EI, m/z) 176 (12, M^+), 145 (22), 127 (16), 117 (100), 115 (60), 91 (31), 77 (10), 65 (12), 51 (12).

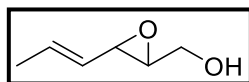


Figure 3.11. (*E*)-3-(prop-1-en-1-yl) oxiran-2-yl methanol

1f: ^1H NMR (400 MHz, CDCl_3) δ = 5.96 (qd, J = 7.0, 14.9 Hz, 1 H), 5.22 (qdd, J = 1.6, 8.2, 15.3 Hz, 1 H), 3.93 (dd, J = 2.2, 12.7 Hz, 1 H), 3.66 (dd, J = 3.7, 12.7 Hz, 1 H), 3.37 (dd, J = 2.2, 8.4 Hz, 1 H), 3.09 - 3.04 (m, 1 H), 1.74 (td, J = 1.6, 6.7 Hz, 3 H). ^{13}C NMR (101 MHz, CDCl_3) δ = 132.5, 127.6, 61.3, 59.9, 55.8, 17.9.

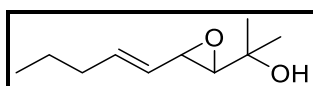


Figure 3.12. (*E*)-2-(3-(pent-1-en-1-yl) oxiran-2-yl) propan-2-ol

1g: ^1H NMR (400 MHz, CDCl_3) δ = 5.90 (td, J = 7.1, 15.2 Hz, 1 H), 5.33 (ddd, J = 1.6, 7.8, 15.3 Hz, 1 H), 3.17 (d, J = 8.2 Hz, 1 H), 2.06 (q, J = 7.0 Hz, 2 H), 1.77 (d, J = 19.6 Hz, 1 H), 1.42 (qd, J = 7.4, 14.7 Hz, 2 H), 1.34 (s, 3 H), 1.28 (s, 3 H), 0.91 (t, J = 7.4 Hz, 3 H). ^{13}C NMR (101 MHz, CDCl_3) δ = 137.5, 125.2, 110.0, 64.4, 60.0, 34.6, 24.6, 22.2, 18.9, 13.6.

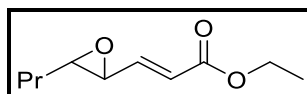


Figure 3.13. Ethyl (*E*)-3-(3-propyloxiran-2-yl) acrylate

1h: ^1H NMR (400 MHz, CDCl_3) δ = 6.68 (dd, J = 7.0, 15.7 Hz, 1 H), 6.12 (d, J = 15.7 Hz, 1 H), 4.20 (q, J = 7.0 Hz, 2 H), 3.20 (dd, J = 1.6, 7.0 Hz, 1 H), 2.89 (dt, J = 2.2, 5.6 Hz, 1 H), 1.68 - 1.38 (m, 4 H), 1.29 (t, J = 7.2 Hz, 3 H), 1.00 - 0.94 (m, 3 H). ^{13}C NMR (101 MHz, CDCl_3) δ = 165.7, 144.8, 123.5, 61.3, 60.5, 56.3, 33.9, 19.1, 14.2, 13.8.

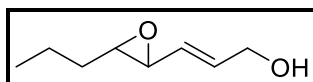


Figure 3.14. (*E*)-3-(3-propyloxiran-2-yl) prop-2-en-1-ol

1i: ^1H NMR (400 MHz, CDCl_3) δ = 6.06 (td, J = 5.5, 15.7 Hz, 1 H), 5.48 (tdd, J = 1.9, 7.8, 15.5 Hz, 1 H), 4.17 (dd, J = 1.4, 5.3 Hz, 2 H), 3.12 (dd, J = 2.3, 7.8 Hz, 1 H), 2.84 (dt, J = 2.0, 5.5 Hz, 1 H), 1.63 - 1.41 (m, 4 H), 0.99-0.92 (m, 3 H). ^{13}C NMR (101 MHz, CDCl_3) δ = 133.9, 129.0, 62.7, 60.4, 57.9, 34.0, 19.2, 13.9.

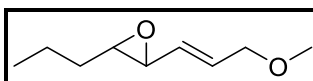


Figure 3.15. (*E*)-2-(3-methoxyprop-1-en-1-yl)-3-propyloxirane

1j: ^1H NMR (400 MHz, CDCl_3) δ = 5.96 (td, J = 5.6, 15.8 Hz, 1 H), 5.47 (tdd, J = 1.4, 7.8, 15.6 Hz, 1 H), 3.93 (dd, J = 1.6, 5.9 Hz, 2 H), 3.34 (s, 3 H), 3.11 (dd, J = 2.3, 7.8 Hz, 1 H), 2.83 (dt, J = 2.3, 5.5 Hz, 1 H), 1.60 - 1.41 (m, 4 H), 0.95 (t, J = 7.4 Hz, 3 H). ^{13}C NMR (101 MHz, CDCl_3) δ = 131.2, 130.5, 72.1, 60.3, 58.1, 57.9, 34.0, 19.2, 13.9.

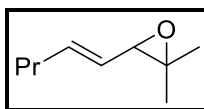


Figure 3.16. (*E*)-2,2-dimethyl-3-(pent-1-en-1-yl) oxirane

1k: ^1H NMR (400 MHz, CDCl_3) δ = 5.90 (td, J = 7.0, 15.7 Hz, 1 H), 5.31 (dd, J = 7.8, 15.7 Hz, 1 H), 3.17 (d, J = 7.8 Hz, 1 H), 2.07 (q, J = 7.0 Hz, 2 H), 1.42 (sxt, J = 7.4 Hz, 2 H), 1.34 (s, 3 H), 1.28 (s, 3 H), 0.91 (t, J = 7.0 Hz, 2 H). ^{13}C NMR (101 MHz, CDCl_3) δ = 137.5, 125.2, 64.4, 60.0, 34.6, 24.6, 22.2, 18.9, 13.6.

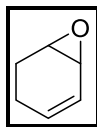


Figure 3.17. 7-Oxabicyclo [4.1.0] hept-2-ene

1l: ^1H NMR (400 MHz, CDCl_3) δ = 6.07 - 5.81 (m, 2 H), 3.51 (td, J = 1.4, 2.7 Hz, 1 H), 3.24 (dt, J = 1.8, 4.0 Hz, 1 H), 2.31 - 2.17 (m, 1 H), 2.13 - 1.99 (m, 2 H), 1.68 - 1.57 (m, 1 H). ^{13}C NMR (101 MHz, CDCl_3) δ = 133.1, 123.1, 55.2, 47.1, 20.8, 20.6.

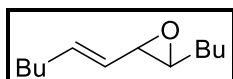
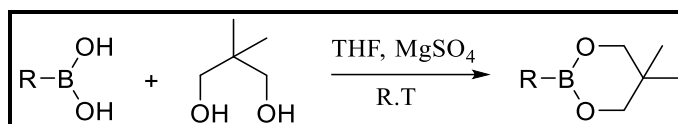


Figure 3.18(E)-2-butyl-3-(hex-1-en-1-yl)oxirane

1m: ^1H NMR (400 MHz, CDCl_3) δ = 5.89 (td, J = 7.0, 14.5 Hz, 1 H), 5.16 (ddd, J = 1.2, 8.0, 15.5 Hz, 1 H), 3.05 (d, J = 8.2 Hz, 1 H), 2.80 (t, J = 5.5 Hz, 1 H), 2.05 (q, J = 6.9 Hz, 2 H), 1.59 - 1.23 (m, 10 H), 0.89 (q, J = 7.4 Hz, 6 H). ^{13}C NMR (101 MHz, CDCl_3) δ = 136.6, 127.5, 60.4, 58.8, 32.0, 31.7, 31.0, 28.0, 22.5, 22.2, 14.0, 13.9.

3.5.3.Synthesis of Organoborons

Table 3.1.Synthesis of neopentyl glycol esters



R (yield%)				
2a (91%)	2b (89%)	2c (85%)	2d (87%)	2e (71%)
2f (90%)	2g (82%)	2h (90%)	2i (92%)	2j (92%)
2k (87%)	2l (76%)	2m (75%)	2n (97%)	2o (90%)
		2p (80%)		

Under Ar gas and at rt, to the dry THF (15 mL) solution of organoboronic acid (10 mmol) was added MgSO₄ (14 mmol, 1.7 g). Then, 2,2 dimethyl propan-1,3-diol (11 mmol, 1.2 g) was added to the mixture and stirred overnight. The crude mixture was concentrated under reduced pressure and boronic acid neopentyl glycol ester derivative was purified on silica gel column (hexane-EtOAc as an eluent) (Matthew et al. 2014). The yields are given in Table 3.1.

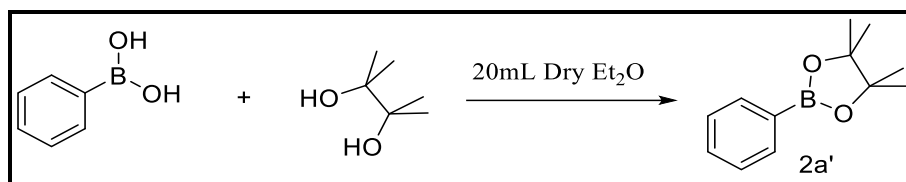


Figure 3.19. Synthesis of phenyl boronic acid pinacol ester

Phenylboronic acid (610 mg, 5 mmol) and 2,3-dimethyl-2,3-butanediol (590 mg, 5 mmol) in 20 mL of diethyl ether were stirred overnight at rt. The crude mixture was concentrated under reduced pressure and phenylboronic acid pinacol ester (2a') was obtained after chromatographed on silica gel column (hexane-EtOAc as an eluent). (Morandi, et al. 2005).

3.5.4.Characterization of Organoborons

The synthesized boronic acid ester derivatives were analyzed by NMR spectra recorded on a 400 MHz spectrometer.

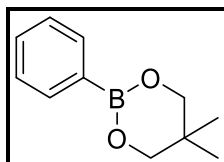


Figure 3.20. 5,5-Dimethyl-2-phenyl-1,3,2-dioxaborinane

2a: $^1\text{H NMR}$ (400MHz, CDCl_3) δ : 7.80 (d, $J = 7.0$ Hz, 2H), 7.46-7.39 (m, 1H), 7.38-7.32 (m, 2H), 3.77 (d, $J = 0.8$ Hz, 4H), 1.02 (s, 6H).

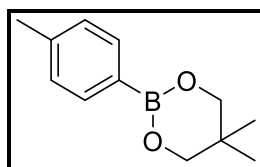


Figure 3.21. 5,5-Dimethyl-2-(p-tolyl)-1,3,2-dioxaborinane

2b: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ : 7.71 (d, $J = 7.7$ Hz, 2H), 7.19 (d, $J = 7.6$ Hz, 2H), 3.77 (s, 4H), 2.37 (s, 3H), 1.03 (s, 6H).

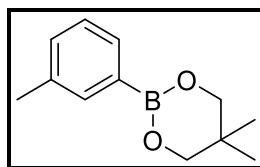


Figure 3.22. 5,5-Dimethyl-2-(o-tolyl)-1,3,2-dioxaborinane

2c: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ : 7.24-7.62 (4H, m), 3.77 (4H, s), 2.35 (3H, s), 1.02 (6H, s),

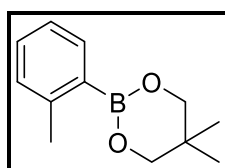


Figure 3.23. 5,5-Dimethyl-2-(m-tolyl)-1,3,2-dioxaborinane

2d: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ = 7.77 - 7.69 (m, 1 H), 7.31 - 7.25 (m, 1 H), 7.19 - 7.11 (m, 2 H), 3.78 (s, 4 H), 2.52 (s, 3 H), 1.04 (s, 6 H).

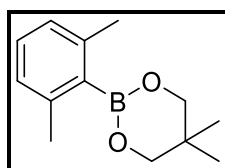


Figure 3.24. 2-(2,6-Dimethylphenyl)-5,5-dimethyl-1,3,2-dioxaborinane

2e: $^1\text{H NMR}$ (400MHz, CDCl_3) δ = 7.10 (t, J = 7.0 Hz, 1 H), 6.94 (d, J = 7.4 Hz, 2 H), 3.89 - 3.70 (m, 4 H), 2.39 (s, 6 H), 1.22 - 1.00 (m, 6 H).

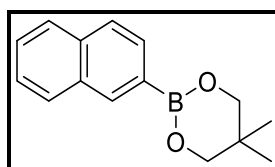


Figure 3.25. 5,5-Dimethyl-2-(naphthalen-2-yl)-1,3,2-dioxaborinane

2f: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ : 8.35 (s, 1 H), 7.80-7.89 (m, 4 H), 7.44-7.51 (m, 2 H), 3.84 (s, 4H), 1.06 (s, 6H).

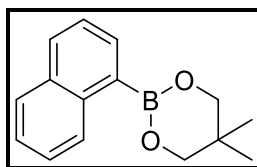


Figure 3.26. 5,5-Dimethyl-2-(naphthalen-1-yl)-1,3,2-dioxaborinane

2g: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ : 8.0 (dd, $J = 8.4, 1.2$ Hz, 1H), 8.08 (dd, $J = 6.9, 1.4$ Hz, 1H), 7.93 (d, $J = 8.2$ Hz, 1H), 7.86 (dd, $J = 8.1, 1.5$ Hz, 1H), 7.59-7.42 (m, 3H), 3.91 (s, 4H), 1.11 (s, 6H).

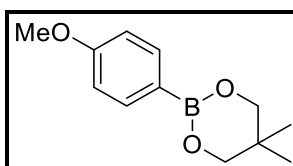


Figure 3.27. 2-(4-methoxyphenyl)-5,5-dimethyl-1,3,2-dioxaborinane

2h: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ : 7.76 (d, $J = 8.5$ Hz, 2H), 6.90 (d, $J = 8.5$ Hz, 2H), 3.83 (s, 3H), 3.76 (s, 4H), 1.02 (s, 6H).

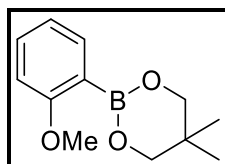


Figure 3.28. 2-(2-Methoxyphenyl)-5,5-dimethyl-1,3,2-dioxaborinane

2i: $^1\text{H NMR}$ (400 MHz, CDCl_3) $\delta = 7.65$ (d, $J = 7.4$ Hz, 1 H), 7.36 (t, $J = 7.8$ Hz, 1 H), 6.94 (t, $J = 7.4$ Hz, 1 H), 6.86 (d, $J = 8.2$ Hz, 1 H), 3.84 (s, 3 H), 3.79 (s, 4 H), 1.04 (s, 6 H).

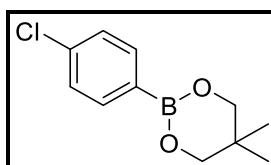


Figure 3.29. 2-(4-Chlorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane

2j: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ : 7.75 – 7.67 (m, 2 H), 7.35 – 7.29 (m, 2 H), 3.76 (s, 4H), 1.02 (s, 6H).

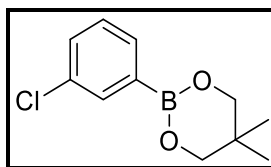


Figure 3.30. 2-(3-Chlorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane

2k: $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.77 (s, 1H), 7.66 (d, 1H), 7.39 (m, 1H), 7.27 (t, 1H), 3.78 (s, 4H), 1.03 (s, 6H).

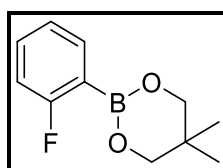


Figure 3.31. 2-(2-Fluorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane

2l: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ = 7.75 - 7.69 (m, 1 H), 7.43 - 7.35 (m, 1 H), 7.12 (t, J = 7.4 Hz, 1 H), 7.00 (t, J = 8.6 Hz, 1 H), 3.80 (s, 4 H), 1.12 - 0.99 (m, 6 H).

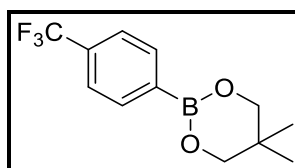


Figure 3.32. 5,5-Dimethyl-2-(4-(trifluoromethyl) phenyl)-1,3,2-dioxaborinane

2m: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ : 7.91 (d, J = 7.7 Hz, 2H), 7.60 (d, J = 7.8 Hz, 2H), 3.79 (s, 4H), 1.03 (s, 6H).

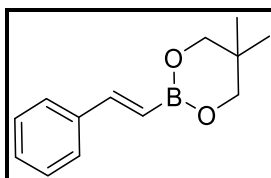


Figure 3.33. (*E*)-5,5-Dimethyl-2-styryl-1,3,2-dioxaborinane

2n: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ : 7.48 (d, J = 7.2 Hz, 2H), 7.40-7.16 (m, 4H), 6.11 (d, J = 18.3 Hz, 1H), 3.69 (s, 4H), 1.00 (s, 6H).

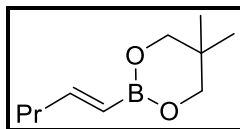


Figure 3.34. (*E*)-5,5-Dimethyl-2-(pent-1-en-1-yl)-1,3,2-dioxaborinane

2o: ^1H NMR (400MHz, CDCl_3) δ : 6.60 - 6.46 (m, 1 H), 5.34 (d, $J = 17.6$ Hz, 1 H), 3.63 (s, 4 H), 2.11 (dq, $J = 1.6, 6.7$ Hz, 2 H), 1.43 (sxt, $J = 7.4$ Hz, 2 H), 0.97 (s, 7 H), 0.90 (t, $J = 7.2$ Hz, 3 H).

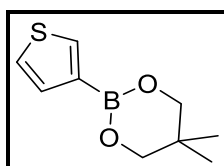


Figure 3.35. 5,5-Dimethyl-2-(thiophen-3-yl)-1,3,2-dioxaborinane

2p: ^1H NMR (400 MHz, CDCl_3) δ : 7.84 (dd, $J = 2.6, 0.8$ Hz, 1H), 7.38 (dd, $J = 4.8, 0.9$ Hz, 1H), 7.31 (dd, $J = 4.8, 2.7$ Hz, 1H), 3.75 (s, 4H), 1.02 (s, 6H).

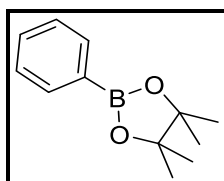


Figure 3.36. 4,4,5,5-Tetramethyl-2-phenyl-1,3,2-dioxaborolane

2a': ^1H NMR (400 MHz, CDCl_3) δ : 7.77 – 7.86 (m, 2 H), 7.46 (tt, $J = 7.40, 1.60$ Hz, 1H) 7.37 (t, $J = 7.80$ Hz, 2H), 1.35 (s, 12H).

3.6. General Method for Palladium-Catalyzed Reactions of Vinyl Oxiranes

Successively, catalyst, ligand, and dry solvent (half of the volume necessary for the reaction) were added in to the schlenk flask which was dried in oven and cooled under Ar gas. The mixture was stirred for 30 min. at 25 °C. After that successively, organoboron, base, and the solution of epoxide compound in dry solvent (other half volume) and

prescribed amount of degassed water was added to the schlenk flask and the mixture was stirred in water or oil bath. The reaction was controlled with help of TLC. When the reaction process was judged to be complete, the mixture was concentrated under reduced pressure. It must be noted based on our experience that the color of the reaction solution usually turned from yellow to dark brown when the reaction cycle is halted. The residue was purified using silica gel on column chromatography and the alcohol product was obtained. In part of the optimization studies the crude was filtered through a short silica gel column (height 10 cm and width 2 cm), washed with Et₂O and evaporated under reduced atmosphere. The residue was analyzed by ¹H NMR using *p*-anisaldehyde as the internal standard.

The synthesized products were analyzed by GC and GC-MS. NMR spectra in CDCl₃ or C₆D₆ were recorded on a 400 MHz spectrometer.

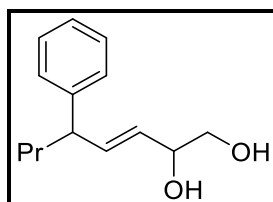


Figure 3.37. (*E*)-5-Phenyloct-3-ene-1,2-diol

3aa: ¹H NMR (400 MHz, CDCl₃) δ: 7.31–7.26 (m, 2H), 7.22–7.13 (m, 3H), 5.91 (dd, *J* = 7.8, 15.7 Hz, 1H), 5.46 (dd, *J* = 6.5, 15.7 Hz, 1H), 4.20 (dt, *J* = 3.5, 6.5 Hz, 1H), 3.61 (dd, *J* = 3.5, 11.3 Hz, 1H), 3.49–3.42 (m, 1H), 3.26 (q, *J* = 7.4 Hz, 1H), 2.16 (br, s, 2H), 1.68 (q, *J* = 7.4 Hz, 2H), 1.31–1.20 (m, 2H), 0.88 (t, *J* = 7.4 Hz, 3H), ¹³C NMR (101 MHz, CDCl₃) δ: 144.3, 137.6, 128.5, 127.8, 127.5, 126.2, 73.0, 66.5, 48.3, 37.9, 20.6, 14.0, ¹H NMR (400 MHz, C₆D₆) δ: 7.20–7.16 (m, 2 H), 7.12–7.02 (m, 3 H), 5.81 (ddd, *J* = 1.4, 7.8, 15.5 Hz, 1H) (major), 5.78 (ddd, *J* = 1.4, 7.8, 15.5 Hz, 1H) (minor), 5.34 (ddd, *J* = 15.5, 6.6, 1.2 Hz, 1H) (major), 5.31 (ddd, *J* = 15.5, 6.6, 1.2 Hz, 1H) (minor), 3.99–3.93 (m, 1H), 3.36 (dd, *J*_{AB} = 11.2 Hz, *J*_{AX} = 7.7 Hz, 1H), 3.35 (dd, *J*_{AB} = 11.2 Hz, *J*_{BX} = 3.6 Hz, 1H), 3.11 (q, *J* = 8.6 Hz, 1H), 2.27–1.92 (m, 2H), 1.57 (q, *J* = 7.7 Hz, 2H), 1.25–1.14 (m, 2H), 0.83 (t, *J* = 7.2 Hz, 3H), ¹³C NMR (101 MHz, C₆D₆) δ: 144.5, 136.3, 128.4, 128.4, 127.5, 126.1, 72.7, 66.5, 48.4, 37.9, 20.6, 13.8, MS (EI, *m/z*): 220 (M⁺, 1), 202 (2), 189 (56), 171 (23), 134 (12), 129 (57), 115 (33), 91 (100), 77 (8). FTIR (ν_{max}/cm⁻¹): 3398, 2925, 1724, 1601, 1453, 1343, 1030, 879, 700.

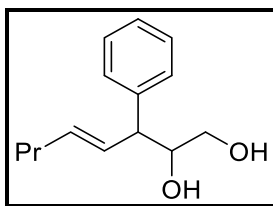


Figure 3.38. (*E*)-3-Phenyloct-4-ene-1,2-diol

4aa: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ : 7.35–7.28 (m, 2 H), 7.25–7.18 (m, 3 H), 5.72 (dd, $J_{AB} = 15.3$ Hz, $J_{AX} = 8.0$ Hz, 1H), 5.66 (dd, $J_{AB} = 15.3$ Hz, $J_{BX} = 5.9$ Hz, 1H), 3.84 (ddd, $J = 2.9, 5.9, 8.6$ Hz, 1H), 3.52 (dd, $J = 2.9, 11.5$ Hz, 1H), 3.39–3.31 (m, 2H), 2.05 (q, $J = 7.2$ Hz, 2H), 1.40 (sex, $J = 7.2$ Hz, 2H), 0.89 (t, $J = 7.2$ Hz, 3H), $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ : 134.8, 129.5, 128.8, 128.6, 127.8, 126.8, 74.3, 64.2, 52.7, 34.7, 22.4, 13.7, $^1\text{H NMR}$ (400 MHz, C_6D_6) δ : 7.15–6.94 (m, 5H), 5.68 (dd, $J = 8.6, 15.5$ Hz, 1H), 5.42 (td, $J = 6.7, 15.5$ Hz, 1H), 3.68 (ddd, $J = 3.1, 6.7, 7.9$ Hz, 1H), 3.40 (dd, $J = 3.1, 11.3$ Hz, 1H), 3.28 (dd, $J = 6.5, 11.3$ Hz, 2H), 1.85 (q, $J = 7.2$ Hz, 2H), 1.22 (sex, $J = 7.2$ Hz, 2H), 0.79 (t, $J = 7.2$ Hz, 3H), $^{13}\text{C NMR}$ (101 MHz, C_6D_6) δ : 141.8, 133.4, 130.1, 128.5, 128.0, 126.4, 74.4, 64.3, 52.4, 34.6, 22.4, 13.4, MS (EI, m/z): 220 (M^+ , 29), 205 (100), 145 (11), 105 (9), 91 (5). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3393, 2925, 1724, 1601, 1453, 1343, 1030, 879, 700.

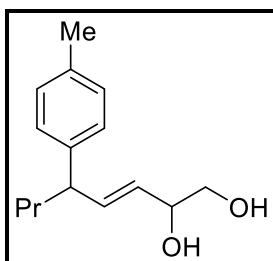


Figure 3.39. (*E*)-5-(*p*-Tolyl) oct-3-ene-1,2-diol

3ab: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ = 7.15 – 7.08 (m, 2H), 7.08 – 7.02 (m, 2H), 5.89 (ddd, $J = 1.2, 7.8, 15.7$ Hz, 1H), 5.43 (ddd, $J = 1.0, 6.4, 15.4$ Hz, 1H), 4.22 – 4.13 (m, 1H), 3.58 (dd, $J_{AB} = 11.3$ Hz, $J_{AX} = 7.5$ Hz, 1H), 3.43 (dd, $J_{AB} = 11.3$ Hz, $J_{BX} = 3.5$ Hz, 1H), 3.22 (q, $J = 7.4$ Hz, 1H), 2.60 (br, s., 2H), 2.32 (s, 3H), 1.66 (q, $J = 7.7$ Hz, 2H), 1.35 – 1.18 (m, 2H), 0.89 (t, $J = 7.4$ Hz, 3H), $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ = 141.3, 137.7, 135.7, 129.2, 127.6, 127.4, 73.0, 66.5, 47.9, 37.9, 21.0, 20.7, 14.0, $^1\text{H NMR}$ (400 MHz, C_6D_6) δ = 7.09 – 6.98 (m, 4H), 5.91 (ddd, $J = 1.2, 7.7, 15.4$ Hz, 1H), 5.43 (ddd,

$J = 1.2, 6.0, 15.6$ Hz, 1H), 4.19 – 4.09 (m, 1H), 3.54 – 3.31 (br, s., 2H), 3.48 (dd, $J_{AB} = 11.1$ Hz, $J_{AX} = 8.1$ Hz, 1H), 3.39 (dd, $J_{AB} = 11.1$ Hz, $J_{BX} = 2.8$ Hz, 1H), 3.17 (q, $J = 7.4$ Hz, 1H), 2.15 (s, 3H), 1.64 (q, $J = 7.6$ Hz, 2H), 1.34 – 1.16 (m, 2H), 0.86 (t, $J = 7.2$ Hz, 3H), ^{13}C NMR (101 MHz, C_6D_6) $\delta = 141.5, 136.7, 135.2, 1291., 128.1, 127.5, 72.9, 66.7, 48.0, 38.1, 20.7, 20.6, 13.9$, MS (EI, m/z) 234 (4, M^+), 204 (3), 185 (39), 160 (5), 147 (14), 143 (100), 131 (38), 105 (98), 91 (25), 77 (10), 55 (15), 41 (18). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3362, 2924, 2870 1513, 1457, 1378, 1075, 1021, 971, 874, 812, 721.

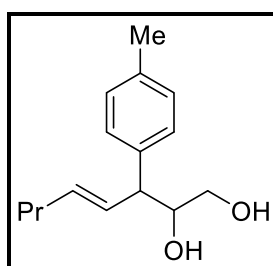


Figure 3.40. (*E*)-3-(*p*-Tolyl) oct-4-ene-1,2-diol

4ab: ^1H NMR (400 MHz, CDCl_3) $\delta = 7.17 - 7.05$ (m, 4 H), 5.75 - 5.60 (m, 2 H), 3.81 (ddd, $J = 3.1, 6.1, 8.8$ Hz, 1 H), 3.50 (d, $J = 3.1$ Hz, 1 H), 3.36 (dd, $J = 6.3, 11.3$ Hz, 1 H), 3.30 (t, $J = 8.2$ Hz, 1 H), 2.32 (s, 3 H), 2.07 - 2.00 (m, 2 H), 1.65 - 1.51 (m, 2 H), 1.40 (qd, $J = 7.1, 14.6$ Hz, 2 H), 0.88 (t, $J = 7.4$ Hz, 3 H), MS (EI, m/z) 234 (1>, M^+), 174(45), 143 (6), 131 (100), 118 (14), 91 (23), 77 (58), 61 (79), 43 (58). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3385, 2924, 2856, 1514, 1457, 1378, 1261, 1075, 1022, 878, 721.

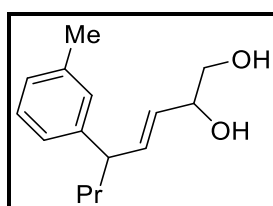


Figure 3.41. (*E*)-5-(*m*-Tolyl) oct-3-ene-1,2-diol

3ac: ^1H NMR (400 MHz, CDCl_3) $\delta = 7.17$ (t, $J = 8.6$ Hz, 1H), 7.03 - 6.90 (m, 3 H), 5.88 (ddd, $J = 1.2, 8.0, 15.5$ Hz, 1H), 5.43 (ddd, $J = 1.0, 6.3, 15.5$ Hz, 1 H), 4.22 - 4.11 (m, 1 H), 3.57 (dd, $J = 3.3, 11.2$ Hz, 1H), 3.41 (dd, $J = 7.6, 11.2$ Hz, 1 H), 3.20 (q, $J = 7.4$ Hz, 1 H), 2.67 (br. s., 2H), 2.32 (s, 3H), 1.65 (q, $J = 7.6$ Hz, 2H), 1.31 - 1.20 (m, 2H), 0.88 (t, $J = 7.4$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 144.4, 138.0, 137.6, 128.3,$

128.3, 127.7, 126.9, 124.5, 73.0, 66.5, 48.3, 38.0, 21.5, 20.7, 14.0. ^1H NMR (400MHz, C_6D_6) δ = 7.14 - 7.11 (m, 2 H), 7.05 - 7.02 (m, 2 H), 5.74 (ddd, J = 1.6, 7.6, 15.5 Hz, 1 H), 5.29 (ddd, J = 1.2, 5.9, 15.7 Hz, 1 H), 3.96 - 3.90 (m, 1 H), 3.40 (q, J = 7.4 Hz, 1 H), 3.32 (dd, J_{AB} = 11.0 Hz, J_{AX} = 7.9 Hz, 1H), 3.21 (dd, J_{AB} = 11.0 Hz, J_{BX} = 3.5 Hz, 1H), 2.24 (s, 3H) (Minor) 2.17 (s, 3 H) (Major), 1.94 (br. s, 2 H), 1.59 (q, J = 7.7 Hz, 2 H), 1.28 - 1.13 (m, 2 H), 0.83 (t, J = 7.4 Hz, 3 H). ^{13}C NMR (101MHz, C_6D_6) δ = 142.4, 135.9, 130.3, 128.4, 126.4, 126.3, 125.9, 72.7, 66.5, 43.4, 37.5, 20.6, 19.4, 13.9.), MS (EI, m/z) 234 (2, M^+), 216(1), 203(44), 185(63), 173(5), 143 (100), 131 (40), 105 (95), 91 (29), 77 (10), 55 (10), 41 (10). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3368, 2925, 2870, 1606, 1458, 1378, 1075, 1030, 971, 876, 704.

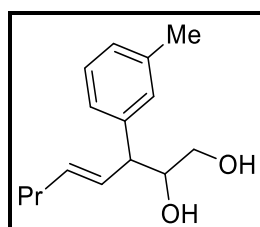


Figure 3.42. (*E*)-3-(*m*-Tolyl) oct-4-ene-1,2-diol

4ac: ^1H NMR (400 MHz, CDCl_3) δ = 7.23 - 7.17 (m, 1 H), 7.08 - 6.95 (m, 3 H), 5.78 - 5.60 (m, 2 H), 3.83 (ddd, J = 2.9, 6.1, 8.6 Hz, 1 H), 3.52 (dd, J = 3.1, 11.3 Hz, 1 H), 3.36 (dd, J = 6.3, 11.3 Hz, 1 H), 3.29 (t, J = 8.4 Hz, 1 H), 2.33 (s, 3 H), 2.23 (br. s., 0 H), 2.04 (q, J = 5.9 Hz, 2 H), 1.94 (br. s., 1 H), 1.41 (sxt, J = 7.8 Hz, 2 H), 0.89 (t, J = 7.4 Hz, 3 H). ^{13}C NMR (101 MHz, CDCl_3) δ = 141.0, 134.7, 129.6, 128.6, 128.5, 127.5, 124.8, 74.3, 64.2, 52.7, 34.7, 29.7, 22.4, 21.5, 13.7. MS (EI, m/z) 216(<1), 203(46), 159(7), 145(7), 115 (14), 105 (22), 91 (15), 77 (6), 55 (4), 43 (4). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3385, 2923, 2854, 1606, 1460, 1378, 1076, 1038, 970, 877, 783, 705.

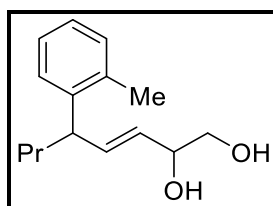


Figure 3.43. (*E*)-5-(*o*-Tolyl) oct-3-ene-1,2-diol

3ad: ^1H NMR (400 MHz, CDCl_3) $\delta = 7.20 - 7.06$ (m, 4H), 5.86 (ddd, $J = 1.2, 7.5, 15.6$ Hz, 1H), 5.42 (ddd, $J = 1.2, 6.5, 15.5$ Hz, 1H), 4.23 – 4.16 (m, 1H), 3.60 (dd, $J_{AB} = 11.2$ Hz, $J_{AX} = 7.5$ Hz, 1H), 3.52 (q, $J = 7.6$ Hz, 1H), 3.45 (dd, $J_{AB} = 11.2$ Hz, $J_{BX} = 3.5$ Hz, 1H), 2.31 (s, 3H), 1.76 – 1.59 (m, 2H), 1.40 – 1.16 (m, 2H), 0.89 (t, $J = 7.4$ Hz, 3H), ^{13}C NMR (101 MHz, CDCl_3) $\delta = 142.2, 137.2, 135.7, 130.4, 127.8, 126.3, 126.2, 125.9, 73.0, 66.5, 43.3, 37.5, 20.7, 19.6, 14.1$, ^1H NMR (400 MHz, C_6D_6) $\delta = 7.18 - 7.07$ (m, 2H), 7.06 – 6.98 (m, 2H), 5.80 (ddd, $J = 1.6, 7.6, 15.5$ Hz, 1H), 5.34 (ddd, $J = 1.2, 6.1, 15.5$ Hz, 1H), 4.08 – 4.01 (m, 2H), 3.43 (q, $J = 7.0$, 1H), 3.41 (dd, $J_{AB} = 13.4$ Hz, $J_{AX} = 6.9$ Hz, 1H), 3.31 (dd, $J_{AB} = 13.4$ Hz, $J_{AX} = 4.4$ Hz, 1H), 2.92 (br, s, 2H), 2.18 (s, 3H), 1.61 (q, $J = 7.6$ Hz, 2H), 1.30 – 1.18 (m, 2H), 0.84 (t, $J = 7.4$ Hz, 3H), ^{13}C NMR (101 MHz, C_6D_6) $\delta = 142.5, 136.0, 135.6, 130.4, 128.4, 126.5, 126.4, 125.9, 72.9, 66.7, 43.5, 37.6, 20.7, 19.5, 14.0$, MS (EI, m/z) 234 (4, M^+), 203 (45), 191 (8), 185 (66), 143 (100), 173 (4), 147 (13), 143 (100), 105 (90), 91 (32), 77 (10), 55 (10), 41 (10). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3364, 2926, 2870, 1460, 1378, 1074, 972, 874, 756, 726.

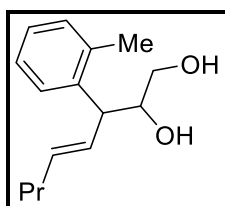


Figure 3.44. (*E*)-3-(*o*-Tolyl) oct-4-ene-1,2-diol

4ad: ^1H NMR (400 MHz, CDCl_3) $\delta = 7.21 - 7.08$ (m, 4H), 5.70 - 5.58 (m, 2H), 3.96 - 3.85 (m, 1H), 3.64 - 3.58 (m, 1H), 3.55 (d, $J = 11.0$ Hz, 1H), 3.38 - 3.27 (m, 1H), 2.34 (s, 3H), 2.08 - 1.99 (m, 2H), 1.39 (sxt, $J = 7.3$ Hz, 2H), 0.88 (t, $J = 7.4$ Hz, 3H), ^{13}C NMR (101 MHz, CDCl_3) $\delta = 139.4, 135.9, 134.8, 130.7, 129.8, 126.7, 126.3, 126.3, 74.1, 64.0, 47.6, 34.6, 22.4, 19.8, 13.7$, MS (EI, m/z) 234 (1, M^+), 174 (47), 159 (5), 145 (6), 131 (100), 119 (11), 115 (13), 105 (22), 91 (17), 77 (5), 65 (2), 55 (3), 43 (3). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3364, 2922, 2853, 1660, 1463, 1377, 969, 755, 724.

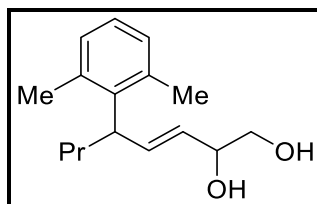


Figure 3.45. (*E*)-5-(2,6-Dimethylphenyl) oct-3-ene-1,2-diol

3ae: $^1\text{H NMR}$ (400 MHz, CDCl_3) $\delta = 7.04 - 6.92$ (m, 3H), 6.06 (dd, $J = 5.1, 15.7$ Hz, 1H), 5.35 (ddd, $J = 2.0, 6.6, 15.7$ Hz, 1H), 4.22 (dt, $J = 3.7, 6.9$ Hz, 1 H), 3.85 (q, $J = 7.4$ Hz, 1H), 3.61 (dd, $J = 3.5, 11.3$ Hz, 1H), 3.45 (dd, $J = 7.6, 11.2$ Hz, 1 H), 2.35 - 2.26 (m, 6H), 1.88 - 1.73 (m, 2H), 1.33 (s, 1 H), 1.24 - 1.12 (m, 1H), 0.89 (t, $J = 7.2$ Hz, 3H). $^1\text{H NMR}$ (400MHz, C_6D_6) $\delta = 7.03 - 6.91$ (m, 3 H), 5.91 (ddd, $J = 1.4, 5.1, 15.8$ Hz, 1 H), 5.25 (ddd, $J = 2.0, 6.2, 15.7$ Hz, 1 H), 3.97 - 3.88 (m, 1 H), 3.75 (q, $J = 6.3$ Hz, 1 H), 3.29 (dd, $J = 3.5, 11.0$ Hz, 1 H), 3.16 (dd, $J = 7.6, 10.8$ Hz, 1 H), 2.22 (s, 6 H), 1.69 (q, $J = 7.8$ Hz, 3 H), 1.27 - 1.07 (m, 2 H), 0.81 (t, $J = 7.4$ Hz, 3 H). MS (EI, m/z) 248 (4, M^+), 247 (5), 220 (28), 205 (70), 177 (26), 170 (18), 161 (17), 133 (32), 105 (26), 91 (18), 57 (92), 41 (100). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3368, 2923, 2854, 1465, 1378, 1075, 973, 874, 767.

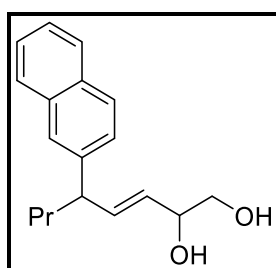


Figure 3.46. (*E*)-5-(Naphthalen-2-yl) oct-3-ene-1,2-diol

3af: $^1\text{H NMR}$ (400 MHz, CDCl_3) $\delta = 7.84 - 7.74$ (m, 3H), 7.60 (s, 1H), 7.52 - 7.39 (m, 2H), 7.31 (dd, $J = 1.8, 8.4$ Hz, 1H), 5.98 (ddd, $J = 1.2, 7.4, 15.7$ Hz, 1H), 5.48 (ddd, $J = 1.2, 6.3, 15.7$ Hz, 1H), 4.23 - 4.14 (m, 1H), 3.58 (dd, $J_{AB} = 11.2$ Hz, $J_{AX} = 7.5$ Hz, 1H), 3.43 (dd, $J_{AB} = 11.2$ Hz, $J_{BX} = 3.5$ Hz, 1H), 3.42 (q, $J = 7.4$ Hz, 1H), 2.54 (d, $J = 8.6$ Hz, 1 H), 1.78 (q, $J = 7.6$ Hz, 1H), 1.37 - 1.18 (m, 2H), 0.91 (q, $J = 7.2$ Hz, 2H), $^{13}\text{C NMR}$ (101 MHz, CDCl_3) $\delta = 141.7, 137.4, 133.6, 132.2, 128.1, 128.1, 127.6, 127.6, 126.1, 126.0, 125.9, 125.4, 73.0, 66.5, 48.4, 37.7, 20.7, 14.0$, $^1\text{H NMR}$ (400 MHz, C_6D_6) $\delta = 7.66$ (dd, $J = 8.2, 17.6$ Hz, 3H), 7.56 (s, 1H), 7.31 - 7.22 (m, 3H), 5.95 (ddd, $J = 1.2,$

7.5, 15.6 Hz, 1H), 5.44 (ddd, $J = 1.0, 6.0, 15.4$ Hz, 1H), 4.13 (m, 1H), 3.47 (dd, $J_{AB} = 11.3$ Hz, $J_{AX} = 8.1$ Hz, 1H), 3.38 (dd, $J_{AB} = 11.3$ Hz, $J_{BX} = 3.2$ Hz, 1H), 3.29 (q, $J = 7.3$ Hz, 1H), 1.72 – 1.63 (m, 2H), 1.34 – 1.13 (m, 3H), 0.85 (t, $J = 7.4$ Hz, 3H), ^{13}C NMR (101 MHz, C_6D_6) $\delta = 142.0, 136.2, 133.9, 132.5, 128.6, 128.1, 127.6, 126.2, 126.0, 125.8, 125.2, 72.9, 66.6, 48.5, 37.8, 20.7, 13.9$, MS (EI, m/z) 270 (36, M^+), 239 (18), 221 (42), 209 (10), 195 (10), 179 (100), 167 (54), 152 (20), 141 (56), 128 (19), 115 (10), 55 (4), 41 (4), 31 (4). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3365, 2927, 2870, 1600, 1457, 1377, 1260, 1024, 815, 746.

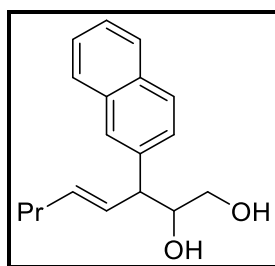


Figure 3.47. (*E*)-3-(Naphthalen-2-yl) oct-4-ene-1,2-diol

4af: ^1H NMR (400 MHz, CDCl_3) $\delta = 7.85 - 7.75$ (m, 3H), 7.66 (d, $J = 1.6$ Hz, 1H), 7.51 – 7.42 (m, 3H), 7.35 (dd, $J = 1.6, 8.6$ Hz, 1H), 5.82 (tdd, $J = 1.2, 8.6, 15.3$ Hz, 1H), 5.71 (td, $J = 6.3, 15.3$ Hz, 1H), 3.97 (ddd, $J = 2.9, 6.0, 8.7$ Hz, 1H), 3.55 (dd, $J_{AB} = 11.4$ Hz, $J_{AX} = 6.3$ Hz, 1H), 3.53 (t, $J = 8.6$ Hz, 1H), 3.40 (dd, $J_{AB} = 11.4$ Hz, $J_{BX} = 3.1$ Hz, 1H), 2.06 (q, $J = 7.7$ Hz, 2H), 1.41 (qd, $J = 7.4, 14.7$ Hz, 2H), 0.89 (t, $J = 7.4$ Hz, 3H), ^{13}C NMR (101 MHz, CDCl_3) $\delta = 138.6, 135.0, 133.6, 130.6, 129.4, 128.5, 127.6, 127.6, 126.4, 126.2, 126.1, 125.7, 74.3, 64.2, 52.7, 34.7, 22.4, 13.7$, MS (EI, m/z) 270 (1, M^+), 253 (1), 236 (1), 210 (54), 193 (6), 178 (20), 167 (100), 152 (20), 141 (22), 128 (15), 115 (13), 81 (6), 44 (8), 32 (10). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3385, 2924, 2854, 1724, 1600, 1463, 1377, 1260, 1075, 971, 816, 746.

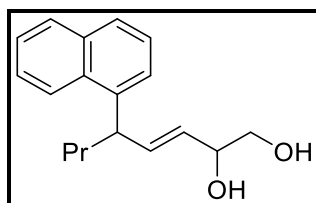


Figure 3.48. (*E*)-5-(Naphthalen-1-yl) oct-3-ene-1,2-diol

3ag: ^1H NMR (400 MHz, C_6D_6) δ = 8.08 (d, J = 8.6 Hz, 1H), 7.67 (dd, J = 1.0, 7.6 Hz, 1H), 7.59 – 7.52 (m, 1H), 7.40 – 7.22 (m, 3H), 7.16 (s, 1H), 5.96 (dd, J = 7.4, 15.3 Hz, 1H), 5.43 (dd, J = 5.7, 15.5 Hz, 1H), 4.07 (m, 1H), 4.02 (q, J = 7.8, 1H), 3.44 – 3.36 (m, 1H), 3.35 – 3.14 (m, 3H), 1.84 – 1.66 (m, 2H), 1.38 – 1.15 (m, 2H), 0.82 (t, J = 7.4 Hz, 3H), ^{13}C NMR (101 MHz, C_6D_6) δ = 140.7, 135.9, 134.3, 131.9, 129.0, 128.8, 126.7, 125.6, 125.6, 125.3, 124.1, 123.4, 72.9, 66.6, 42.7, 37.7, 20.9, 14.0, MS (EI, m/z) 270 (25), 239 (13), 221 (35), 179 (100), 167 (88), 165 (96), 152 (53), 141 (78), 41 (56). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3361, 2928, 2870, 1596, 1509, 1457, 1365, 1074, 1027, 972, 875, 777.

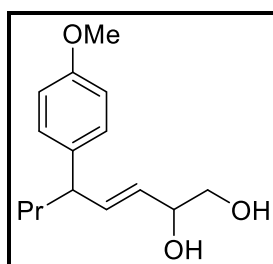


Figure 3.49. (*E*)-5-(4-Methoxyphenyl) oct-3-ene-1,2-diol

3ah: ^1H NMR (400 MHz, CDCl_3) δ = 7.12 – 7.03 (m, 2H), 6.88 – 6.79 (m, 2H), 5.88 (ddd, J = 1.2, 7.6, 15.5 Hz, 1H), 5.42 (ddd, J = 1.2, 6.4, 15.6 Hz, 1H), 4.23 – 4.15 (m, J = 3.5 Hz, 1H), 3.78 (s, 3H), 3.59 (dd, J_{AB} = 11.3 Hz, J_{AX} = 7.9 Hz, 1H), 3.36 (dd, J_{AB} = 11.3 Hz, J_{BX} = 3.5 Hz, 1H), 3.21 (q, J = 7.4 Hz, 1H), 2.31 (br, s, 2H), 1.72 – 1.56 (m, 2H), 1.32 – 1.15 (m, 2H), 0.88 (t, J = 7.4 Hz, 3H), ^{13}C NMR (101 MHz, CDCl_3) δ = 157.9, 137.9, 136.3, 128.4, 127.5, 113.8, 73.0, 66.5, 55.2, 47.4, 38.0, 20.6, 14.0, ^1H NMR (400 MHz, C_6D_6) δ = 7.06 – 6.99 (m, 2H), 6.86 – 6.79 (m, 2H), 5.86 (dd, J = 7.6, 15.5 Hz, 1H), 5.38 (dd, J = 6.1, 15.5 Hz, 1H), 4.08 – 4.01 (m, 1H), 3.42 (dd, J_{AB} = 10.7 Hz, J_{AX} = 7.5 Hz, 1H), 3.35 (s, 3H), 3.32 (dd, J_{AB} = 10.7 Hz, J_{BX} = 3.5 Hz, 1H), 3.13 (q, J = 7.4 Hz, 1H), 2.45 (br, s, 2H), 1.60 (q, J = 7.6 Hz, 2H), 1.33 – 1.15 (m, 2H), 0.86 (t, J = 7.4 Hz, 3H), ^{13}C NMR (101 MHz, C_6D_6) δ = 158.3, 136.9, 136.4, 128.5, 128.0, 113.9, 72.8, 66.6, 54.4, 47.5, 38.1, 20.7, 13.9, MS (EI, m/z) 250 (16, M^+), 232 (3), 219 (5), 207 (35), 201 (24), 189 (10), 176 (6), 171 (8), 159 (100), 147 (38), 135 (10), 128 (7), 121 (54), 115 (17), 77 (10), 65 (5), 55 (7), 41 (7), 31 (7).

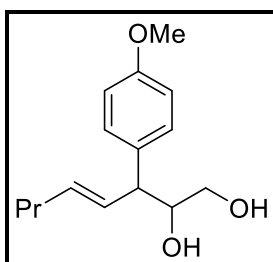


Figure 3.50. (*E*)-3-(4-Methoxyphenyl) oct-4-ene-1,2-diol

4ah: $^1\text{H NMR}$ (400 MHz, CDCl_3) $\delta = 7.15 - 7.10$ (m, 2H), 6.89 – 6.82 (m, 2H), 5.69 (dd, $J_{AB} = 15.3$ Hz, $J_{AX} = 8.1$ Hz, 1H), 5.64 (dd, $J_{AB} = 15.3$ Hz, $J_{BX} = 5.2$ Hz, 1H), 3.79 (s, 3H), 3.83 – 3.75 (m, 1H), 3.52 (dd, $J_{AB} = 11.4$ Hz, $J_{AX} = 6.5$ Hz, 1H), 3.36 (dd, $J_{AB} = 11.4$ Hz, $J_{BX} = 2.9$ Hz, 1H), 3.30 (t, $J = 8.2$ Hz, 1 H), 2.04 (q, $J = 7.2$, 2H), 1.40 (sxt, $J = 6.7$ Hz, 2H), 0.89 (t, $J = 7.4$ Hz, 3H). $^1\text{H NMR}$ (400 MHz, C_6D_6) $\delta = 7.04 - 6.99$ (m, 2H), 6.78 – 6.73 (m, 2H), 5.68 (tdd, $J = 1.5, 8.8, 15.3$ Hz, 1H), 5.44 (td, $J = 7.0, 15.3$ Hz, 1H), 3.71 – 3.65 (m, 1H), 3.46 (dd, $J = 3.1, 11.3$ Hz, 1H), 3.36 – 3.26 (m, 2H), 3.30 (s, 3H), 1.87 (q, $J = 6.7$ Hz, 2H), 1.24 (sxt, $J = 7.4$ Hz, 2H), 0.80 (t, $J = 7.0$ Hz, 3H).

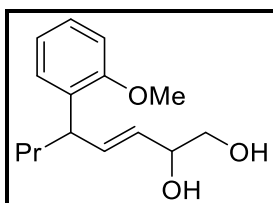


Figure 3.51. (*E*)-5-(2-Methoxyphenyl) oct-3-ene-1,2-diol

3ai: $^1\text{H NMR}$ (400 MHz, CDCl_3) $\delta = 7.20 - 7.10$ (m, 2 H), 6.91 (dt, $J = 1.2, 7.4$ Hz, 1 H), 6.85 (dd, $J = 0.8, 8.2$ Hz, 1 H), 5.95 (ddd, $J = 1.2, 7.8, 15.7$ Hz, 1 H), 5.46 (ddd, $J = 1.2, 6.5, 15.5$ Hz, 1 H), 4.24 - 4.16 (m, 1 H), 3.81 (s, 3 H), 3.75 (q, $J = 7.8$ Hz, 1 H), 3.65 - 3.55 (m, 1 H), 3.53 - 3.41 (m, 1 H), 1.97 (br. s., 1 H), 1.88 (br. s., 1 H), 1.67 (q, $J = 7.6$ Hz, 2 H), 1.33 - 1.19 (m, 2 H), 0.88 (t, $J = 7.4$ Hz, 3 H). $^1\text{H NMR}$ (400 MHz, C_6D_6) $\delta = 7.20 - 7.13$ (m, 1 H), 7.07 (dt, $J = 2.0, 7.8$ Hz, 1 H), 6.91 (t, $J = 7.4$ Hz, 1 H), 6.57 (d, $J = 8.2$ Hz, 1 H), 5.93 (dd, $J = 7.8, 15.7$ Hz, 1 H), 5.44 (dd, $J = 6.1, 15.5$ Hz, 1 H), 3.98 - 3.89 (m, 2 H), 3.32 (s, 3 H), 3.32 (dd, $J_{AB} = 10.7$ Hz, $J_{AX} = 7.3$ Hz, 1H), 3.22 (dd, $J_{AB} = 10.7$ Hz, $J_{BX} = 3.3$ Hz, 1H), 3.22 (dd, $J = 7.0, 11.3$ Hz, 1 H), 1.70 (qd, $J = 6.6, 9.3$ Hz, 2 H), 1.42 - 1.20 (m, 2 H), 0.88 (t, $J = 7.2$ Hz, 3 H). $^{13}\text{C NMR}$ (101 MHz, C_6D_6) $\delta = 157.0, 136.0, 132.9, 128.5, 126.9, 120.7, 110.6, 72.8, 66.5, 54.6, 54.6, 40.8, 37.2, 20.8, 13.9$. MS

(EI, m/z) 250 (5, M⁺), 219 (12), 201 (31), 159 (62), 121 (100), 91 (61), 77 (22), 43 (63), 41 (76). FTIR ($\nu_{\max}/\text{cm}^{-1}$): 3372, 2925, 1491, 1240, 1029, 874, 752.

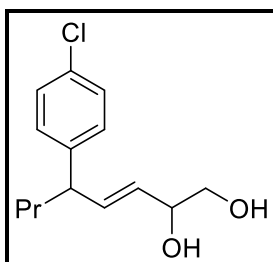


Figure 3.52. (*E*)-5-(4-Chlorophenyl) oct-3-ene-1,2-diol

3aj: ¹H NMR (400 MHz, CDCl₃) δ = 7.34 – 7.18 (m, 2H), 7.13 – 7.03 (m, 2H), 5.86 (ddd, J = 1.2, 7.6, 15.5 Hz, 1H), 5.42 (ddd, J = 1.2, 6.3, 15.7 Hz, 1H), 4.23 – 4.15 (m, 1H), 3.59 (dd, J_{AB} = 11.3 Hz, J_{AX} = 8.1 Hz, 1H), 3.43 (dd, J_{AB} = 11.3 Hz, J_{BX} = 3.3 Hz, 1H), 3.23 (q, J = 7.4 Hz, 1H), 2.48 (br, s., 2H), 1.72 – 1.56 (m, 2H), 1.33 – 1.12 (m, 2H), 0.87 (t, J = 7.4 Hz, 3H), ¹³C NMR (101 MHz, CDCl₃) δ = 142.8, 136.9, 131.8, 128.9, 128.6, 128.2, 72.9, 66.5, 47.6, 37.8, 20.5, 13.9. ¹H NMR (400 MHz, C₆D₆) δ = 7.19 – 7.11 (m, 2H), 6.86 – 6.77 (m, 2H), 5.77 (ddd, J = 1.2, 7.8, 15.7 Hz, 1H), 5.34 (ddd, J = 1.0, 5.9, 15.5 Hz, 1H), 4.12 (m, 1H), 3.47 (dd, J_{AB} = 11.2 Hz, J_{AX} = 7.9 Hz, 1H), 3.36 (dd, J_{AB} = 11.2 Hz, J_{BX} = 3.1 Hz, 1H), 3.01 (q, J = 7.4 Hz, 1H), 1.56 – 1.40 (m, 2H), 1.24 – 1.04 (m, 2H), 0.82 (t, J = 7.4 Hz, 3H), ¹³C NMR (101 MHz, C₆D₆) δ = 142.9, 135.8, 131.8, 128.9, 128.5, 128.5, 72.8, 66.6, 47.6, 37.7, 20.5, 13.8. MS (EI, m/z) 254 (4, M⁺), 236 (1), 223 (72), 218 (1), 205 (22), 193 (5), 179 (8), 163 (42), 151 (30), 139 (18), 125 (100), 115 (42), 103 (6), 89 (6), 77 (6), 57 (14), 41 (8), 31 (8). FTIR ($\nu_{\max}/\text{cm}^{-1}$): 3363, 2926, 2870, 1491, 1365, 1090, 1014, 821, 719.

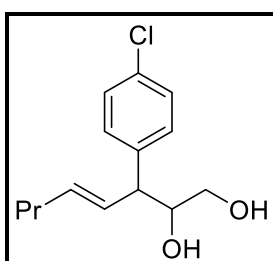


Figure 3.53. (*E*)-3-(4-Chlorophenyl) oct-4-ene-1,2-diol

4aj: ^1H NMR (400 MHz, CDCl_3) $\delta = 7.31 - 7.26$ (m, 2H), 7.18 – 7.12 (m, 2H), 5.67 (dd, $J_{AB} = 15.6$ Hz, $J_{AX} = 15.6$ Hz, 1H), 5.64 (dd, $J_{AB} = 15.6$ Hz, $J_{AX} = 6.9$ Hz, 1H), 3.80 (ddd, $J = 3.1, 6.0, 8.5$ Hz, 1 H), 3.53 (dd, $J_{AB} = 11.4$ Hz, $J_{AX} = 6.1$ Hz, 1H), 3.48 (q, $J = 7.0$ Hz, 1 H), 3.34 (dd, $J_{AB} = 11.4$ Hz, $J_{BX} = 2.9$ Hz, 1H), 3.37 – 3.31 (m, 2H), 2.08 – 2.00 (m, 2H), 1.40 (sxt, $J = 7.4$ Hz, 2H), 0.88 (t, $J = 7.2$ Hz, 3H). MS (EI, m/z) 223 (1), 194 (54), 177 (1), 159 (16), 151 (100), 138 (24), 125 (42), 115 (40), 99 (3), 91 (6), 77 (7), 61 (10), 55 (8), 43 (8). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3361, 2922, 2853, 1463, 1377, 1074, 774.

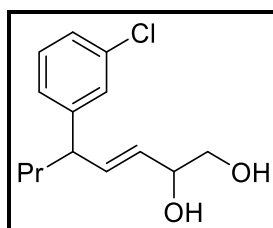


Figure 3.54. (*E*)-5-(3-Chlorophenyl) oct-3-ene-1,2-diol

3ak: ^1H NMR (400 MHz, C_6D_6) $\delta = 7.23 - 7.12$ (m, 2 H), 7.08 - 7.02 (m, 1 H), 6.91 - 6.79 (m, 2 H), 5.73 (tdd, $J = 1.2, 7.8, 16.0$ Hz, 1 H), 5.33 (ddd, $J = 1.2, 5.9, 15.3$ Hz, 1 H), 4.09 - 4.00 (m, 1 H), 3.42 (ddd, $J = 0.8, 3.1, 11.0$ Hz, 1 H), 3.31 (dt, $J = 1.2, 9.6$ Hz, 1 H), 3.19-3.66 (br. s, 2 H), 2.99 (q, $J = 7.4$ Hz, 1 H), 1.46 (q, $J = 7.4$ Hz, 2 H), 1.24 - 1.02 (m, 2 H), 0.79 (t, $J = 7.2$ Hz, 3 H). ^{13}C NMR (101 MHz, C_6D_6) $\delta = 146.9, 135.4, 134.3, 129.7, 128.9, 127.7, 126.3, 125.7, 72.7, 66.5, 48.1, 37.6, 20.5, 13.7$. MS (EI, m/z) 254 (1, M^+), 223 (23), 205 (7), 167 (20), 125 (70), 115 (100), 103 (19), 89 (6), 77 (36), 57 (58), 43 (60), 41 (90). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3354, 2926, 2871, 1595, 1571, 1077, 1028, 971, 874, 782, 696.

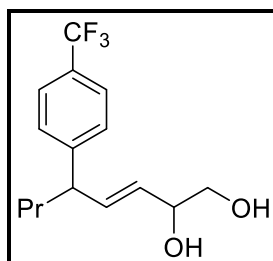


Figure 3.55. (*E*)-5-(4-(Trifluoromethyl) phenyl) oct-3-ene-1,2-diol

3am: ^1H NMR (400 MHz, CDCl_3) $\delta = 7.54$ (d, $J = 7.8$ Hz, 2H), 7.27 (d, $J = 8.6$ Hz, 2H), 5.89 (ddd, $J = 1.4, 7.7, 15.6$ Hz, 1H), 5.47 (ddd, $J = 1.2, 6.3, 15.7$ Hz, 1H), 4.24

– 4.17 (m, 1H), 3.61 (dd, $J_{AB} = 11.1$ Hz, $J_{AX} = 7.7$ Hz, 1H), 3.45 (dd, $J_{AB} = 11.1$ Hz, $J_{BX} = 3.3$ Hz, 1H), 3.33 (q, $J = 7.6$ Hz, 1 H), 2.28 (br, s, 2H), 1.76 – 1.61 (m, 2H), 1.35 – 1.12 (m, 2H), 0.88 (t, $J = 7.8$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 148.4, 148.4, 136.4, 128.6, 128.4, 127.9, 125.4, 72.8, 66.5, 48.2, 37.7, 20.6, 13.9$. ^1H NMR (400 MHz, C_6D_6) $\delta = 7.38$ (d, $J = 8.2$ Hz, 2H), 6.90 (d, $J = 7.4$ Hz, 2H), 5.78 – 5.67 (m, 1H), 5.36 – 5.26 (m, 1H), 4.10 – 3.98 (m, 1H), 3.46 – 3.36 (m, 1H), 3.34 – 3.25 (m, 1 H), 3.02 (q, $J = 7.4$ Hz, 1H), 1.45 (td, $J = 7.5, 15.5$ Hz, 2H), 1.20 – 0.99 (m, 2H), 0.80 (t, $J = 7.4$ Hz, 3H), ^{13}C NMR (101 MHz, C_6D_6) $\delta = 148.6, 135.2, 129.0, 128.9, 127.8, 125.3, 123.4, 72.6, 66.5, 48.1, 37.6, 20.5, 13.7$. MS (EI, m/z) 269 (11), 257 (100), 237 (11), 213 (7), 201 (43), 181 (18), 165 (10), 159 (71), 145 (12), 127 (12), 115 (12), 95 (1), 87 (10), 71 (5), 57 (30), 43 (10), 31 (8). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3381, 2926, 1324, 1120, 1067, 972, 835.

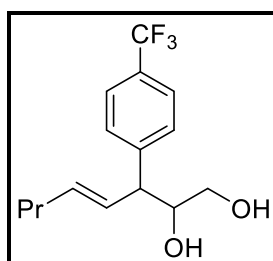


Figure 3.56. (*E*)-3-(4-(Trifluoromethyl) phenyl) oct-4-ene-1,2-diol

4am: ^1H NMR (400 MHz, C_6D_6) $\delta = 7.32$ (d, $J = 7.8$ Hz, 2H), 6.93 (d, $J = 8.2$ Hz, 2H), 5.56 (tdd, $J = 1.5, 8.8, 15.3$ Hz, 1H), 5.35 (td, $J = 6.7, 15.5$ Hz, 1H), 3.51 (dt, $J = 3.5, 6.8$ Hz, 1H), 3.25 (dd, $J_{AB} = 10.8$ Hz, $J_{AX} = 6.9$ Hz, 1H), 3.18 (t, $J = 8.2$ Hz, 1H), 3.11 (dd, $J_{AB} = 10.8$ Hz, $J_{BX} = 2.5$ Hz, 1H), 1.85 (dq, $J = 1.6, 6.7$ Hz, 2H), 1.23 (q, $J = 7.4$ Hz, 2H), 0.80 (t, $J = 7.4$ Hz, 3H). MS (EI, m/z) 228 (70), 208 (65), 185 (100), 172 (93), 165 (35), 159 (54), 145 (11), 129 (20), 103 (4), 91 (6), 69 (12), 61 (54), 55 (18), 43 (20), 31 (6). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3388, 2925, 2854, 1464, 1326, 1125, 1068, 836.

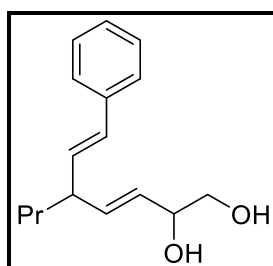


Figure 3.57. (*E*)-5-((*E*)-Styryl) oct-3-ene-1,2-diol

3an: ^1H NMR (400 MHz, C_6D_6) $\delta = 7.33 - 7.24$ (m, 2H), 7.20 – 7.11 (m, 2H), 7.09 – 7.01 (m, 1H), 6.45 (d, $J = 16.0$ Hz, 1H) (minor), 6.36 (d, $J = 16.0$ Hz, 1H) (major), 6.19 (dd, $J = 7.8, 16.0$ Hz, 1H) (minor), 6.05 (dd, $J = 7.8, 16.0$ Hz, 1H) (major), 5.75 (ddd, $J = 1.2, 7.4, 15.7$ Hz, 1H), 5.48 (ddd, $J = 1.2, 6.0, 15.6$ Hz, 1H), 4.29 – 4.16 (m, 1H), 3.57 (dd, $J_{AB} = 11.3$ Hz, $J_{AX} = 8.1$ Hz, 1H), 3.46 (dd, $J_{AB} = 11.3$ Hz, $J_{BX} = 3.2$ Hz, 1H), 2.78 (quin, $J = 7.3$ Hz, 1H), 1.45 – 1.36 (m, 2H), 1.36 – 1.24 (m, 2H), 0.89 (t, $J = 6.7$ Hz, 3H) (major), 0.86 (t, $J = 6.7$ Hz, 3H) (minor), ^{13}C NMR (101 MHz, C_6D_6) $\delta = 135.2, 132.9, 129.8, 128.6, 128.4, 127.0, 126.3, 126.2, 73.0, 66.7, 45.7, 37.1, 20.4, 13.9$, MS (EI, m/z) 228 (2), 215 (6), 197 (10), 185 (78), 167 (18), 155 (30), 143 (65), 129 (60), 115 (50), 107 (20), 91 (100), 77 (15), 55 (10), 43 (10). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3378, 2924, 2852, 1464, 1260, 1092, 1025, 799.

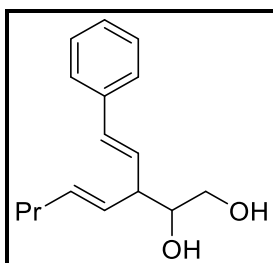


Figure 3.58. (*E*)-3-((*E*)-Styryl) oct-4-ene-1,2-diol

4an: ^1H NMR (400 MHz, CDCl_3) $\delta = 7.39 - 7.28$ (m, 4H), 7.26 – 7.20 (m, 1H), 6.47 (d, $J = 16.0$ Hz, 1H), 6.14 (dd, $J = 8.2, 16.0$ Hz, 1H), 5.66 (td, $J = 6.7, 15.3$ Hz, 1H), 5.50 (dd, $J = 8.6, 15.3$ Hz, 1H), 3.76 (d, $J = 11.3$ Hz, 1H), 3.67 (t, $J = 8.6$ Hz, 1H), 3.64 – 3.57 (m, 1H), 3.04 (q, $J = 7.8$ Hz, 1H), 2.07 (q, $J = 7.0$ Hz, 2H), 1.43 (sxt, $J = 7.4$ Hz, 2H), 0.92 (t, $J = 7.4$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 137.0, 134.6, 131.8, 128.6, 128.5, 128.2, 127.4, 126.2, 110.0, 73.7, 64.5, 50.1, 34.8, 22.4, 13.7$. MS (EI, m/z) 246 (1, M^+), 215 (2), 185 (72), 165 (4), 155 (10), 143 (95), 129 (66), 115 (42), 105 (12), 91 (100), 77 (12), 65 (8), 55 (10), 43 (10), 31 (4).

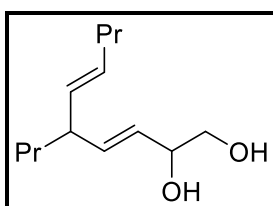


Figure 3.59. (*3E,6E*)-5-Propyldeca-3,6-diene-1,2-diol

3ao: ^1H NMR (400 MHz, C_6D_6) δ = 5.67 (ddd, J = 1.2, 7.4, 14.5 Hz, 1H), 5.43 (q, J = 6.3 Hz, 1 H), 5.39 (t, J = 6.3 Hz, 1H), 5.30 (dd, J = 7.8, 15.7 Hz, 1H), 4.15 – 4.06 (m, 1H), 3.48 (dd, J_{AB} = 11.0 Hz, J_{AX} = 7.9 Hz, 1H), 3.37 (dd, J_{AB} = 11.0 Hz, J_{BX} = 3.5 Hz, 1H), 3.17 (q, J = 7.4 Hz, 1H), 2.65 (td, J = 6.8, 13.7 Hz, 1H), 2.55 (br, s, 2H), 1.96 (q, J = 6.8 Hz, 2H), 1.42 – 1.25 (m, 6H), 0.98 – 0.81 (m, 6H), ^{13}C NMR (101MHz, C_6D_6) δ = 136.1, 133.1, 130.0, 128.1, 73.0, 66.7, 45.3, 37.3, 34.7, 22.7, 20.4, 13.9, 13.5. MS (EI, m/z) 195 (1), 181 (26), 151 (27), 107 (29), 95 (38), 91 (22), 81 (43), 67 (79), 55 (66), 43 (79), 41 (100). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3368, 2926, 2871, 1458, 1378, 1074, 1029, 970, 874.

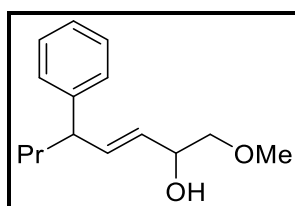


Figure 3.60. (*E*)-1-Methoxy-5-phenyloct-3-en-2-ol

3ba: ^1H NMR (400 MHz, CDCl_3) δ : 7.32–7.26 (m, 2H), 7.22–7.14 (m, 3H), 5.92 (dd, J = 7.8, 15.7 Hz, 1H), 5.44 (dd, J = 6.3, 15.7 Hz, 1H), 4.32–4.24 (m, 1H), 3.41–3.36 (m, 1H), 3.38 (s, 3H), 3.29–3.21 (m, 2 H), 2.37 (br, s., 1H), 1.68 (q, J = 7.4 Hz, 2H), 1.31–1.22 (m, 2H), 0.88 (t, J = 7.2 Hz, 3H), ^{13}C NMR (101 MHz, CDCl_3) δ : 144.5, 137.2, 128.4, 127.6, 127.6, 126.1, 76.7, 71.1, 59.0, 48.3, 38.0, 20.6, 14.0. ^1H NMR (400 MHz, C_6DC_6) δ : 7.21–7.16 (m, 2 H), 7.13–7.01 (m, 3 H), 5.93 (ddd, J = 1.4, 7.8, 15.5 Hz, 1H) (major), 5.92 (ddd, J = 1.4, 7.8, 15.5 Hz, 1H) (minor), 5.45 (ddd, J = 0.8, 6.3, 15.3 Hz, 1H), 4.22 (m, 1H), 3.16 (q, J = 7.8 Hz, 1H), 3.08 (dd, J_{AB} = 8.6 Hz, J_{AX} = 6.2 Hz, 1H), 3.10 (dd, J_{AB} = 8.6 Hz, J_{BX} = 2.4 Hz, 1H), 3.00 (s, 3H) (minor), 2.99 (s, 3 H) (major), 2.29 (br, s., 1H), 1.60 (q, J = 7.5 Hz, 2H), 1.28–1.17 (m, 2H), 0.83 (t, J = 7.5 Hz, 3 H). ^{13}C NMR (101 MHz, C_6D_6) δ : 144.7, 136.1, 128.4, 128.4, 127.6, 126.0, 76.8, 70.7, 58.2, 48.5, 38.1, 20.6, 13.8. MS (EI, m/z): 234 (M^+ , 1), 216 (1), 189 (54), 171 (29), 145 (15), 134 (24), 129 (44), 115 (32), 105 (20), 101 (18), 91 (100), 77 (7), 69 (6), 57 (13), 45 (26). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3435, 2926, 1601, 1452, 1120, 969, 699, 614.

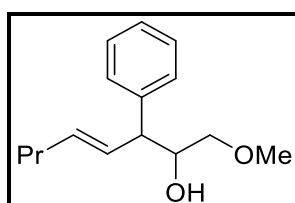


Figure 3.61. (*E*)-1-Methoxy-3-phenyloct-4-en-2-ol

4ba: ^1H NMR (400 MHz, CDCl_3) δ : 7.34–7.28 (m, 3H), 7.25–7.19 (m, 2H), 5.76 (dd, $J = 9.8, 15.3$ Hz, 1H), 5.61 (td, $J = 6.6, 15.3$ Hz, 1H), 3.98–3.91 (m, 1H), 3.36 (t, $J = 8.2$ Hz, 1H), 3.33–3.26 (m, 4 H), 3.14 (dd, $J = 6.8, 9.6$ Hz, 1H), 2.04 (q, $J = 7.0$ Hz, 2H), 1.39 (sex, $J = 7.2$ Hz, 2H), 0.88 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ : 141.8, 134.0, 129.4, 128.6, 127.9, 126.6, 74.5, 73.2, 59.0, 52.4, 34.7, 22.4, 13.7. ^1H NMR (400 MHz, C_6D_6) δ : 7.22–7.03 (m, 5H), 5.94 (dd, $J = 8.4, 15.4$ Hz, 1H) (major), 5.68 (dd, $J = 8.4, 15.4$ Hz, 1H) (minor), 5.48 (td, $J = 7.0, 15.4$ Hz, 1H), 4.04 (m, 1H) (minor), 3.99 (m, 1H) (major), 3.43 (t, $J = 7.6$ Hz, 1H), 3.18 (dd, $J_{AB} = 9.4$ Hz, $J_{AX} = 7.1$ Hz, 1H), 3.10 (dd, $J_{AB} = 9.4$ Hz, $J_{BX} = 3.4$ Hz, 1H), 3.03 (s, 3H) (minor), 2.97 (s, 3 H) (major), 2.25 (br, s., 1H), 1.93 (q, $J = 7.2$ Hz, 2H), 1.28 (qd, $J = 7.2, 14.6$ Hz, 2H), 0.82 (t, $J = 7.2$ Hz, 3H). ^{13}C NMR (101 MHz, C_6D_6) δ : 142.5, 132.7, 130.3, 128.4, 128.2, 126.3, 74.8, 73.2, 58.2, 52.2, 34.7, 22.4, 13.4. MS (EI, m/z): 217 (48), 199 (5), 185 (15), 173 (26), 159 (34), 119 (52), 113 (7), 104 (100), 91 (11), 81 (8), 67 (7), 53 (10), 45 (20). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3447, 2923, 2852, 1733, 1454, 1123, 971, 700.

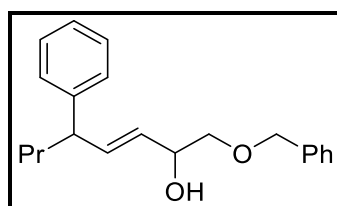


Figure 3.62. (*E*)-1-(Benzyloxy)-5-phenyloct-3-en-2-ol

3ca: ^1H NMR (400 MHz, CDCl_3) δ = 7.42 – 7.27 (m, 7H), 7.25 – 7.16 (m, 3H), 5.94 (dd, $J = 7.8, 15.7$ Hz, 1H), 5.47 (dd, $J = 6.5, 15.5$ Hz, 1H), 4.57 (s, 2H), 4.38 – 4.30 (m, 1H), 3.51 (dd, $J_{AB} = 9.5$ Hz, $J_{AX} = 8.2$ Hz, 1H), 3.37 (dd, $J_{AB} = 9.5$ Hz, $J_{BX} = 3.5$ Hz, 1H), 3.28 (q, $J = 7.2$ Hz, 1H), 2.55 (br, s., 1H), 1.70 (q, $J = 7.4$ Hz, 2H), 1.40 – 1.17 (m, 2H), 0.91 (t, $J = 7.4$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ = 144.5, 137.9, 137.2, 128.5, 128.4, 127.8, 127.8, 127.7, 127.6, 126.2, 74.3, 73.3, 71.3, 48.4, 38.0, 20.7, 14.1. ^1H NMR (400 MHz, C_6D_6) δ = 7.21 – 7.12 (m, 6H), 7.12 – 7.03 (m, 4H), 5.93 (ddd, $J = 1.2, 7.8, 15.7$ Hz, 1H), 5.45 (ddd, $J = 1.2, 5.9, 15.7$ Hz, 1H), 4.29 – 4.24 (m, 1H), 4.22 (s, 2H), 3.23 (dd, $J_{AB} = 9.4$ Hz, $J_{AX} = 8.1$ Hz, 1H), 3.19 (dd, $J_{AB} = 9.4$ Hz, $J_{BX} = 3.6$ Hz, 1H), 3.15 (q, $J = 7.4$ Hz, 1 H), 2.39 (br, s., 1H), 1.59 (q, $J = 7.6$ Hz, 2H), 1.31 – 1.12 (m, 2H), 0.82 (t, $J = 7.4$ Hz, 3H). ^{13}C NMR (101 MHz, C_6D_6) δ = 144.7, 138.3, 136.0, 128.5, 128.4,

128.2, 127.6, 127.5, 126.0, 74.5, 72.8, 70.9, 48.5, 38.1, 20.7, 13.8, MS (EI, m/z) 310 (1>, M⁺), 189 (14), 177 (15), 133 (22), 91 (100). FTIR ($\nu_{\max}/\text{cm}^{-1}$): 3436, 2925, 2857, 1453, 1103, 970, 698.

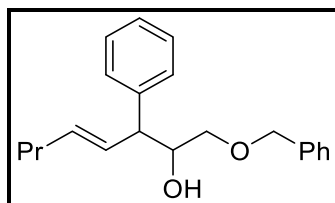


Figure 3.63. (*E*)-1-(Benzyloxy)-3-phenyloct-4-en-2-ol

4ca: ¹H NMR (400 MHz, CDCl₃) δ = 7.38 – 7.27 (m, 8H), 7.25 – 7.17 (m, 2H), 5.77 (dd, J = 9.0, 15.3 Hz, 1H), 5.58 (td, J = 6.8, 15.3 Hz, 1H), 4.49 (d, J_{AB} = 11.9 Hz, 1H), 4.45 (d, J_{AB} = 11.9 Hz, 1H), 4.04 – 3.96 (m, 1H), 3.45 – 3.37 (m, 2H), 3.27 (dd, J = 6.7, 9.8 Hz, 1H), 2.03 (dq, J = 1.2, 7.0 Hz, 2H), 1.38 (qd, J = 7.4, 14.7 Hz, 2H), 0.88 (t, J = 7.4 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ = 141.9, 138.0, 133.9, 129.5, 128.6, 128.4, 128.0, 127.7, 127.7, 126.6, 73.3, 73.3, 72.2, 52.3, 34.7, 22.5, 13.7. ¹H NMR (400MHz, C₆D₆) δ = 7.22 – 7.12 (m, 8H), 7.11 – 7.02 (m, 2H), 5.92 (tdd, J = 1.4, 8.4, 15.4 Hz, 1H), 5.46 (dtd, J = 1.2, 6.7, 15.7 Hz, 1H), 4.21 (d, J_{AB} = 11.7 Hz, 1H), 4.18 (d, J_{AB} = 11.7 Hz, 1H), 4.01 (dt, J = 3.5, 6.7 Hz, 1H), 3.46 (t, J = 7.6 Hz, 1H), 3.33 (dd, J_{AB} = 9.7 Hz, J_{AX} = 6.5 Hz, 1H), 3.24 (dd, J_{AB} = 9.7 Hz, J_{BX} = 3.7 Hz, 1H), 2.22 (br, s., 1H), 1.92 (dq, J = 1.2, 7.0 Hz, 2H), 1.27 (qd, J = 7.2, 14.8 Hz, 2H), 0.81 (t, J = 7.4 Hz, 3H). ¹³C NMR (101 MHz, C₆D₆) δ = 142.5, 138.5, 132.8, 130.2, 128.4, 128.3, 128.2, 126.3, 73.4, 72.9, 72.5, 52.2, 34.7, 22.4, 13.4. MS (EI, m/z) 310 (1>, M⁺), 160 (60), 159 (53), 117 (100), 104 (27), 91 (99). FTIR ($\nu_{\max}/\text{cm}^{-1}$): 3453, 2924, 2857, 1453, 1364, 1260, 1099, 734, 698.

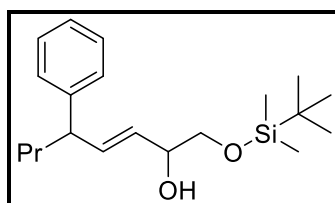


Figure 3.64. (*E*)-1-((*tert*-Butyldimethylsilyl)oxy)-5-phenyloct-3-en-2-ol

3da: ¹H NMR (400 MHz, CDCl₃) δ = 7.33 – 7.27 (m, 2H), 7.23 – 7.15 (m, 3H), 5.91 (dd, J = 7.8, 15.7 Hz, 1H), 5.42 (dd, J = 6.5, 15.5 Hz, 1H), 4.18 – 4.09 (m, 1H), 3.60

(dd, $J_{AB} = 9.9$ Hz, $J_{AX} = 7.6$ Hz, 1H), 3.41 (dd, $J_{AB} = 9.9$ Hz, $J_{BX} = 3.7$ Hz, 1H), 3.26 (q, $J = 7.6$ Hz, 1H), 1.69 (dq, $J = 2.7, 6.7$ Hz, 2H), 1.39 – 1.14 (m, 2H), 0.90 (s, 6H), 0.89 (t, $J = 7.4$ Hz, 3H), 0.06 (s, 6H). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 144.5, 137.2, 128.4, 127.8, 127.6, 126.1, 72.7, 67.2, 48.4, 38.0, 25.9, 20.7, 18.3, 14.0, -5.3, -5.4$. ^1H NMR (400 MHz, C_6D_6) $\delta = 7.21 - 7.14$ (m, 2H), 7.13 – 7.02 (m, 3H), 5.96 (ddd, $J = 1.4, 7.8, 15.5$ Hz, 1H), 5.48 (ddd, $J = 1.2, 5.9, 15.3$ Hz, 1H), 4.19 – 4.11 (m, 1H), 3.47 (dd, $J_{AB} = 10.0$ Hz, $J_{AX} = 7.1$ Hz, 1H), 3.38 (dd, $J_{AB} = 10.0$ Hz, $J_{BX} = 3.8$ Hz, 1H), 3.18 (q, $J = 7.4$ Hz, 1H), 2.39 – 2.29 (bs., 1H), 1.62 (q, $J = 7.6$ Hz, 2H), 1.23 (sptd, $J = 7.0, 28.6$ Hz, 2H), 0.89 (s, 9H), 0.83 (t, $J = 7.4$ Hz, 3H), -0.04 (d, $J = 1.6$ Hz, 6H). ^{13}C NMR (101 MHz, C_6D_6) $\delta = 136.2, 128.7, 128.4, 127.6, 126.0, 72.5, 67.5, 48.6, 38.1, 25.7, 20.7, 18.1, 13.8, -5.7, -5.7$. MS (EI, m/z) 275 (5), 186 (11), 185 (50), 159 (27), 143 (78), 129 (64), 117 (41), 91 (90), 75 (100), 73 (82). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3453, 2926, 2855, 1463, 1108, 836, 777, 699.

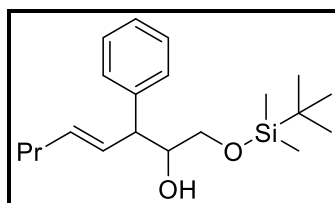


Figure 3.65. (*E*)-1-((*tert*-Butyldimethylsilyl)oxy)-3-phenyloct-4-en-2-ol

4da: ^1H NMR (400 MHz, CDCl_3) $\delta = 7.35 - 7.27$ (m, 2H), 7.25 – 7.16 (m, 3H), 5.79 (dd, $J = 8.6, 15.3$ Hz, 1H), 5.53 (td, $J = 6.7, 15.3$ Hz, 1H), 3.85 (spt, $J = 3.5$ Hz, 1H), 3.48 (dd, $J = 3.5, 10.2$ Hz, 1H), 3.34 (dd, $J = 7.0, 9.8$ Hz, 2H), 2.02 (q, $J = 7.3$ Hz, 2H), 1.38 (sxt, $J = 7.4$ Hz, 2H), 0.97 – 0.76 (m, 12H), -0.01 (s, 6H), ^{13}C NMR (101 MHz, CDCl_3) $\delta = 142.1, 133.2, 129.8, 128.5, 128.0, 126.5, 74.6, 65.0, 51.8, 34.8, 30.3, 25.9, 22.5, 18.2, 13.7, 1.0, -5.4, -5.4$. ^1H NMR (400 MHz, C_6D_6): $\delta = 7.21 - 7.27$ (m, 2H), 7.13 – 7.20 (m, 2H), 7.07 (d, $J = 7.4$ Hz, 1H), 5.97 (ddt, $J = 15.3, 8.3, 1.5$ Hz, 1H), 5.51 (dt, $J = 15.3, 6.8$ Hz, 1H), 3.86 – 3.93 (m, 1H), 3.55 (dd, $J_{AB} = 10.1$ Hz, $J_{AX} = 6.5$ Hz, 1H), 3.48 (t, $J = 7.6$ Hz, 1H), 3.44 (dd, $J_{AB} = 10.1$ Hz, $J_{BX} = 3.7$ Hz, 1H), 1.94 (q, $J = 6.7$ Hz, 4H), 1.28 (sxt, $J = 7.4$ Hz, 4H), 0.92 (s, 9H), 0.82 (t, $J = 7.4$ Hz, 3H), -0.02 ppm (d, $J = 1.6$ Hz, 6H). ^{13}C NMR (101 MHz, C_6D_6) $\delta = 142.6, 132.7, 130.4, 128.4, 128.3, 126.3, 74.6, 65.3, 51.9, 34.7, 25.7, 22.5, 18.1, 13.5, 1.0, -5.7, -5.7$. MS (EI, m/z) 277 (5), 117 (72), 105 (70), 103 (32), 75 (100), 73 (62). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3483, 2927, 2857, 1463, 1256, 1116, 835, 777, 700.

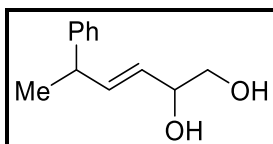


Figure 3.66. (*E*)-5-Phenylhex-3-ene-1,2-diol

3fa: ^1H NMR (400MHz, C_6D_6) δ = 7.21 - 7.14 (m, 2 H), 7.11 - 7.03 (m, 3 H), 5.79 (ddd, J = 1.6, 6.8, 15.5 Hz, 1 H), 5.31 (ddd, J = 1.4, 6.1, 15.7 Hz, 1 H), 3.95 - 3.87 (m, 1 H), 3.36 - 3.30 (m, 1 H), 3.29 - 3.17 (m, 2 H), 1.21 (d, J = 7.0 Hz, 4 H). ^{13}C NMR (101MHz, C_6D_6) δ = 145.9, 137.6, 128.9, 127.6, 126.6, 73.1, 66.9, 42.5, 21.4. MS (EI, m/z): 192 (M^+ , 1>), 174 (10), 161 (22), 143 (100), 128 (32), 115 (31), 105 (27), 91 (55), 77 (40), 55 (26), 43 (23).

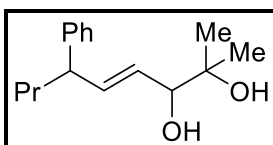


Figure 3.67. (*E*)-2-Methyl-6-phenylnon-4-ene-2,3-diol

3ga: ^1H NMR (400 MHz, C_6D_6) δ = 7.22 - 7.15 (m, 2 H), 7.13 - 7.03 (m, 3 H), 5.80 (ddd, J = 1.2, 7.8, 15.7 Hz, 1 H), 5.51 (ddd, J = 1.2, 7.0, 15.7 Hz, 1 H), 3.77 (dd, J = 1.0, 6.8 Hz, 1 H), 3.17 (q, J = 7.7 Hz, 1 H), 2.42 (br. s, 2 H), 1.61 (q, J = 7.6 Hz, 2 H), 1.32 - 1.16 (m, 2 H), 1.10 (d, J = 1.2 Hz, 6 H), 0.84 (t, J = 7.4 Hz, 3 H). ^{13}C NMR (101 MHz, C_6D_6) δ = 144.7, 137.0, 128.6, 128.4, 127.5, 126.0, 79.2, 72.5, 48.4, 37.9, 26.2, 23.5, 20.7, 13.8. MS (EI, m/z): 231 (1>), 214 (1>), 145 (2), 133 (6), 129 (6), 115 (19), 91 (20), 77 (2), 59 (100), 43 (17). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3401, 2928, 2871, 1453, 1164, 972, 699.

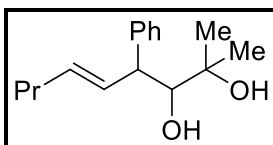


Figure 3.68. (*E*)-2-methyl-4-phenylnon-5-ene-2,3-diol

4ga: ^1H NMR (400MHz, CDCl_3) δ = 7.34 - 7.24 (m, 4 H), 7.24 - 7.17 (m, 1 H), 5.81 (dd, J = 9.4, 15.3 Hz, 1 H), 5.62 (td, J = 6.7, 15.6 Hz, 1 H), 3.68 (d, J = 5.5 Hz, 1 H),

3.49 (dd, $J = 5.9, 9.4$ Hz, 1 H), 2.30 (br. s., 1 H), 2.16 (br. s., 1 H), 2.03 (q, $J = 6.9$ Hz, 2 H), 1.39 (sxt, $J = 7.4$ Hz, 2 H), 1.22 - 1.19 (m, 3 H), 1.08 - 1.02 (m, 3 H), 0.88 (t, $J = 7.2$ Hz, 4 H). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 143.2, 134.5, 130.0, 128.7, 127.9, 126.5, 79.7, 51.7, 34.7, 29.7, 27.1, 25.3, 22.4, 13.7$. MS (EI, m/z): 248 (M^+ , 1>), 160 (36), 117 (41). 104 (22), 91 (21), 77 (7), 71 (10), 59 (100), 43 (37). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3422, 2926, 2858, 1453, 1378, 1160, 973, 700.

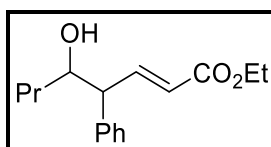


Figure 3.69. Ethyl-(*E*)-5-hydroxy-4-phenyloct-2-enoate

4ha: ^1H NMR (400 MHz, C_6D_6) $\delta = 7.55$ (ddd, $J = 1.4, 8.8, 15.7$ Hz, 1 H), 7.12 - 6.95 (m, 5 H), 5.96 (d, $J = 15.7$ Hz, 1 H), 4.02 (q, $J = 7.3$ Hz, 2 H), 3.57 (br. s., 1 H), 3.21 - 3.11 (m, 1 H), 1.48 - 1.30 (m, 2 H), 1.24 - 1.13 (m, 3 H), 0.96 (t, $J = 7.0$ Hz, 3 H), 0.73 (t, $J = 7.0$ Hz, 3 H). ^{13}C NMR (101MHz, C_6D_6) $\delta = 148.4, 140.7, 128.6, 128.2, 126.7, 123.3, 73.7, 59.8, 55.6, 37.1, 18.8, 13.9, 13.6$. FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3476, 2926, 2873, 1739, 1465, 1380, 1221, 1139, 701.

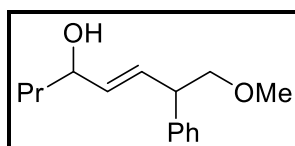


Figure 3.70.(*E*)-8-methoxy-7-phenyloct-5-en-4-ol

3ja: ^1H NMR (400 MHz, C_6D_6) $\delta = 7.22 - 7.12$ (m, 4 H), 7.11 - 7.02 (m, 1 H), 5.85 (ddd, $J = 1.2, 7.4, 15.7$ Hz, 1 H), 5.54 (ddd, $J = 1.2, 6.7, 15.7$ Hz, 1 H), 3.97 (q, $J = 5.5$ Hz, 2 H), 3.57 (q, $J = 6.3$ Hz, 2 H), 3.46 (dd, $J_{AB} = 9.2$ Hz, $J_{AX} = 7.5$ Hz, 1H), 3.42 (dd, $J_{AB} = 9.2$ Hz, $J_{BX} = 6.6$ Hz, 1H), 3.05 (s, 3 H), 1.53 - 1.26 (m, 4 H), 0.83 (t, $J = 7.2$ Hz, 3 H). ^{13}C NMR (101 MHz, CDCl_3) $\delta = 142.2, 135.4, 131.0, 128.4, 128.1, 126.5, 76.5, 72.2, 58.2, 48.5, 39.6, 18.8, 13.9$. MS (EI, m/z) 172 (6), 130 (31), 129 (16), 115 (10), 91 (17), 77 (6), 71 (44), 45 (100), 43 (57), 41 (34). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3422, 2928, 2971, 1453, 1111, 968, 699.

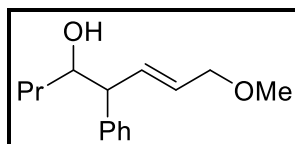


Figure 3.71. (*E*)-8-Methoxy-5-phenyloct-6-en-4-ol

4ja: ^1H NMR (400 MHz, CDCl_3) δ = 7.35 - 7.28 (m, 2 H), 7.25 - 7.17 (m, 3 H), 6.03 (ddd, J = 0.8, 9.0, 15.7 Hz, 1 H), 5.70 (td, J = 5.9, 15.3 Hz, 1 H), 3.93 (d, J = 5.9 Hz, 2 H), 3.81 (q, J = 7.0 Hz, 1 H), 3.36 - 3.24 (m, 1 H), 3.31 (s, 3 H), 1.54 - 1.47 (m, 1 H), 1.37 - 1.28 (m, 3 H), 0.85 (t, J = 6.8 Hz, 3 H). ^{13}C NMR (101 MHz, CDCl_3) δ = 141.8, 133.1, 129.7, 128.7, 128.0, 126.6, 74.0, 72.8, 57.9, 55.9, 36.7, 18.9, 14.0.

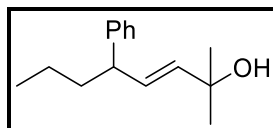


Figure 3.72. (*E*)-2-Methyl-5-phenyloct-3-en-2-ol

3ka: ^1H NMR (400 MHz, CDCl_3) δ = 7.36 - 7.25 (m, 2 H), 7.22 - 7.13 (m, 3 H), 5.75 (dd, J = 7.4, 15.3 Hz, 1 H), 5.61 (d, J = 15.7 Hz, 1 H), 3.22 (q, J = 7.7 Hz, 1 H), 1.66 (q, J = 7.6 Hz, 2 H), 1.35 - 1.14 (m, 2 H), 1.29 (d, J = 3.1 Hz, 6 H), 0.88 (t, J = 7.4 Hz, 3 H). ^{13}C NMR (101 MHz, CDCl_3) δ = 145.0, 137.5, 130.9, 128.4, 127.5, 126.0, 70.7, 48.1, 38.2, 29.9, 29.8, 20.7, 14.0. MS (EI, m/z) 218 (1 $^+$, M^+), 175 (4), 157 (6), 149 (12), 115 (10), 91 (14), 77 (7), 71 (8), 59 (19), 43 (100). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3368, 2959, 2927, 2871, 1453, 1149, 970, 910, 760, 698.

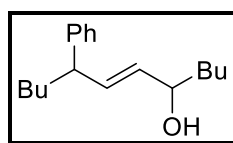


Figure 3.73(E)-8-phenyldodec-6-en-5-ol

3ma: ^1H NMR (400 MHz, C_6D_6) δ = 7.23 - 7.11 (m, 5 H), 7.10 - 7.04 (m, 1 H), 5.72 (dd, J = 7.6, 15.5 Hz, 1 H), 5.45 (dd, J = 6.5, 15.5 Hz, 1 H), 3.88 (q, J = 5.9 Hz, 1 H), 3.16 (q, J = 7.4 Hz, 1 H), 1.66 (q, J = 7.3 Hz, 2 H), 1.49 - 1.13 (m, 10 H), 0.87 - 0.81 (m, 6 H). ^{13}C NMR (101 MHz, C_6D_6) δ = 144.9, 134.2, 133.3, 128.4, 127.5, 126.0, 72.2, 48.6, 37.2, 35.6, 29.8, 27.6, 22.7, 22.6, 13.9, 13.8. MS (EI, m/z) 203 (2), 185 (3), 145 (6),

118 (18), 91 (42), 85 (40), 57 (62), 41 (100). FTIR ($\nu_{\text{max}}/\text{cm}^{-1}$): 3420, 2928, 2975, 1445, 1098, 967, 704.

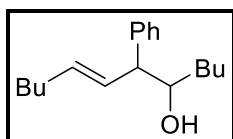


Figure 3.74(E)-6-phenyldodec-7-en-5-ol

4ma: ^1H NMR (400 MHz, CDCl_3) δ = 7.35 - 7.27 (m, 2 H), 7.24 - 7.16 (m, 3 H), 5.76 - 5.57 (m, 2 H), 3.78 - 3.67 (m, 1 H), 3.18 (t, J = 8.0 Hz, 1 H), 2.06 (q, J = 6.7 Hz, 2 H), 1.84 (br. s., 1 H), 1.49 - 1.17 (m, 10 H), 0.86 (td, J = 7.0, 15.7 Hz, 6 H). ^{13}C NMR (101MHz, CDCl_3) δ = 142.4, 134.7, 129.5, 128.6, 127.9, 126.4, 74.0, 56.4, 34.0, 32.3, 31.5, 27.9, 22.6, 22.2, 14.0, 13.9.

CHAPTER 4

RESULTS AND DISCUSSION

In this study, the Pd-catalyzed 1,3-substitution reactions of alkenyl oxiranes with organoborons were studied. For this purpose, the optimization conditions were thoroughly examined

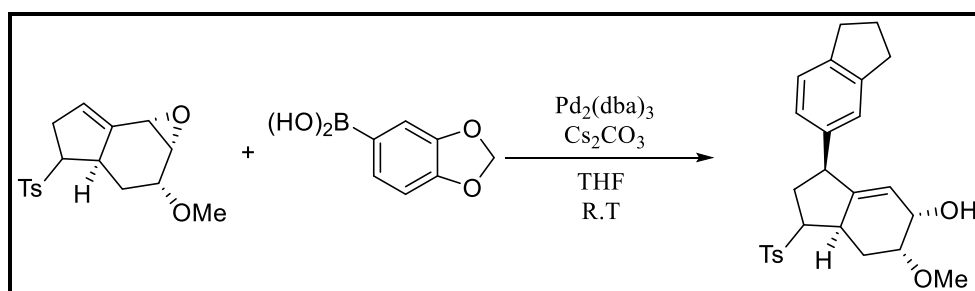


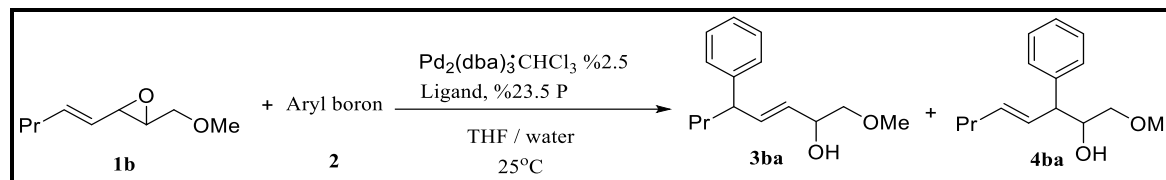
Figure 4.1. A method for the synthesis of montabuphine
(Source: Guan et al. 2012)

It was recently disclosed that as one of the synthesis step that was applied in synthesis of a biologically active compound, Pd-catalyzed coupling of an organoboronic acid with a cyclic vinyl epoxide proceeded stereo- and regio-selectively to give a 1,3-substituted major product (Figure 4.1). Hence, we have preferred to perform our first trial over the conditions reported therein for an acyclic vinyl epoxide compound **1b**, that is with phenyl boronic acid, using Pd₂(dba)₃·CHCl₃ as the Pd-complex, Cs₂CO₃ as a base, in dried THF, and 25 °C. The result was rather disappointing in fact, proceeded with poor regio- and diastereo-selectivity. (Table 4.1, No 1). However, it should be noted that the regio-isomers could be isolated from each other by silica gel column chromatography. In the same conditions, the usage of NaBPh₄ as a boron source decreased the regio-selectivity further (Table 4.1, No 2). While the reaction with phenyl boroxine (PhBO)₃ resulted in low yields, with KPhBF₃ the reactant was recovered as unreacted (Table 4.1, No 3 and 4). The use of phenyl boronic acid neopentyl glycol ester (Phneop) improved the regio-selectivity of the products in favor of 1,3-substitution product, however, diastereo-selectivity was significantly diminished. (Table 4.1, No 5). No significant improvement for the selectivity was observed with the use of an organic base ((*i*-Pr)₂NEt)

(Table 4.1, No 6). It was intriguing to observe however that the existence of water in the reaction mixture drastically improved the stereoselectivity and also caused the shift of this selectivity in favor of other diastereomeric structure. However, this improvement was accompanied by a decrease in regio-isomeric ratio (Table 4.1, No 7). With phenylboronic acid, the reaction gave nearly a similar result (Table 4.1, No 8).

The presence of a bidentate phosphine ligand DPEPhos could not further improve the process (Table 4.1, No 9-13).

Table 4.1. The Effect of conditions on reaction of vinyl epoxide (**1b**) with phenylboron compounds^a



No	Ligand	Aryl Boron (eqv.)	THF/H ₂ O (mL)	Base (eqv.)	t, hour(s)	% 3b (dr) ^b	% 4b (dr) ^b
1	-	PhB(OH) ₂ (2)	2.5:0	CS ₂ CO ₃ (2)	0.83	55 (1:4,7)	29 (7:1)
2	-	NaBPh ₄ (2)	2.5:0	CS ₂ CO ₃ (2)	0.83	55 (N.D.)	45 (N.D.)
3	-	(PhBO) ₃ (2)	2.5:0	CS ₂ CO ₃ (2)	O.N.	30 (N.D.)	(N.D.)
4	-	PhBF ₃ K (2)	2.5:0	CS ₂ CO ₃ (2)	O.N.	-	-
5	-	PhBneop (2)	2.5:0	CS ₂ CO ₃ (2)	21	82 (3:1)	13 (3:1)
6	-	PhBneop (2)	2.5:0	(<i>i</i> -Pr) ₂ NEt (4)	<1	65 (6,6:1)	16 (1,1:1)
7	-	PhBneop (3)	2.0:5	(<i>i</i> -Pr) ₂ NEt (4)	0.25	53 (1:20)	22 (35:1)
8	-	PhB(OH) ₂ (3)	2.0:5	(<i>i</i> -Pr) ₂ NEt (4)	0.33	53 (1:15)	31 (8:1)
9	DPEPhos	PhBneop (3)	2.0:5	(<i>i</i> -Pr) ₂ NEt (4)	8<>24	62 (1:18)	26 (34:1)
10	DPEPhos	PhB(OH) ₂ (3)	2.0:5	(<i>i</i> -Pr) ₂ NEt (4)	7.5	58 (1:29)	22 (23:1)
11	DPEPhos	PhBneop (3)	2.0:0	(<i>i</i> -Pr) ₂ NEt (4)	24	-	-
12	DPEPhos	PhBneop (3)	2:0.5	CS ₂ CO ₃ (2)	O.N.	56 (1:30)	21 (>40:1)
13	DPEPhos	PhBneop (3)	2:0.5	N(Et) ₃ (2)	7	61 (1:32)	20 (>40:1)

^aO.N.: Overnight. ^bDetermined by ¹H NMR using C₆D₆ solvent and N.D.: Not determined.

It was surprising for us to find that the vinyl oxirane **1a** with an unprotected pendant hydroxyl group afforded much better regio-selectivity in the presence of DPEphos ligand and organic base as compared to its unprotected form **1b**, producing **3a** and **4a** in 69% and 6% isolated yields, respectively. The compound containing oxygen group might have affect the activity of the metal as a result of coordination of free hydroxyl group with the palladium core.

In the next stage of the optimization studies, the ligand activity was surveyed. It should be noted that since the tedious column isolation work-up is highly time consuming, the ¹H NMR technique has been the usual preferred technique for the quantitative analysis of the crude products for fast screening of the optimum conditions. However, this technique allowed us only the determination of the **3aa** product accurately and the accurate analysis of the **4aa** product has not been possible since its related proton signals were overlapped with those of other reagents existing within the mixture

The variation of DPEPhos amount had no remarkable influence on the reaction selectivity (Table 4.2, No 2 and 3) and therefore, the ratio of P/Pd was kept nearly 4:1 for the next experiments.

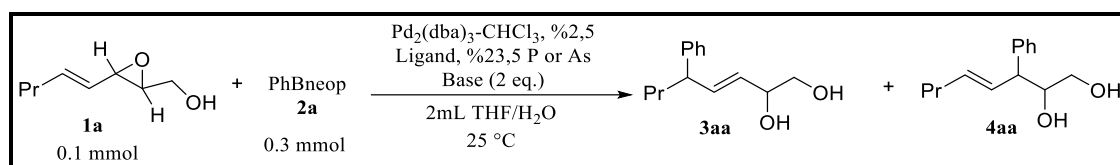
However, it should be noted that no further improvements have been possible by use of any of mono- or bidentate phosphine ligands tested for this study. The bidentate ligands xantphos, dppb, and significant number of electron rich and poor ligands resulted in intricate mixture. No conversion would be possible with the use of dppp and P(2-MeOC₆H₄)₃ ligands. On the other hand, among the other ligands tested only dppf ligand afforded a comparable result with respect to that which was obtained with DPEphos.

At last part of optimization of ligands, several non-phosphorous ligands were examined. The product **3a** formation was lowered significantly with the use of 2,2'-bipyridyl, whereas relatively moderate product formation has been possible with the use of an N-heterocyclic carbene precursor (1,3-bis(2,6-di-*i*-propylphenyl)imidazol-2-ylidene) (Table 4.2, No 25 and 26) On the other hand it has been delightful to find that the best selectivity could be obtained with the presence of AsPh₃ within the reaction medium, which produced **3aa** and **4aa** in 78% and 6% isolated yields, respectively in the presence of NEt₃ base (Table 4.2, No 27). A comparative result was obtained with (*i*-Pr)₂NH base (No 33), with its use **3aa** and **4aa** formations being 79% and 10%, respectively.

It is apparent that the method is quite component to the number of other organic and inorganic bases. The variation of the base type had modest influence on the activity

of the system. (Table 4.2, No 28-34). However, a relatively lower yield was obtained without presence of any base and also without any ligands the reaction was obtained at 30% isolated yield. (Table 4.2, No 35 and No 36).

Table 4.2. Effect of the ligand on pd-catalyzed reaction of alkenyl epoxide (**1a**) with PhBneop



No	Ligand	Base	t, hour(s)	% 3a ^a (dr)	% 4a
1	DPEPhos	N(Et) ₃	3	77-69 ^b (30:1)	6 ^b
2	DPEPhos ^c	N(Et) ₃	1	66	N.D.
3	DPEPhos ^d	N(Et) ₃	1	71	N.D.
4	Xantphos	N(Et) ₃	3	Complex	
5	<i>t</i> -Bu-Xantphos	N(Et) ₃	1	63	N.D.
6	dppe	N(Et) ₃	5 days	44	N.D.
7	dppp	N(Et) ₃	20	-	-
8	dppb	N(Et) ₃	1	Complex	
9	dppf	N(Et) ₃	2	77	N.D.
10	±BINAP	N(Et) ₃	20	22	N.D.
11	PPh ₃	N(Et) ₃	1.5	Complex	
12	Pd(PPh ₃) ₄	N(Et) ₃	2.5	Complex	
13	P(4-MeOC ₆ H ₄) ₃	N(Et) ₃	2.5	Complex	
14	P(2-MeOC ₆ H ₄) ₃	N(Et) ₃	20	-	-
15	P(4-CF ₃)C ₆ H ₄	N(Et) ₃	2.5	Complex	
16	Ph(2-furyl) ₃	N(Et) ₃	1.5	Complex	
17	[(<i>t</i> -Bu) ₃ PH]BF ₄	N(Et) ₃	4.5	56	N.D.
18	[Cy ₃ PH]BF ₄	N(Et) ₃	8	69	N.D.
19	P(Bu) ₃	N(Et) ₃	1.5	44	N.D.

(cont. on next page)

Table 4.2 (cont.)

19	P(Bu) ₃	N(Et) ₃	1.5	44	N.D.
20	P(Ph ₂ Bn) ₃	N(Et) ₃	20	49	N.D.
21	P(Ph ₂ Me) ₃	N(Et) ₃	20	32	N.D.
22	Xphos	N(Et) ₃	3 days	57	N.D.
23	P(OBu) ₃	N(Et) ₃	1.75	62	N.D.
24	P(OPh) ₃	N(Et) ₃	3 days	54	N.D.
25	2,2'-Bipyridyl	N(Et) ₃	5 days	38	N.D.
26	SIPr	N(Et) ₃	1	68	N.D.
27	Ph₃As	N(Et)₃	2.5	84-78^b	6^b
28	Ph ₃ As	K ₃ PO ₄	3	80	N.D.
29	Ph ₃ As	Cs ₂ CO ₃	2.75	83	N.D.
30	Ph ₃ As	K ₂ CO ₃	3.5	78	N.D.
31	Ph ₃ As	KF	3.5	80	N.D.
32	Ph ₃ As	KOH	1.5	82	N.D.
33	Ph₃As	(<i>i</i>-Pr)₂NH	2	84-79^b	10^b
34	Ph ₃ As	(<i>i</i> -Pr) ₂ NEt	4	82	N.D.
35	Ph ₃ As	-	3	72	N.D.
36	-	(<i>i</i> -Pr) ₂ NEt	1.5	33-30 ^b	N.D.

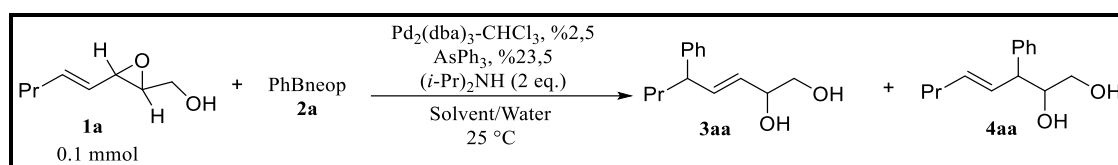
^aDetermined by internal standard using *p*-anisaldehyde by ¹H-NMR. ^bIsolated yield ^c% 10 P. ^d% 15 P

Reducing the reaction concentration to the half or doubling it caused a decrease of the reaction yield (Table 4.3, No 1 and 2). While the ratio of water in the reaction mixture directly affected the reaction time, the NMR yield of these reactions were not affected significantly (Table 4.3, No 3 and 4). According to all results obtained so far, for 0.1 mmol reactant, the available solvent system was set to be 2 mL THF/0.5 mL H₂O.

The various organic solvents were tested (Table 4.3, No 5-12) and among the solvents tried, only was acetone that led to the formation of a comparable yield, albeit the reaction rate was highly diminished and took 2 days for a complete conversion (Table 4.3, No 9). The lower yields were obtained with the use of other solvents, such as DMF, isopropanol, DCM, dioxane, DME, toluene, and acetonitrile.

At 50°C, the reaction rate was relatively accelerated, but the NMR yield was slightly lowered to 78% (Table 4.3, No 13). On the other hand, at 10°C the reaction rate was reasonably decelerated though the reaction yield did not change (Table 4.3, No 14). It is obvious that doubling the reaction scale was tolerated by the method (Table 4.3, No 15). The less usage of PhBneop than 3 eqv. caused a decrease of the yield (Table 4.3, No 16 and 17).

Table 4.3. Optimization of solvent system of pd-catalyzed reaction with PhBneop



No	Solvent	Solvent/H ₂ O (mL)	PhBneop (eqv.)	T °C	t, hour(s)	%3aa ^a	%4aa
1	THF	4/1	3	25	5.5	66	N.D. ^f
2	THF	1/0.25	3	25	2.5	58	N.D.
3	THF	2/0.25	3	25	5	80	N.D.
4	THF	2/1	3	25	2.25	82	N.D.
5	DCM	2/0.5	3	25	5.2	36	N.D.
6	Toluene	2/0.5	3	25	5.2	50	N.D.
7	DMF	2/0.5	3	25	O.N.	65	N.D.
8	Isopropanol	2/0.4	3	25	O.N.	74	N.D.
9 ^c	Acetone	2/0.4	3	25	2 days	82	N.D.
10 ^c	1,4-Dioxane	2/0.5	3	25	5.5	73	N.D.
11	Acetonitrile	2/0.5	3	25	O.N.	48 ^e	N.D.
12	DME ^g	2/0.5	3	25	O.N.	76	N.D.
13	THF	2/0.5	3	50	<1	78	N.D.
14	THF	2/0.5	3	10	O.N.	83	N.D.
15 ^d	THF	4/1	3	25	3	83-76 ^b	12 ^b
16 ^d	THF	4/1	2	25	3.5	70	N.D.
17 ^d	THF	4/1	1.5	25	5	51	N.D.

^aDetermined by ¹H-NMR using *p*-anisaldehyde as the internal standard. ^bIsolated yield. ^cNot dried solvents.

^d0.2 mmol reactant. ^eReactant was not consumed completely. ^fN. D.: Not determined. ^gThe base is NEt₃.

An another type of organoboron ester, the pinacol ester of phenylboronic acid was also reacted with alkenyl oxirane **1a**. The reaction kinetic was slowed by this change and the desired product was obtained with lower yield as 71% (Figure 2.2).

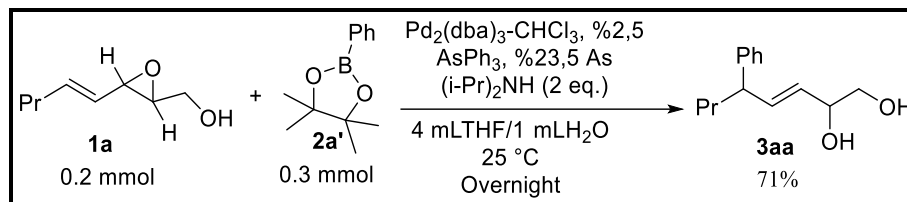
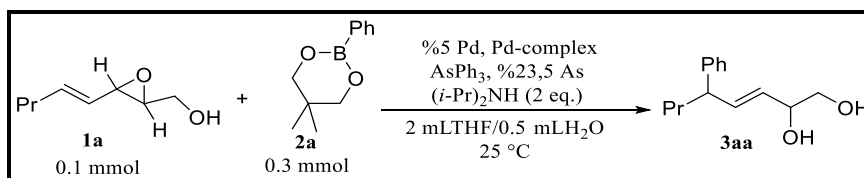


Figure 4.2. The reaction of alkenyl oxirane with pinacol ester of phenylboronic acid

In the arylation reactions, catalytic effects of some of pd complexes were also examined and it was observed that palladium acetate and π -allyl palladium chloride dimer which are Pd(II)-complexes were not effective as compared to Pd (0)-complexes. These catalysts slowed the reaction for a full conversion of the reactant and the lower yields were gained (Table 4.4, No 1 and 2). Non-solvent coordinated $\text{Pd}_2(\text{dba})_3$ complex showed a lower catalytic activity than $\text{Pd}_2(\text{dba})_3 \cdot \text{CHCl}_3$ (Table 4.4, No 3 and Table 4.2, No 27).

Table 4.4. Arylation reactions of the vinyl epoxide **1a** with various Pd-catalysts



No	Pd	T, hour(s)	% 3aa ^a
1	$\text{Pd}(\text{OAc})_2$	2 days	66
2	$[\text{PdCl}(\text{C}_3\text{H}_5)]_2$	2 days	51
3	$\text{Pd}_2(\text{dba})_3$	O. N.	77

^aDetermined by $^1\text{H-NMR}$ using *p*-anisaldehyde as the internal standard.

The table 4.5 includes the results of arylation reaction of the vinyl epoxide **1a** with various RBneop reagents. The method was effectively applicable to electron-rich arylborons (Table 4.5, No 1-3 and 7-8) and 2-naphthylboron reagent (Table 4.5, No 5 and 6). The desired product **3a** was formed with these organoborons at high yields and diastereomeric ratios. Although the *ortho*-substituted methyl, methoxy and 1-naphthyl organoborons substrates which are relatively sterically hindered reagents (Table 4.5, No

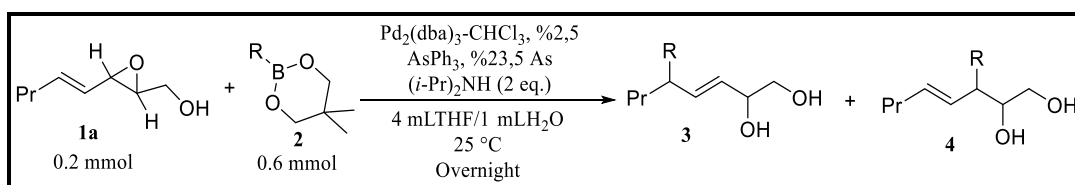
2,6 and 8) slowed down the reaction rate, their performance from the point of allylic substitution yield was more or less comparable to other isomers.

It was noticed that the organoborons having electron-withdrawing groups showed a weak performance in the reaction. Longer reaction periods are required for complete conversion of the vinyl epoxide with these reagents and moderate yields were recovered with *p*- and *m*- substituted ones (Table 4.5, No 9, 10 and 12). A low yield was obtained with a *o*-substituted electron poor organoboron reagent **2m** (Table 4.5), No 11). This reactivity difference might be resulted from in the less reactivity of the electron poor organoborons toward transmetallation step with the intermediate of π -allyl palladium complex.

The reaction with an alkenyl boron derivative containing a phenyl group resulted in the formation of product at 80% yield and with relatively lower 10:1 diastereomeric ratio (Table 4.5, No 13). On the other hand, alkenyl boron compound containing propyl group showed lower activity and the product **3ap** was formed at 30% yield (Table 4.5, No 14).

During a reaction with thiophenylboron ester **2r** palladium black formation was observed in first hour of the reaction and as a result the vinyl epoxide was recovered as it is. (Table 4.5, No 15).

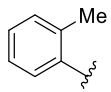
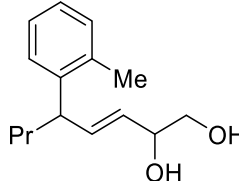
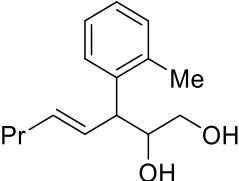
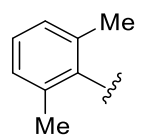
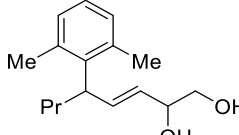
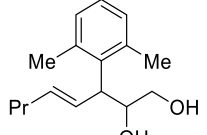
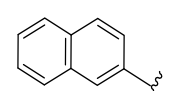
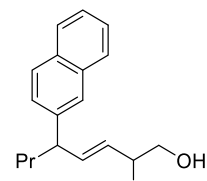
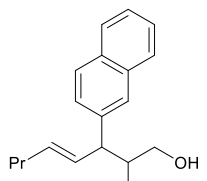
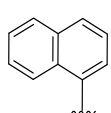
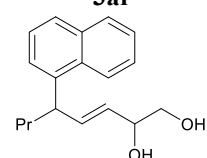
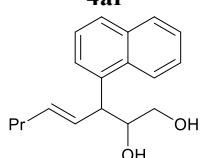
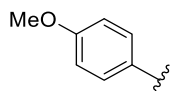
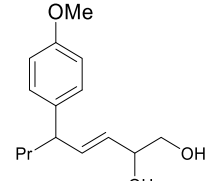
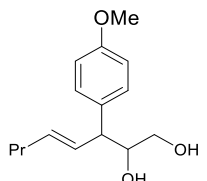
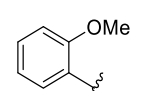
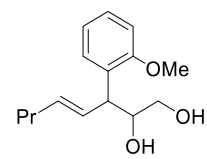
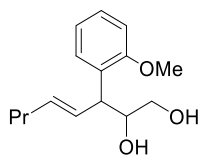
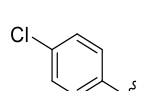
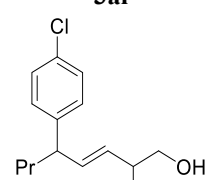
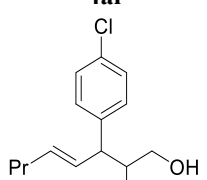
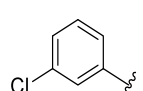
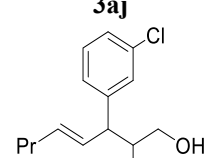
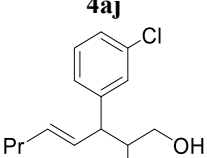
Table 4.5. Reaction of vinyl epoxide **1a** with various neopentyl glycol esters



No	Boron ester, 2	Time, h	Product, 3	3 % (dr)	Product, 4	4 %
1		1.75		76 (>20)		6
2		1.67		75 (>20)		2

(cont. on next page)

Table 4.5 (cont.)

3		O. N.		79 (9:1)		-
	2d		3ad		4ad	
4		O.N.		12 ^x (>20)		-
	2e		3ae		4ae	
5		3.75		74 (>20)		10
	2f		3af		4af	
6		O. N.		75 (>20)		7
	2g		3ag		4ag	
7		1.5		80 (>20)		8
	2h		3ah		4ah	
8		O. N.		78 (>20)		4 ^x
	2i		3ai		4ai	
9		O. N.		56 (>20)		2
	2j		3aj		4aj	
10		O. N.		53 (>20)		-
	2k		3ak		4ak	

(cont. on next page)

Table 4.5 (cont.)

11		4 days		23 (>20)		-
12		O. N.		51 (>20)		5
13		2		80 (10:1)		7
14		O. N.		30 (>20)		<1
15		O. N.		-		-

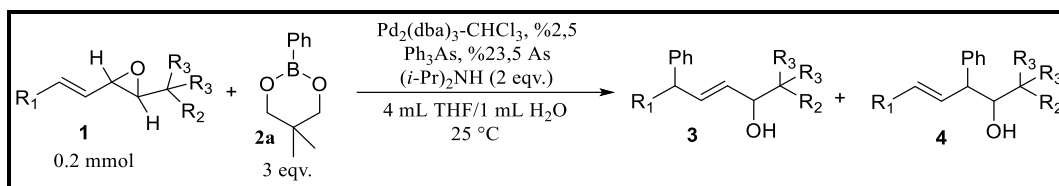
O. N.: Overnight. N. D.: Not determined. Dr: Diastereomeric ratio.

Next, the substrate scope of the method was also surveyed over various alkenyl oxiranes. The use of vinyl epoxide with phenyl group attached to the far alkenyl carbon as the reactant in the reaction resulted in a complex mixture. Although the desired product **3ea** compound could not be isolated, the simple substitution product **4ea** was determined to be 30% yield by NMR technique (Table 4.6, No 1). The reaction with DPEphos ligand of **1e** also did not alter the reaction fate at all (Table 4.6, No 2).

When the hydroxyl group was protected by methyl (**1c**) and silyl (**1d**) groups, the ratio of regio-selectivities of reactions were decreased (Table 4.6, No 3 and 4). It was possible that the coordination of the oxygen containing group which was next to the oxirane ring with the Pd-metal somehow influences the regio-selectivity on the reaction. The reaction's efficiency was not affected with by the presence of a methyl group at R₁ and at R₃. Therefore, the yields of these reactions were obtained at high level (Table 4.6, No 5 and 6).

The reactant having an ester group at R₁ led to only the formation of the simple substitution product **4ha** exclusively (Table 4.6, No 7). While the presence of a hydroxyl group within R1 lowered the regio-selectivity of the process the methyl protection of this group even further worsened the selectivity and lower total yields were obtained (Table 4.6, No 8 and 9).

Table 4.6. The Pd-catalyzed reaction of various alkenyl epoxides with PhBneop



No	t, h	Product 3 (yield%)	Product 4 (yield%)
1	O.N.	 3ea (N.D.)	 4ea (30%) ^a
2 ^b	O.N.	 3ea (3%)	 4ea (12%) ^a
3	7	 3ca (66%)	 4ca (25%)
4	5.3	 3da (65%)	 4da (18%)
5	1.5	 3fa (84%) ^a	 4fa (-)
6	4.7	 3ga (79%)	 4ga (10%)
7	2 days	 3ha (-)	 4ha (51%)
8	6	 3ia (51%) ^a	 4ia (25%) ^a
9	10	 3ja (22%)	 4ja (10%)

^aDetermined by ¹H-NMR using *p*-anisaldehyde as the internal standard. ^bThe reaction was performed using DPEPhos ligand (22.5 P).

In Figure 4.3, with the oxirane having fully alkyl groups, the reaction was resulted in 65% isolated yield after overnight reaction period. And in figure 4.5, the reaction of mono alkyl substituted oxiranes gave 44% isolated yield for **3ma** and 16% isolated yield for **4ma** product.

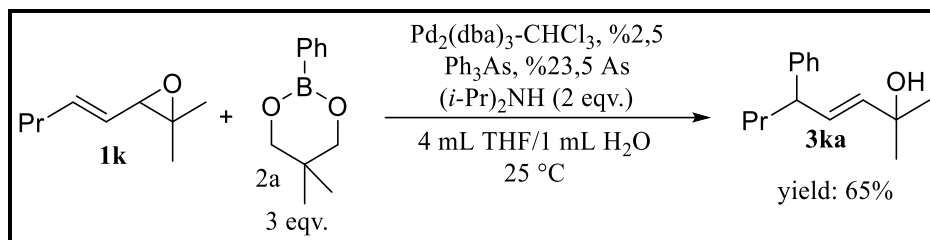


Figure 4.3. Synthesis of (*E*)-2-methyl-5-phenyloct-3-en-2-ol

The reaction also was tried on a cyclic structure. Conversely, the reaction with 3,4-epoxycyclohexene was complete in 1 hour, but a complex mixture was obtained instead of the desired arylated product.

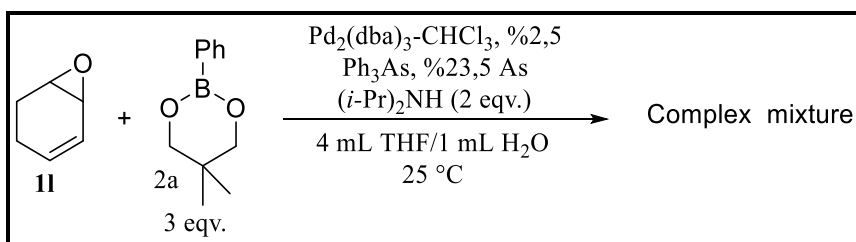


Figure 4.4. The reaction of 3,4-epoxy cyclohexene

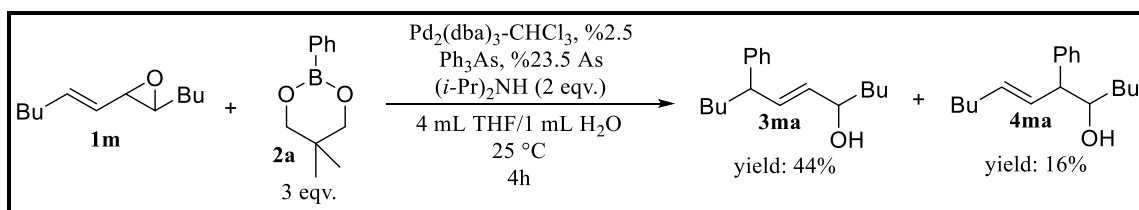


Figure 4.5 The reaction of (*E*)-2-butyl-3-(hex-1-en-1-yl)oxirane

CHAPTER 5

CONCLUSION

As a consequence, a new 1,3-substitution selective arylation reaction of vinyl oxiranes was performed with aryl or alkenyl neopentyl glycol boron esters in the presence of triphenyl arsine ligated tris(dibenzylideneacetone)dipalladium (0) chloroform adduct. The products were obtained with highly diastereomeric ratios

The advantages of reaction are the applicability at room temperature, the use of various stable boron esters in aqueous medium.

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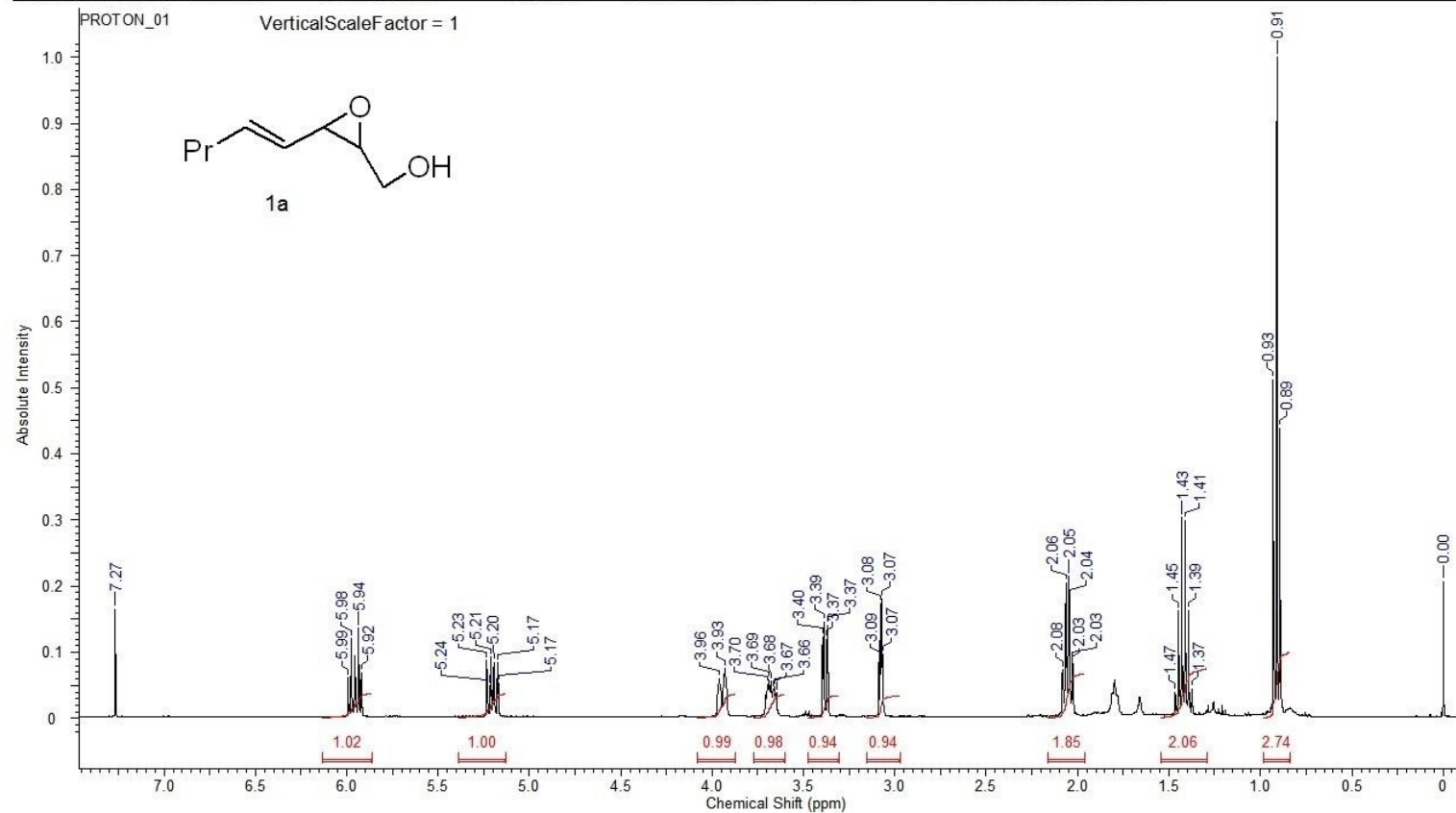
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APPENDIX A

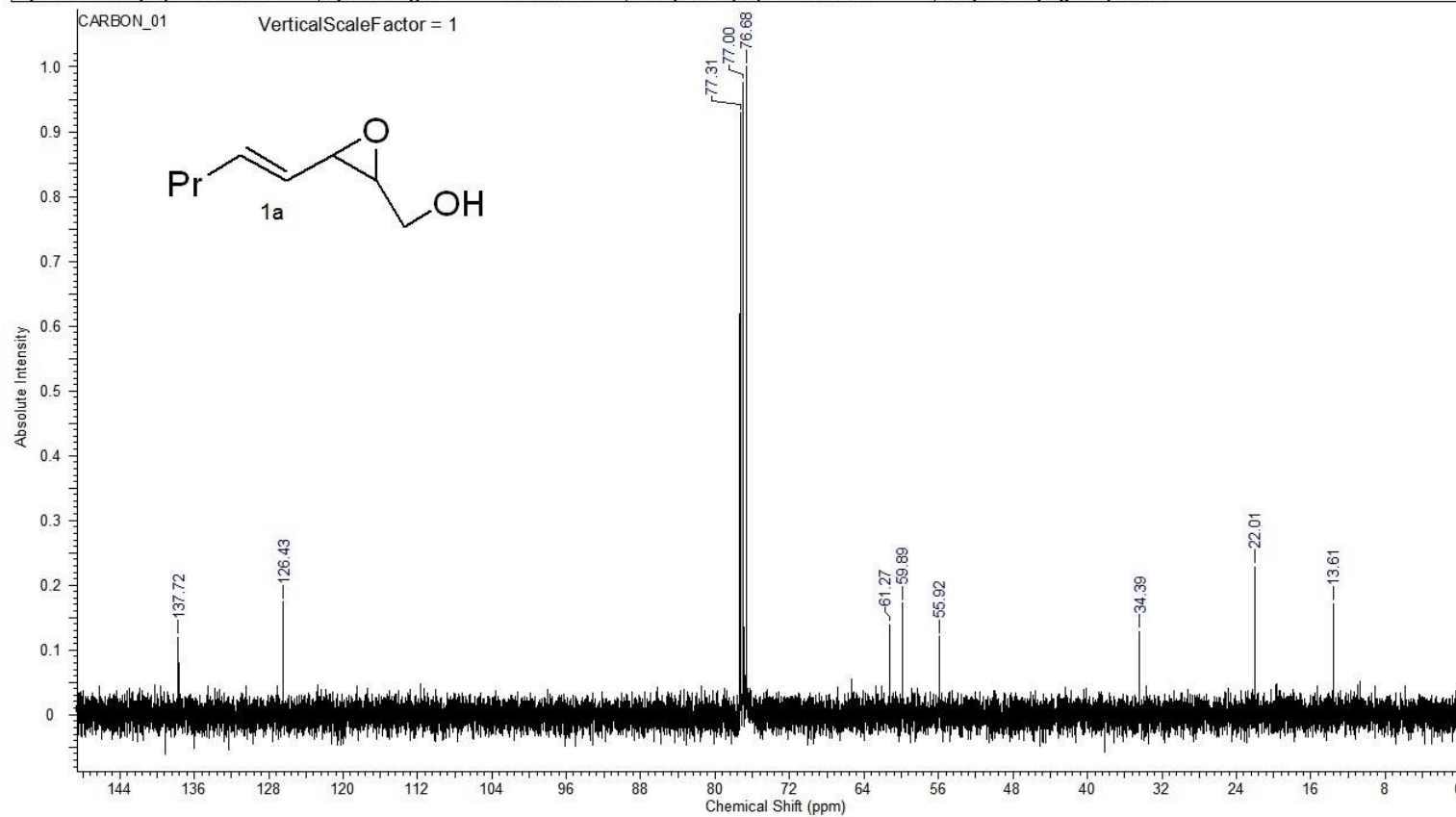
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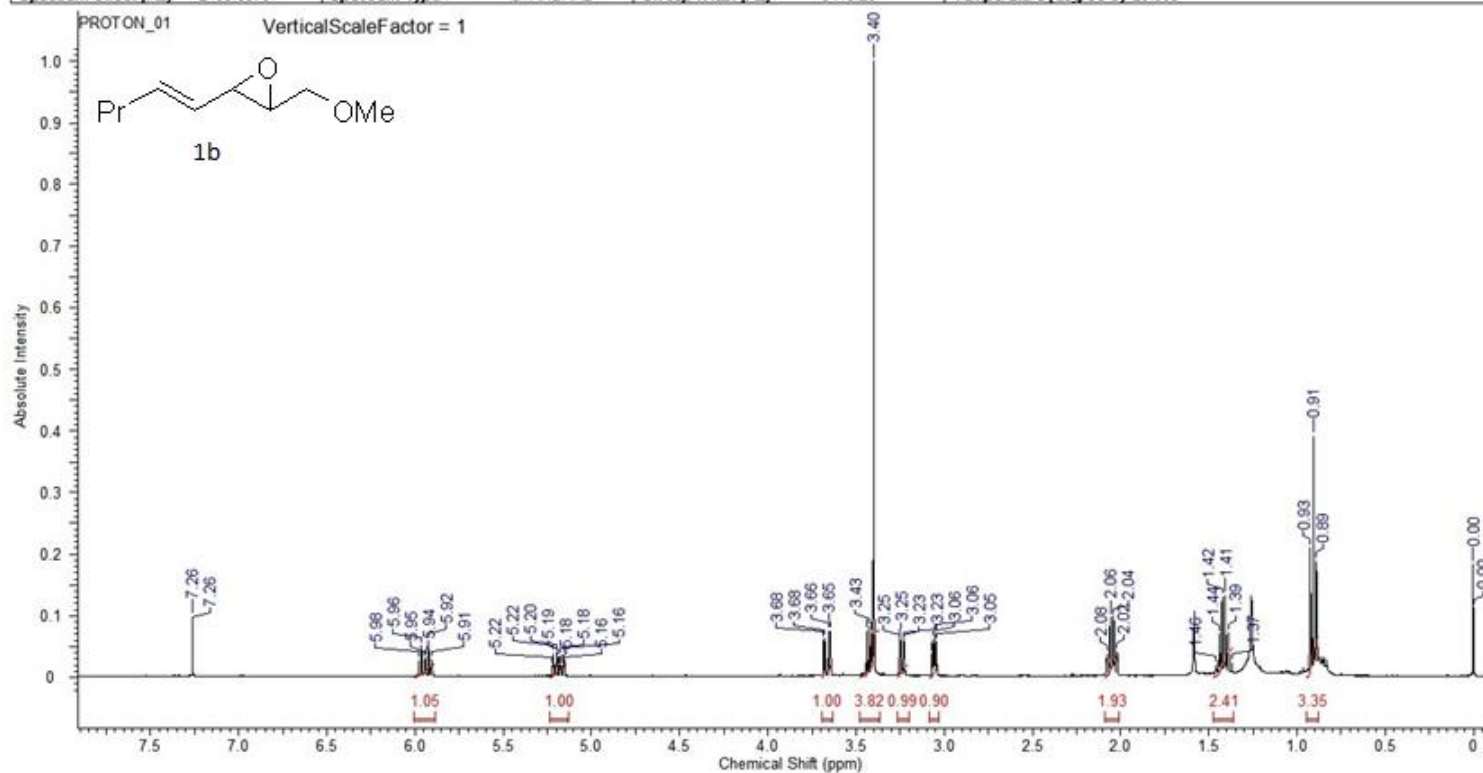
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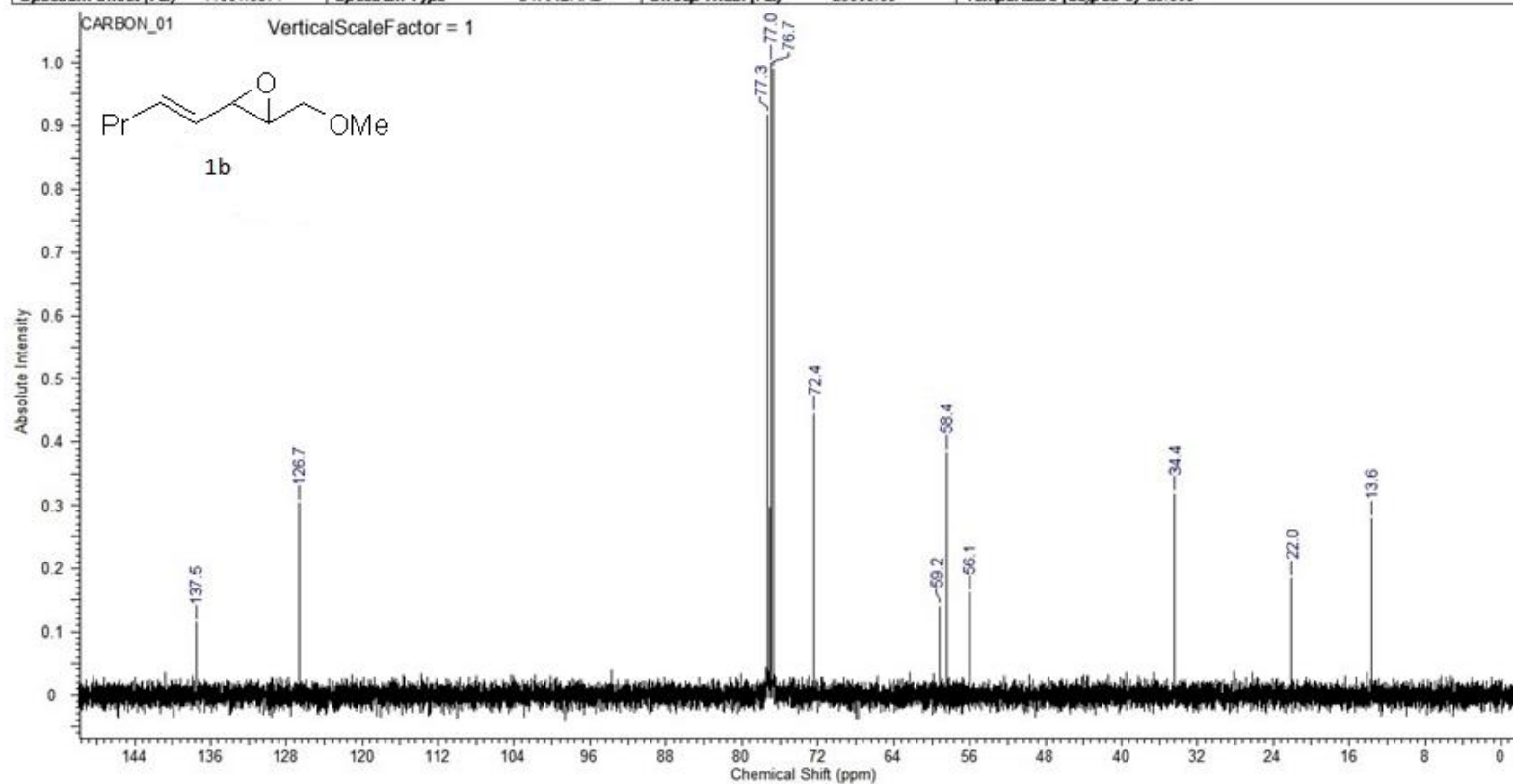
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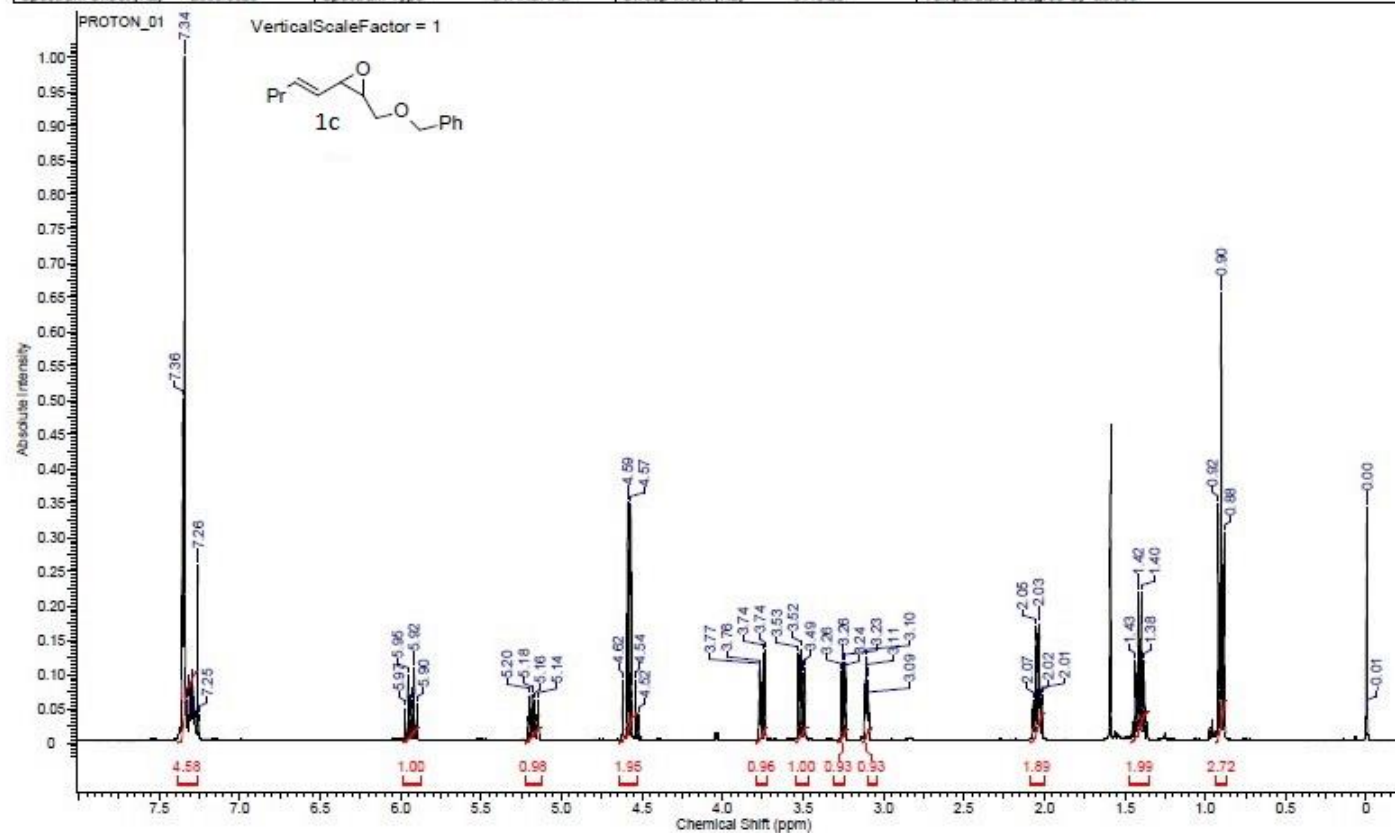
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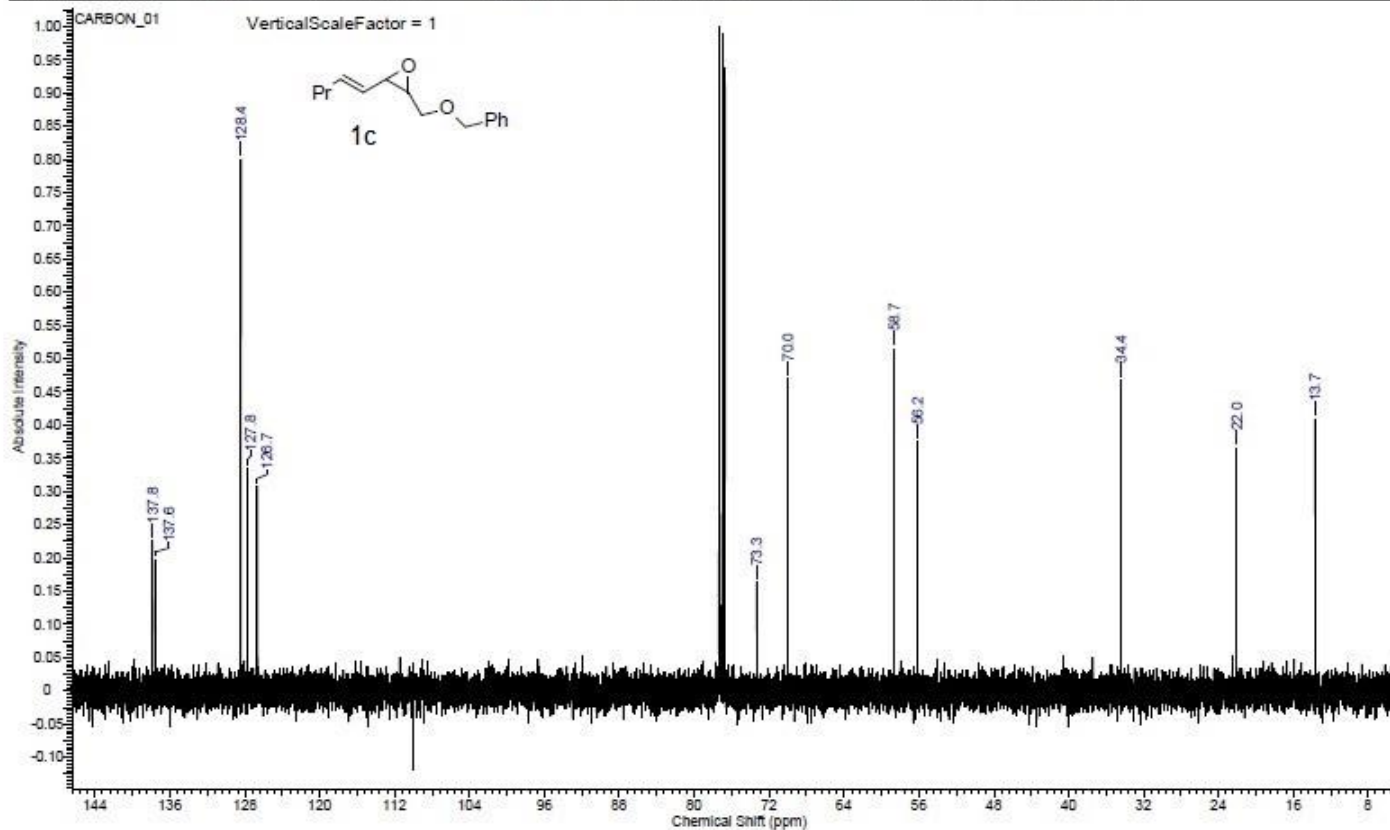


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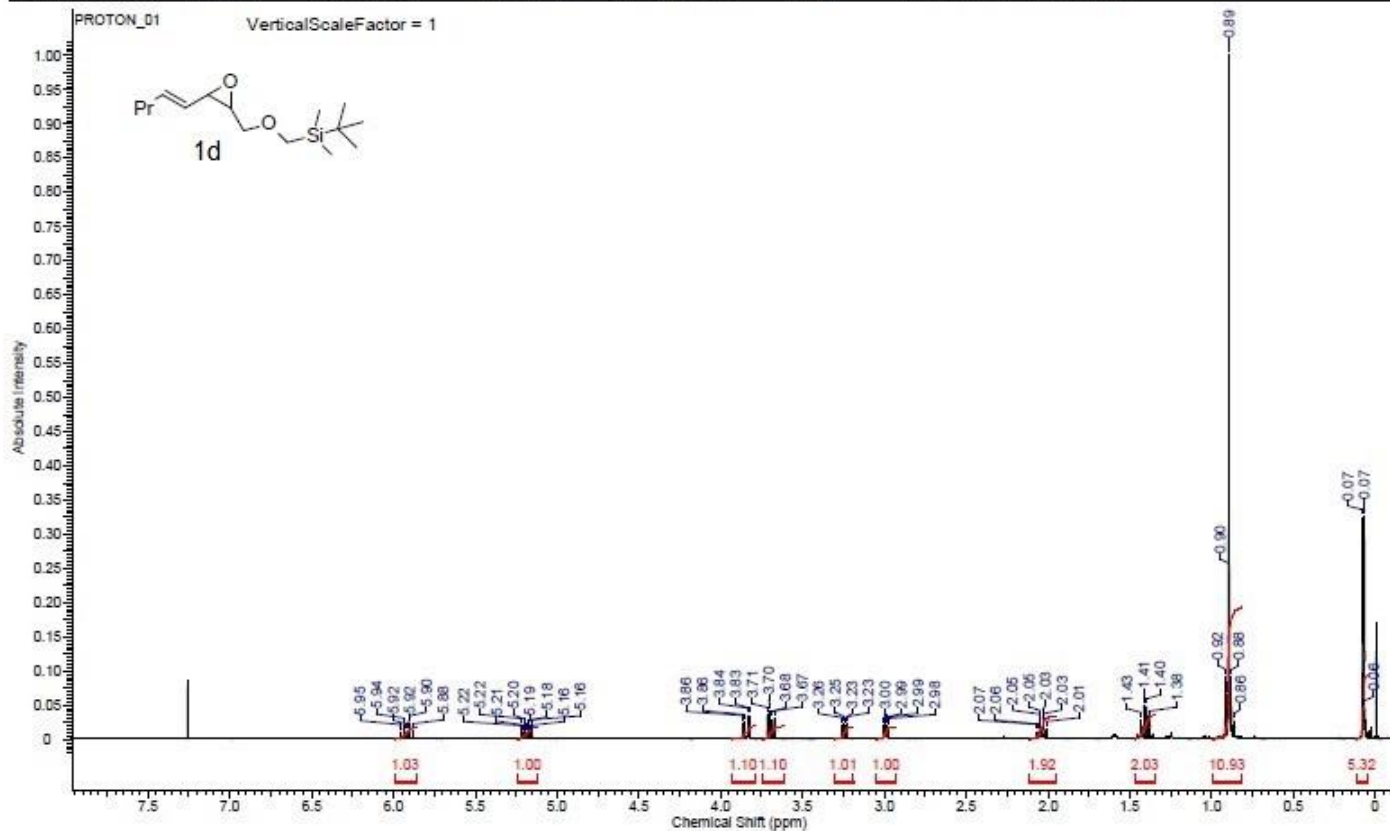
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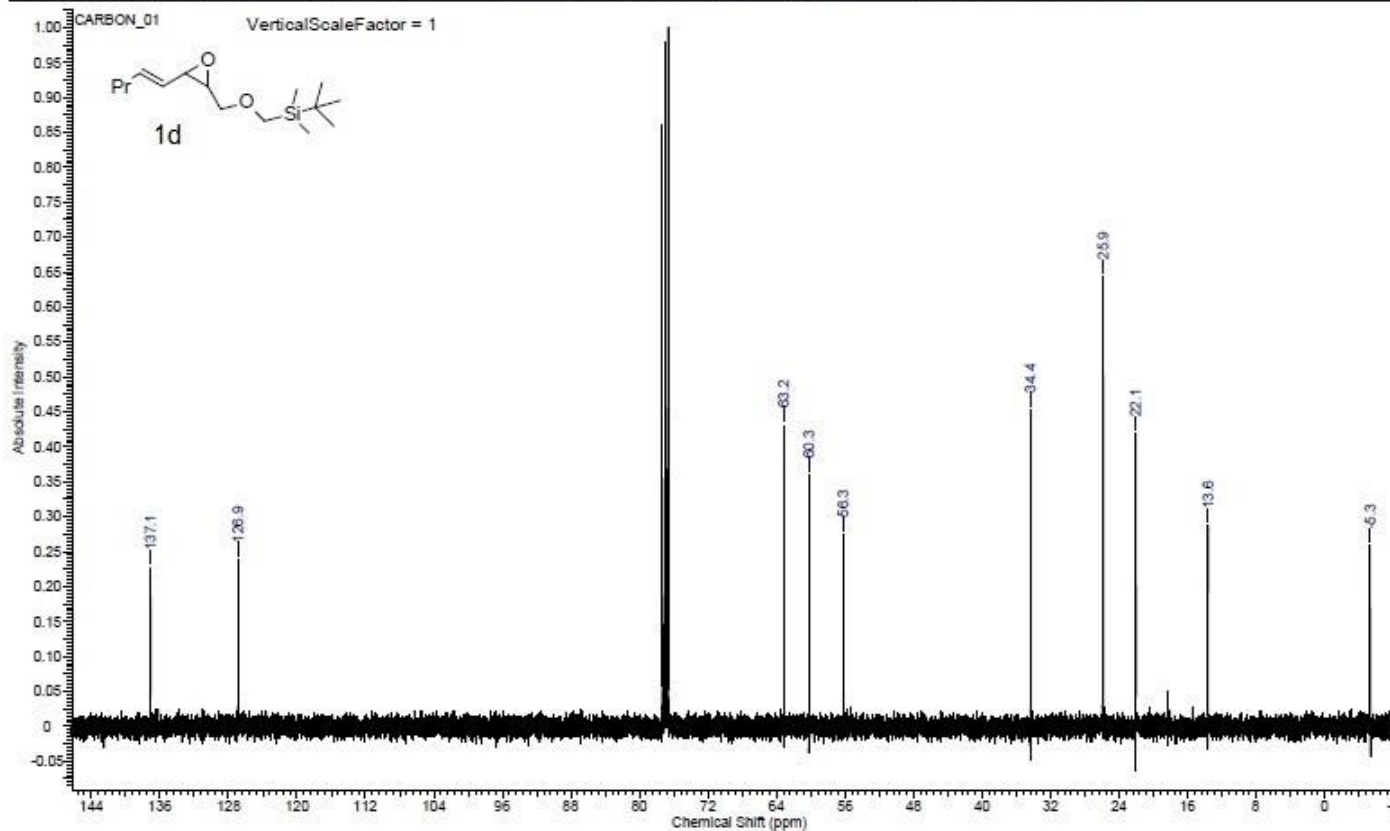
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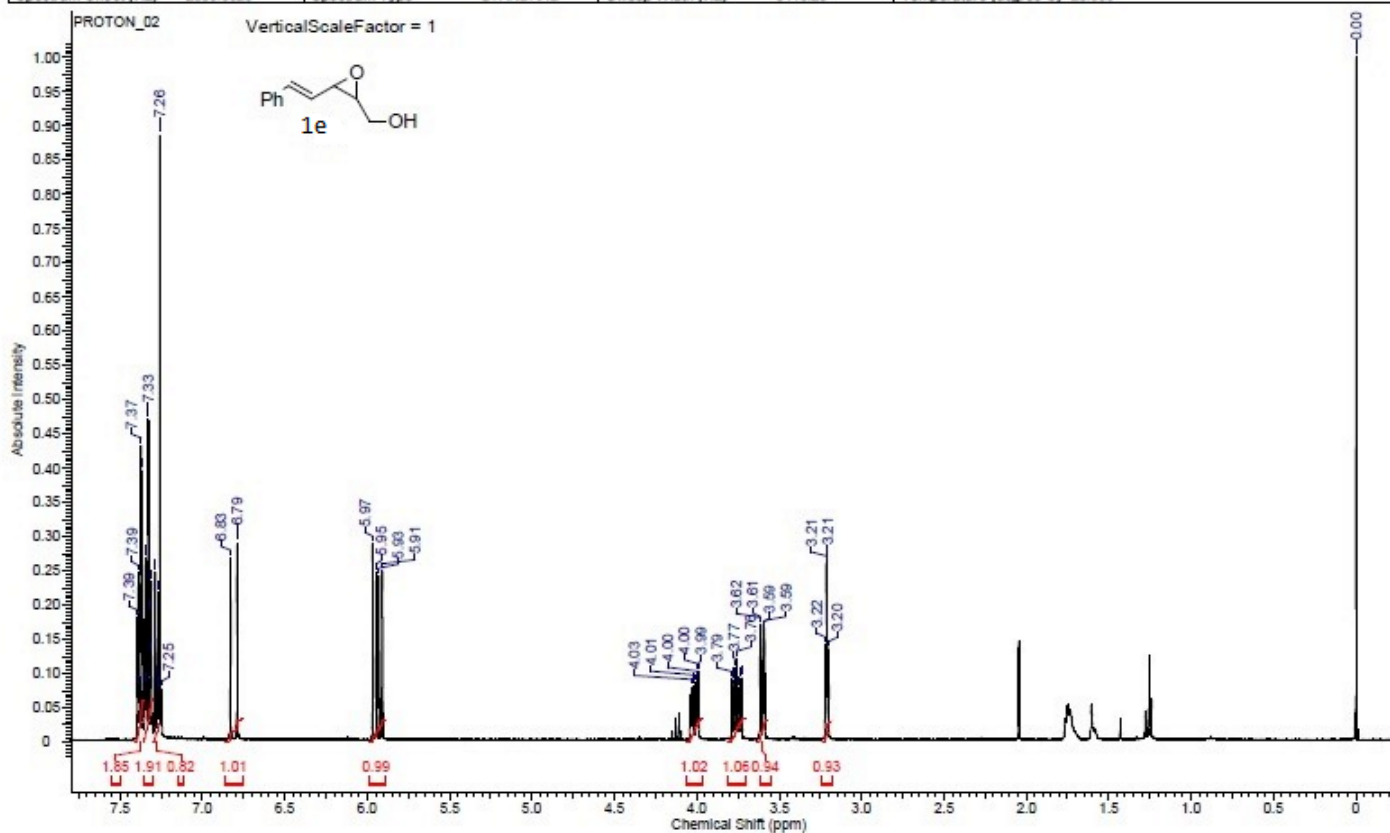
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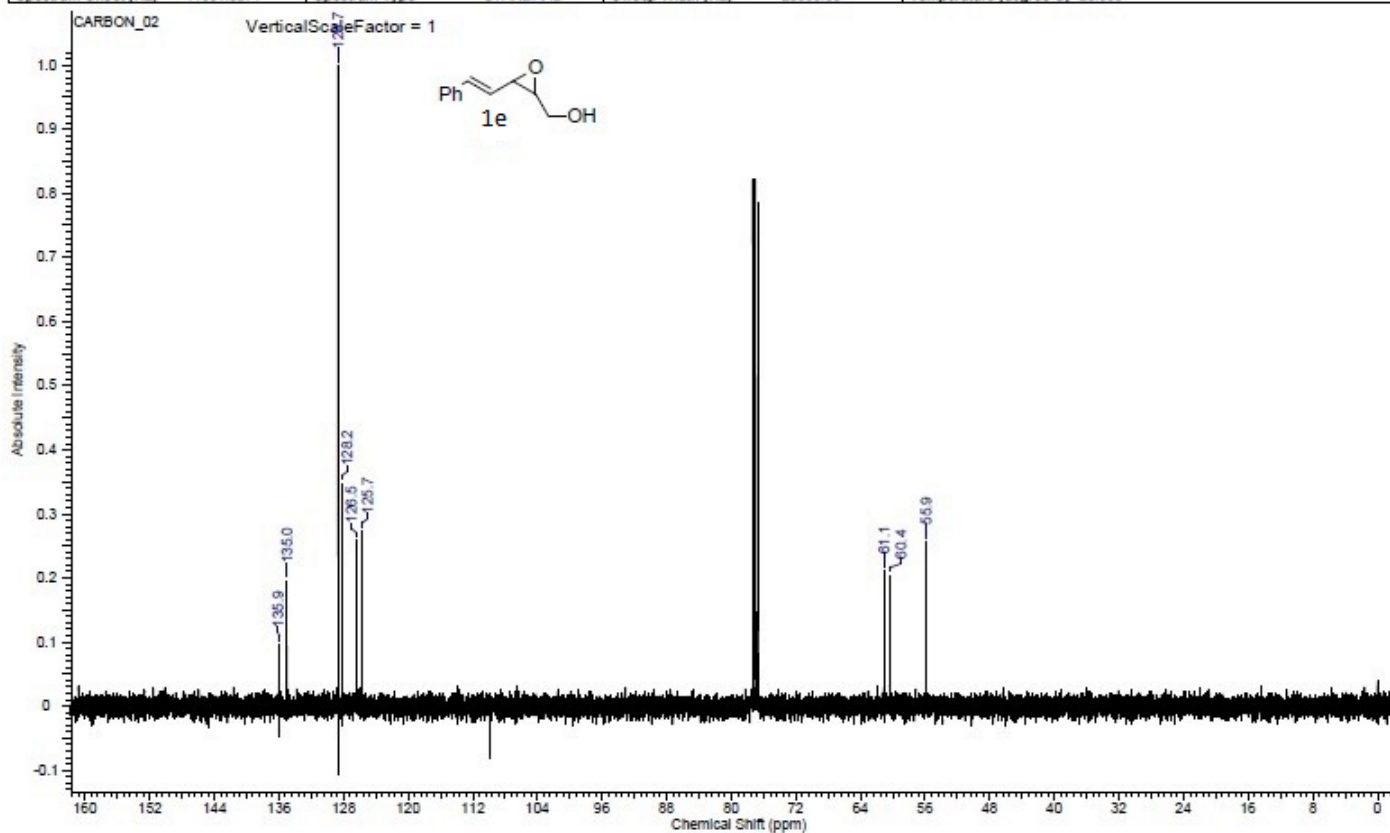
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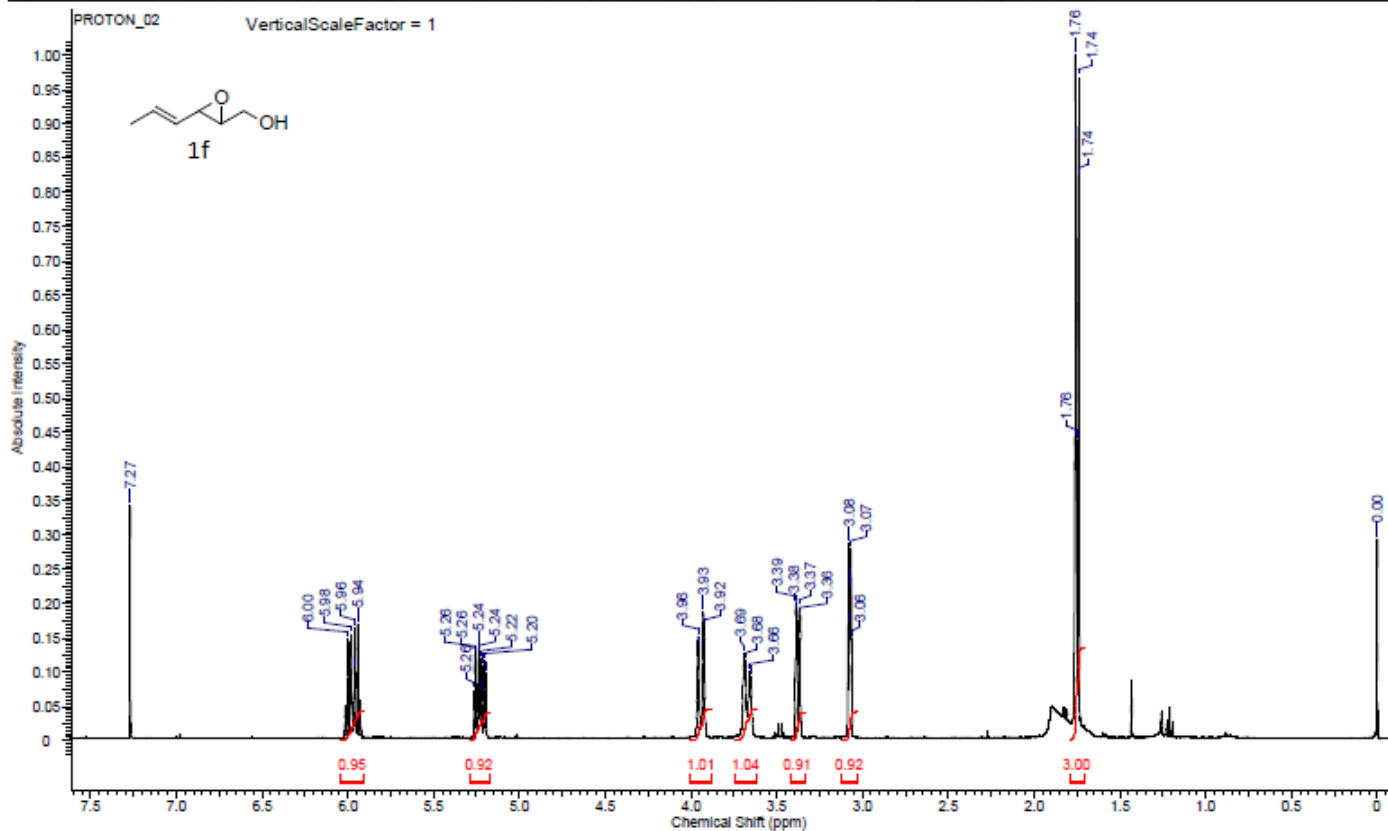
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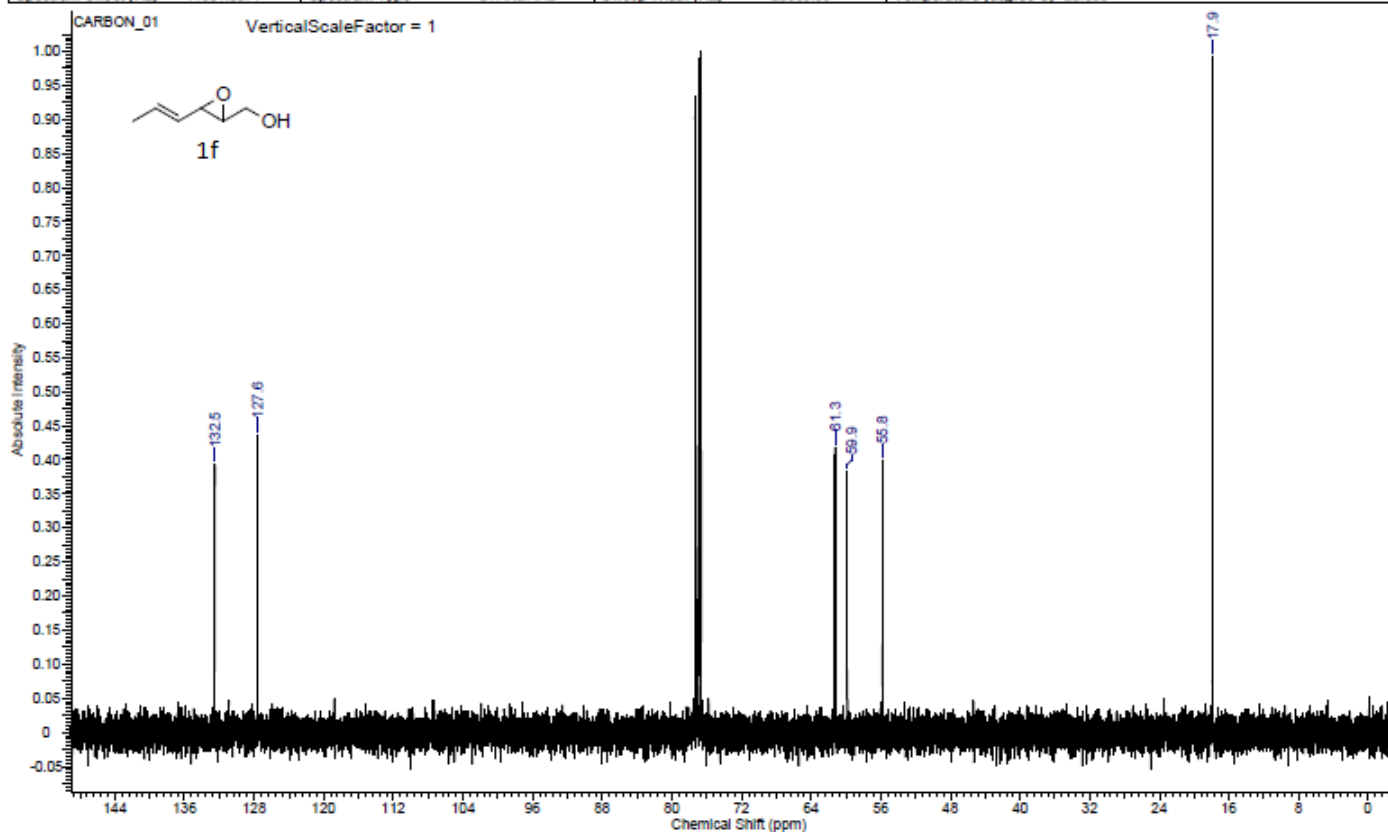
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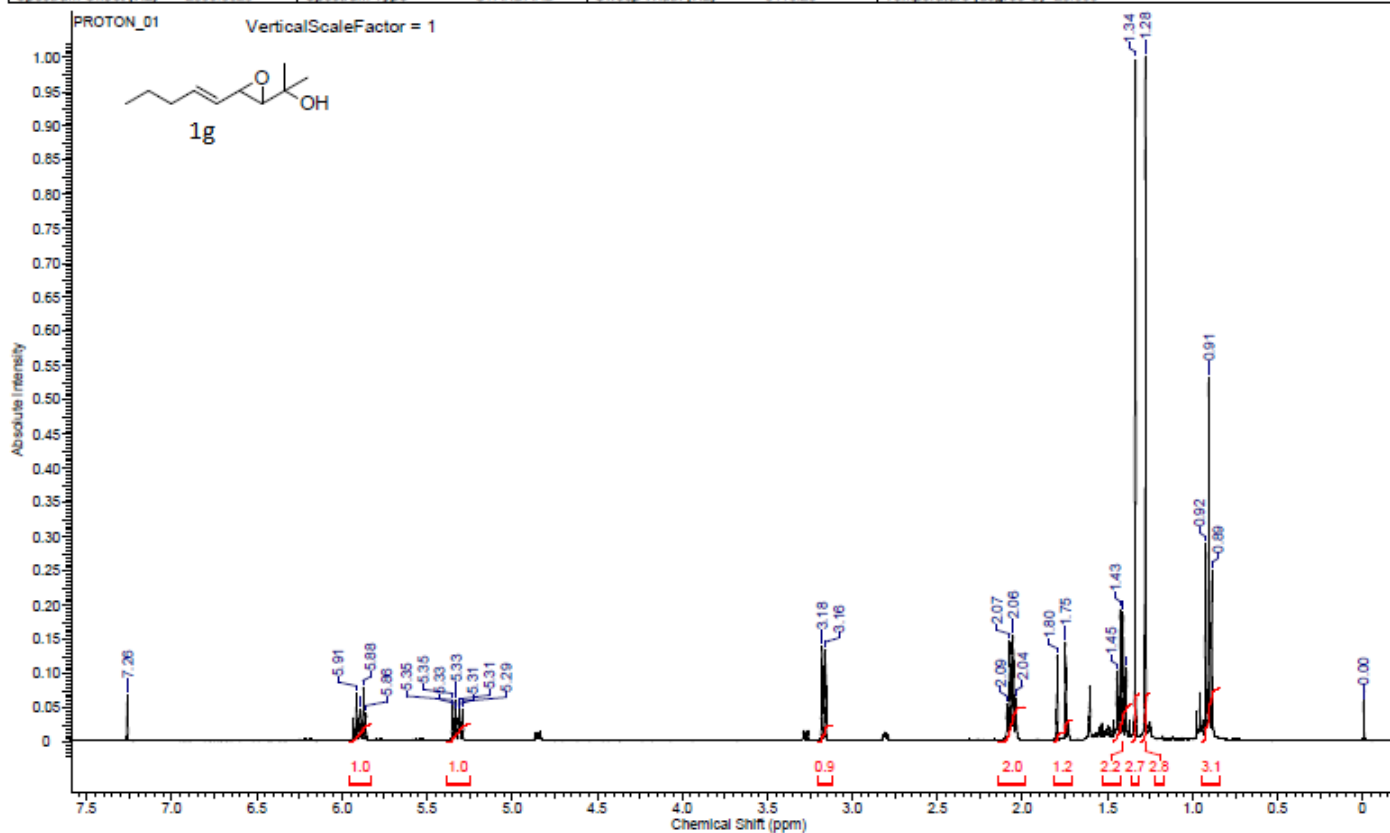
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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2403.0146	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



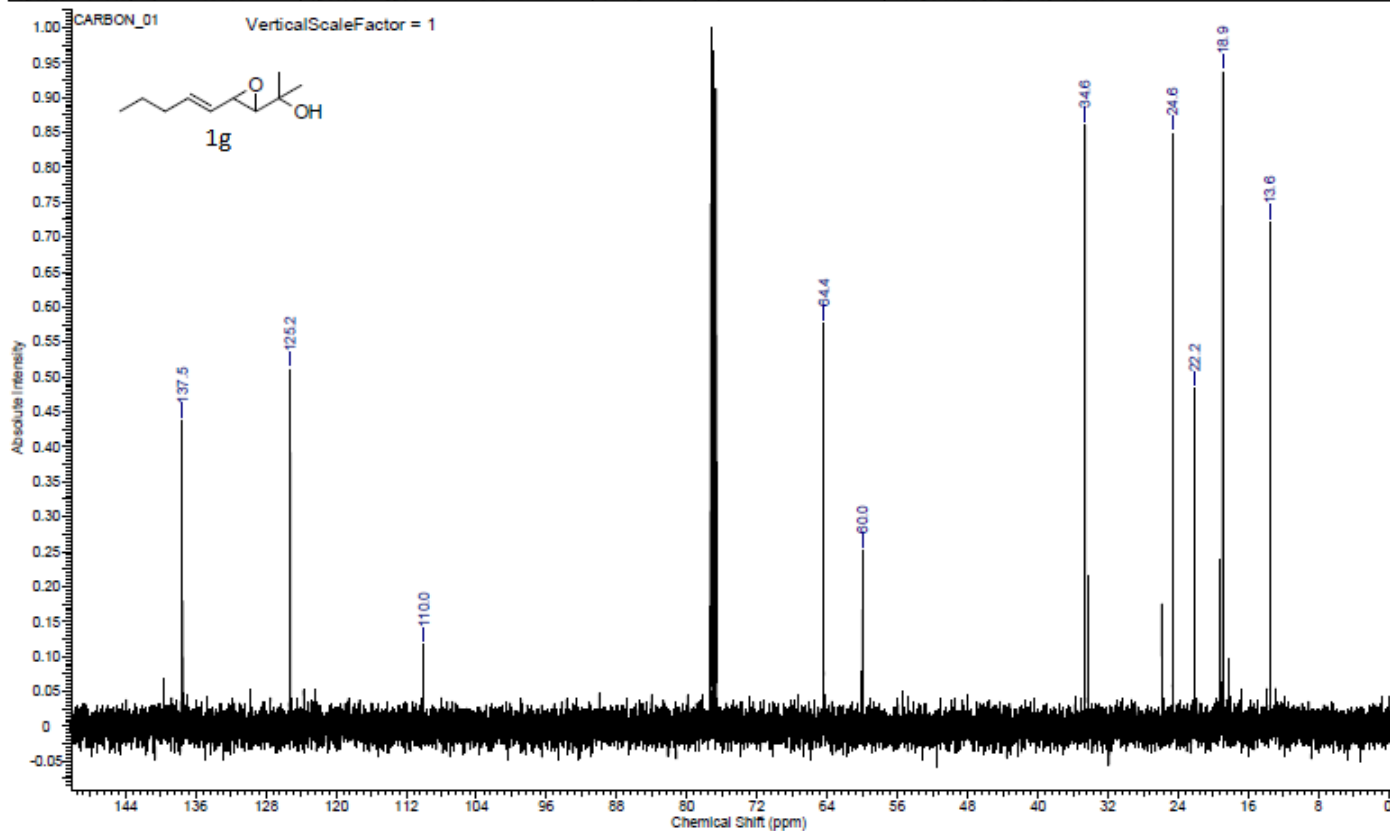
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pu1	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



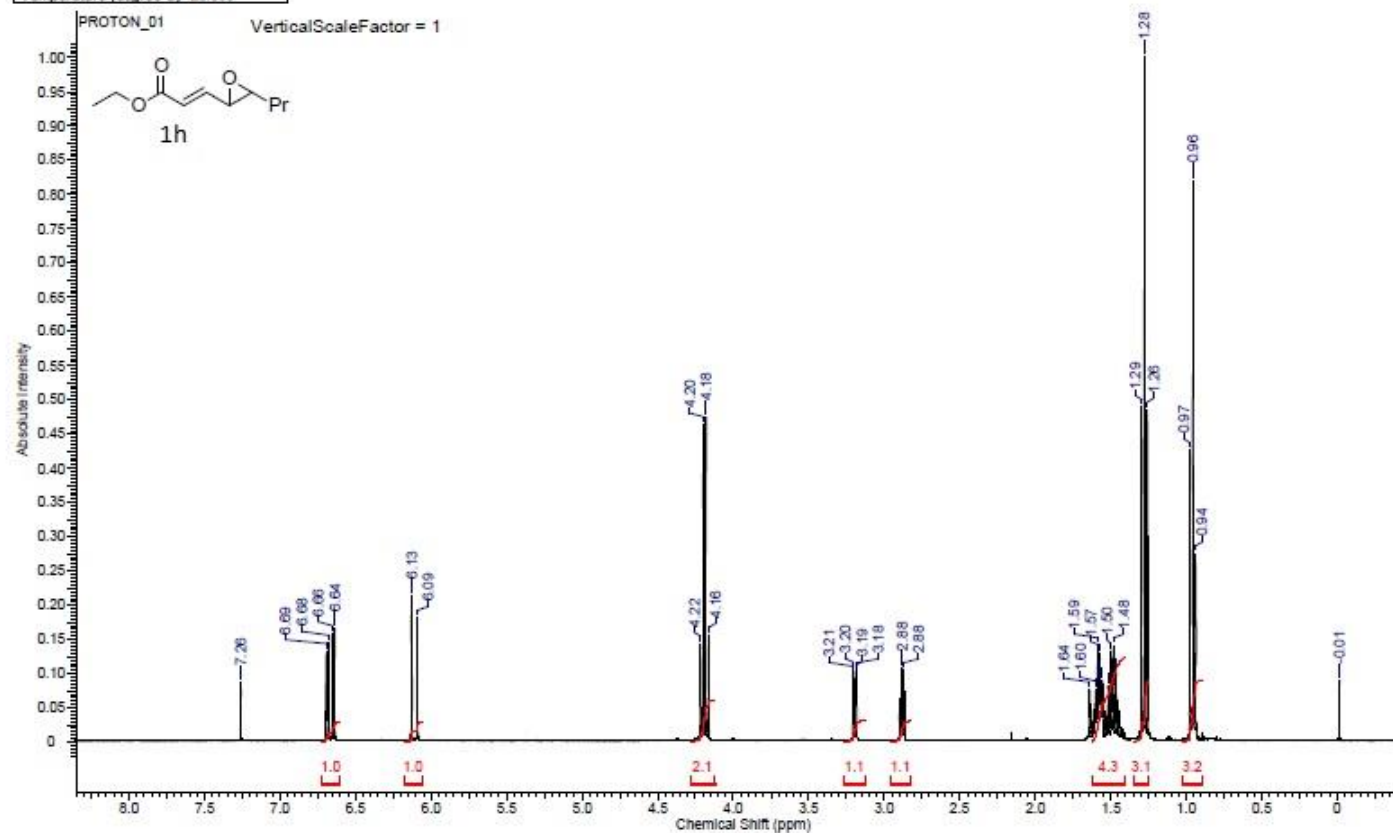
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Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



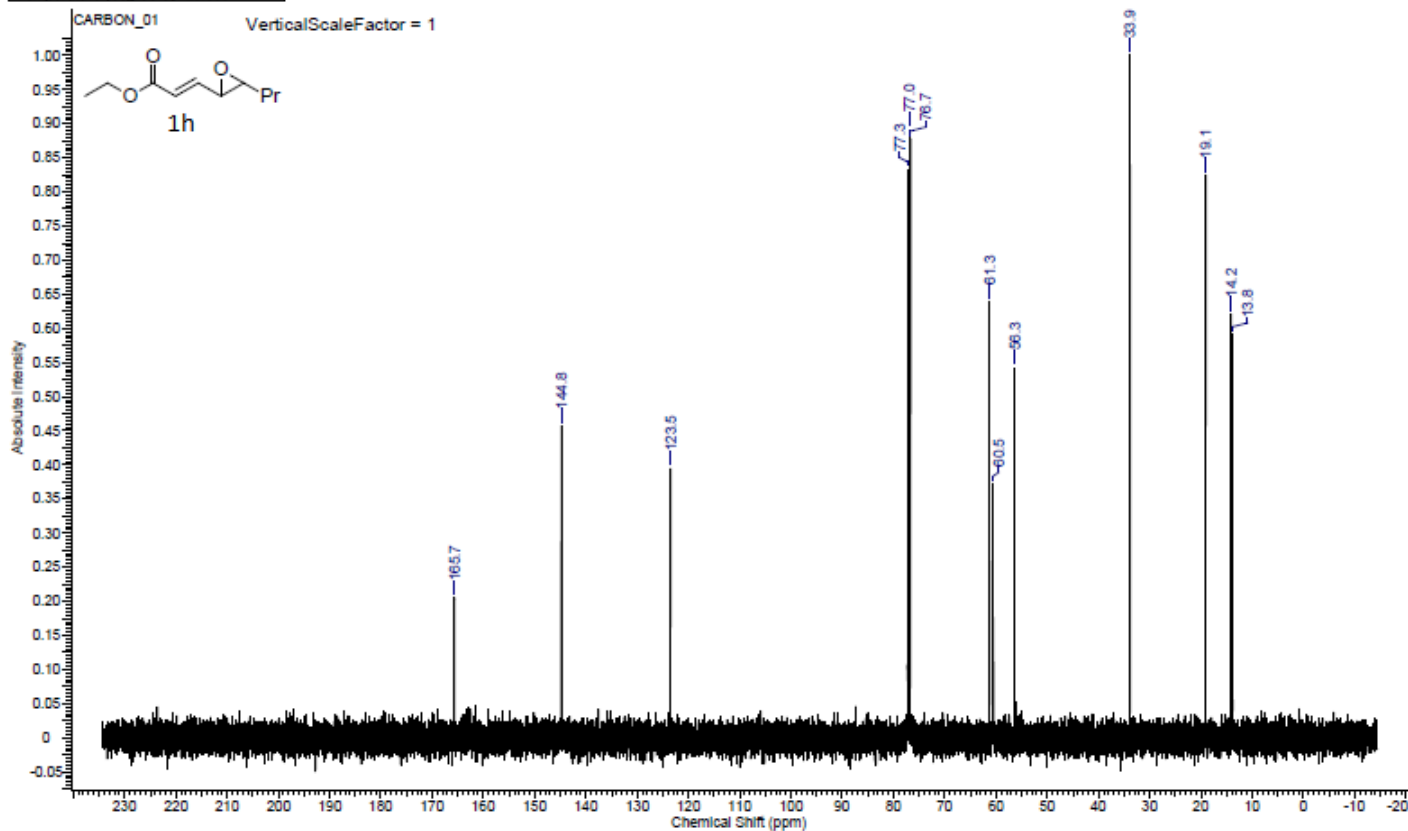
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pu1	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



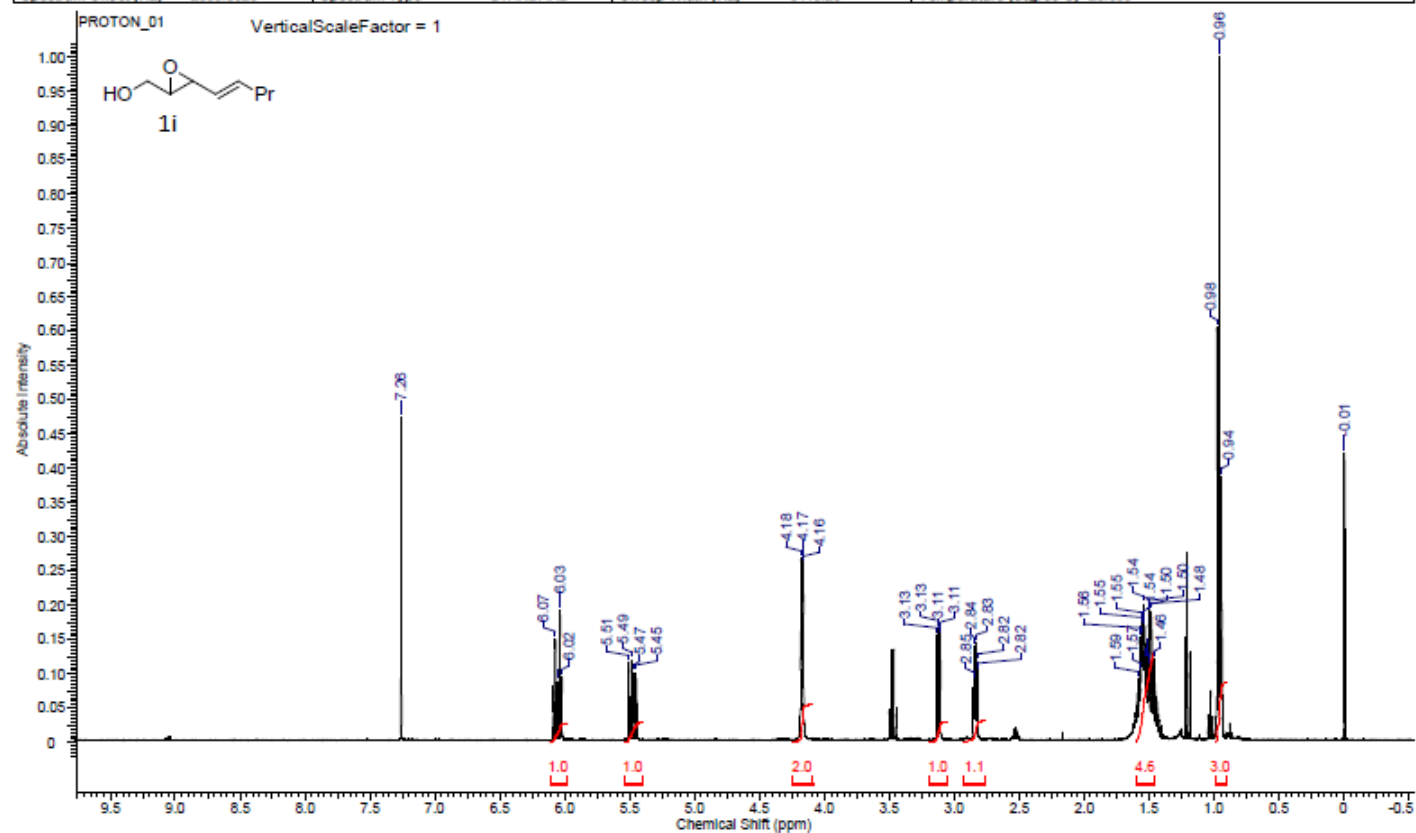
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Number of Transients	16	Original Points Count	16384	Points Count	16384	Pulse Sequence	s2pul
Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	2400.0332	Spectrum Type	STANDARD	Receiver Gain	54.00
Temperature (degree C)	25.000					Sweep Width (Hz)	6410.26



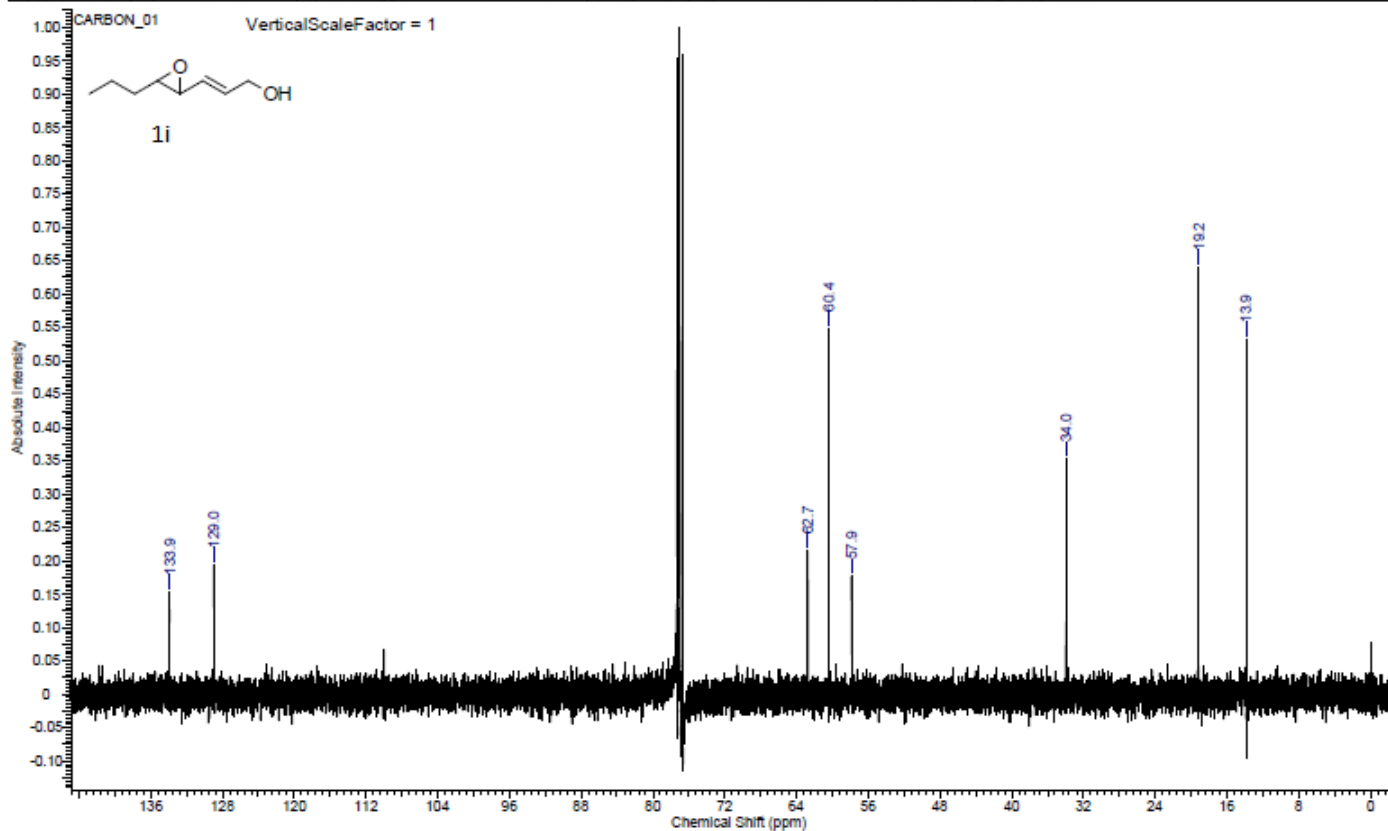
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Number of Transients	256	Original Points Count	32768	Points Count	32768	Pulse Sequence	s2pu
Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Receiver Gain	30.00
Temperature (degree C)	25.000					Sweep Width (Hz)	25000.00



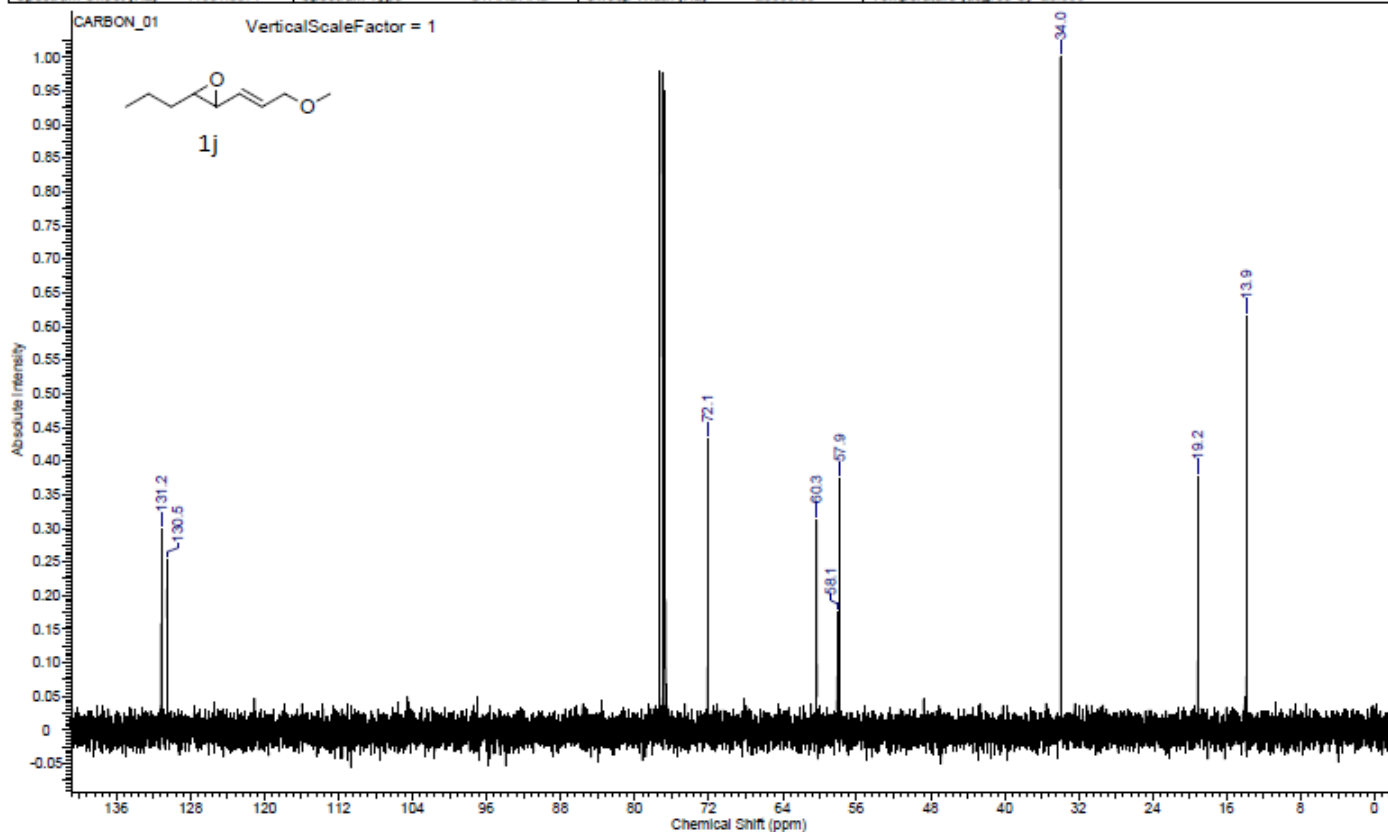
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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



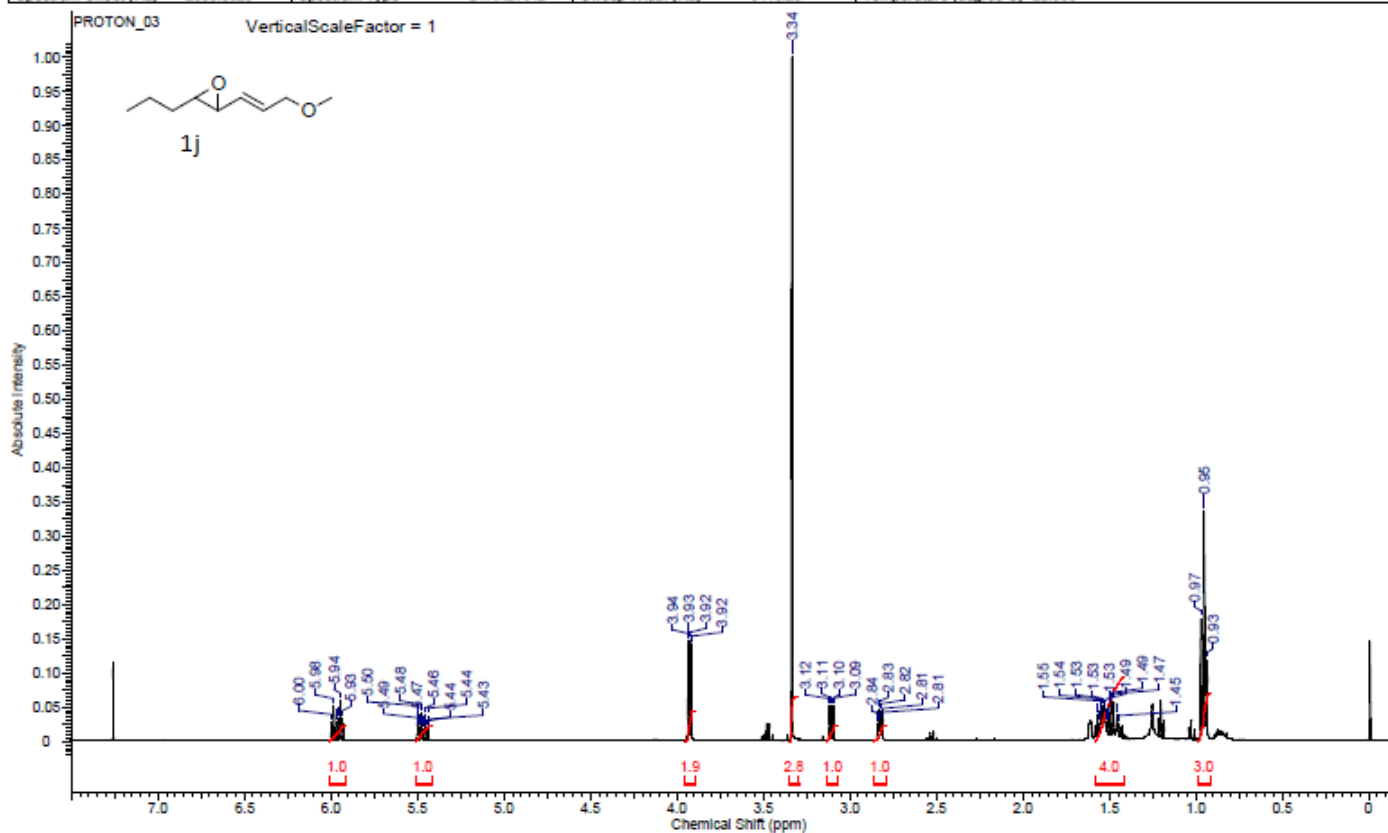
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



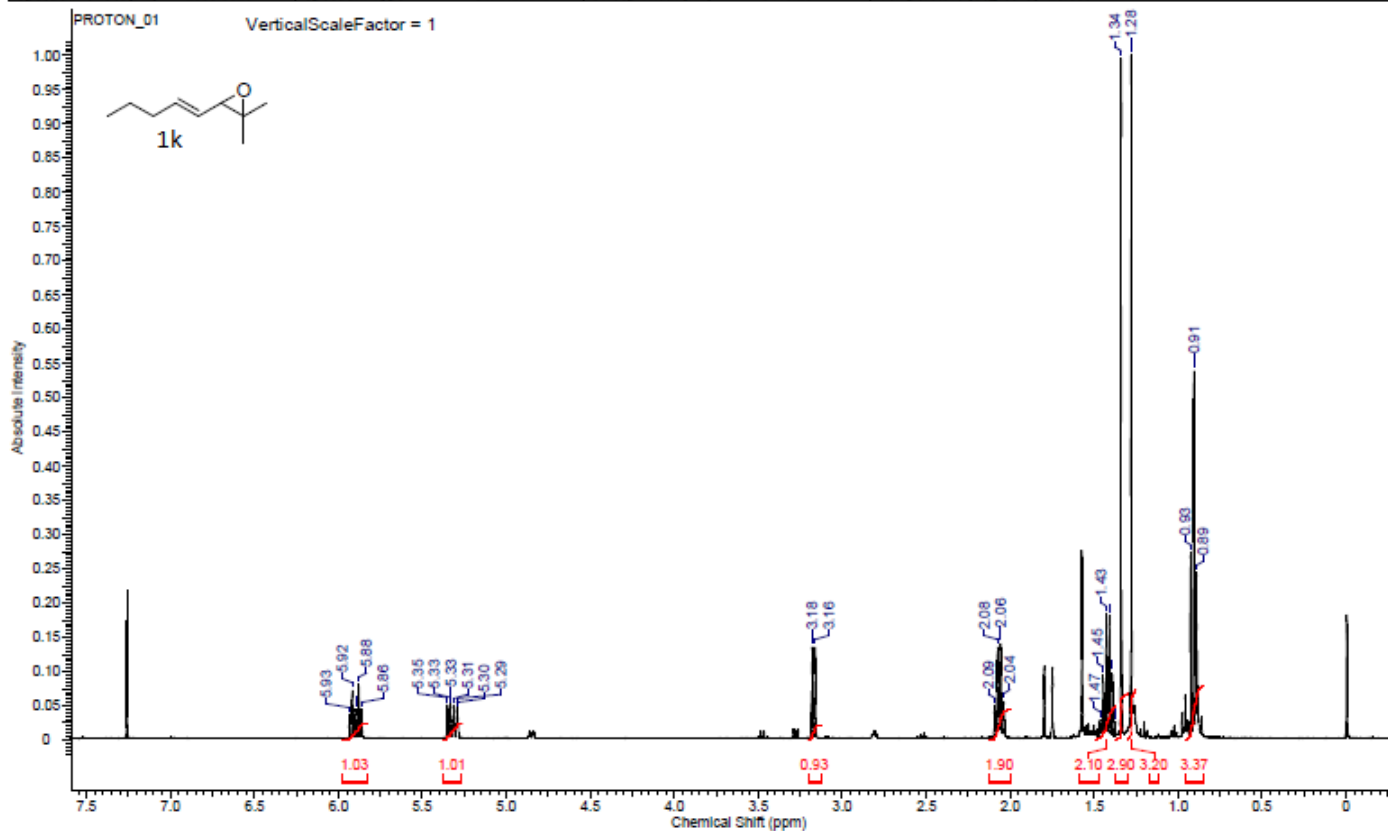
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Frequency (MHz)	100.57	Nucleus	13C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pJ	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



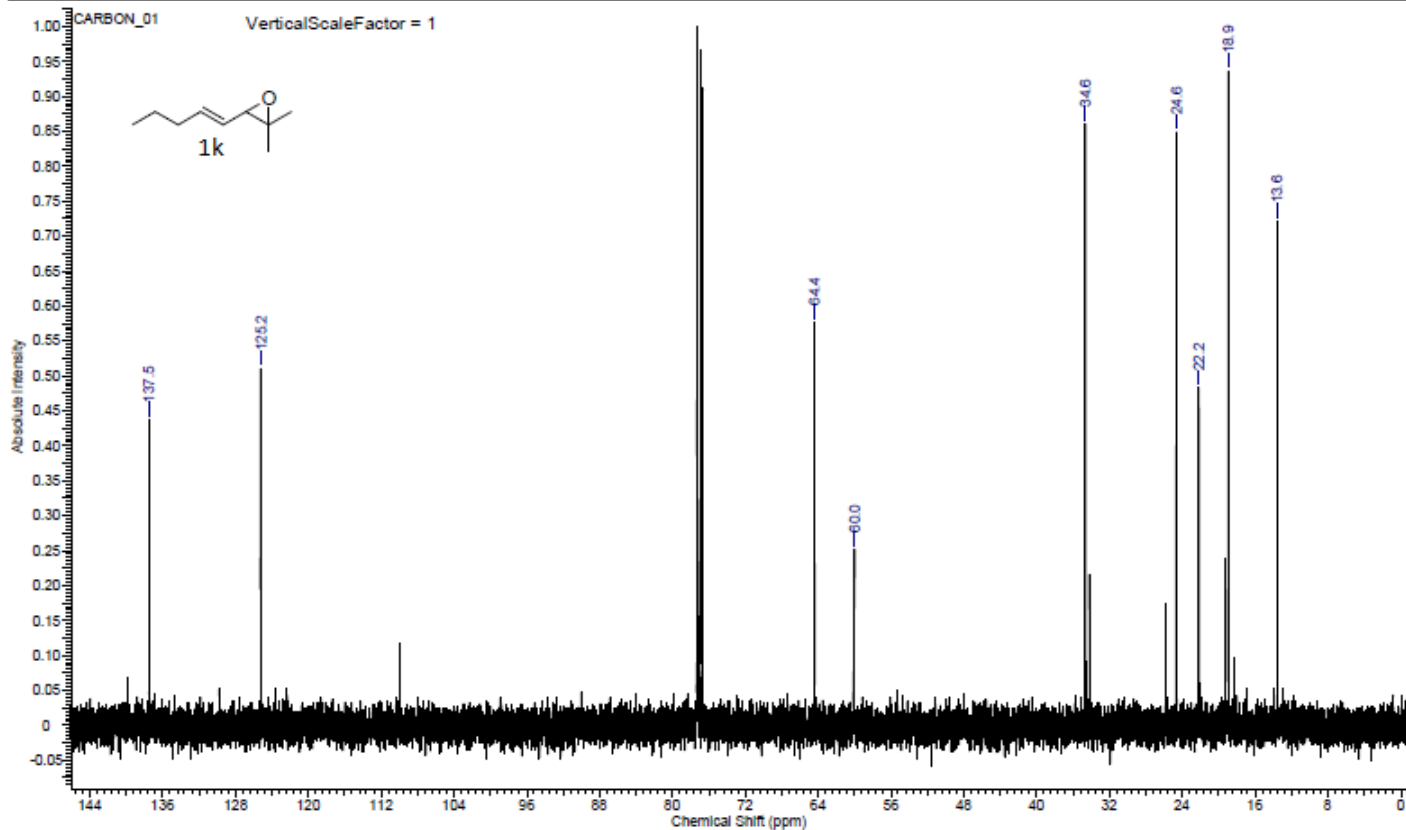
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Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



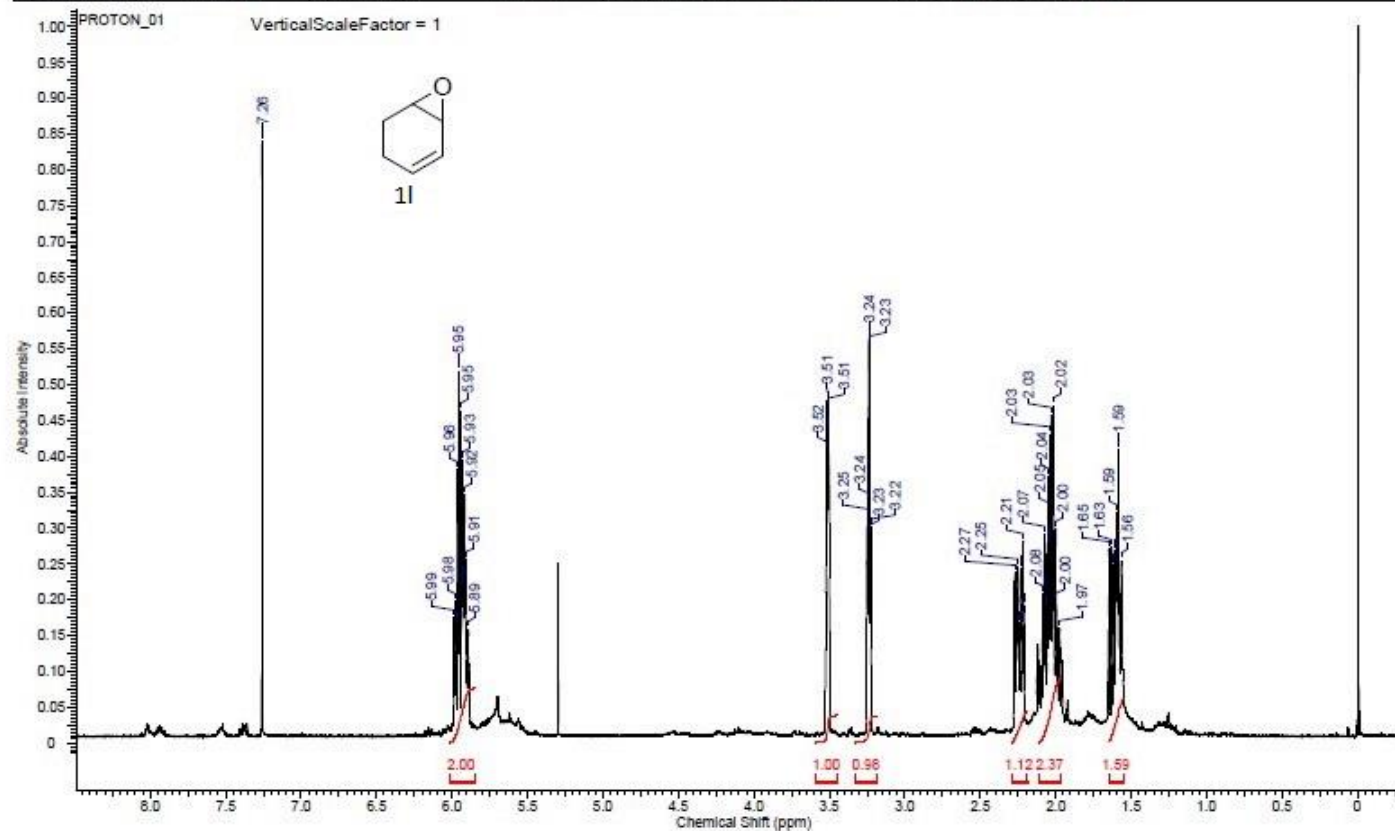
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Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



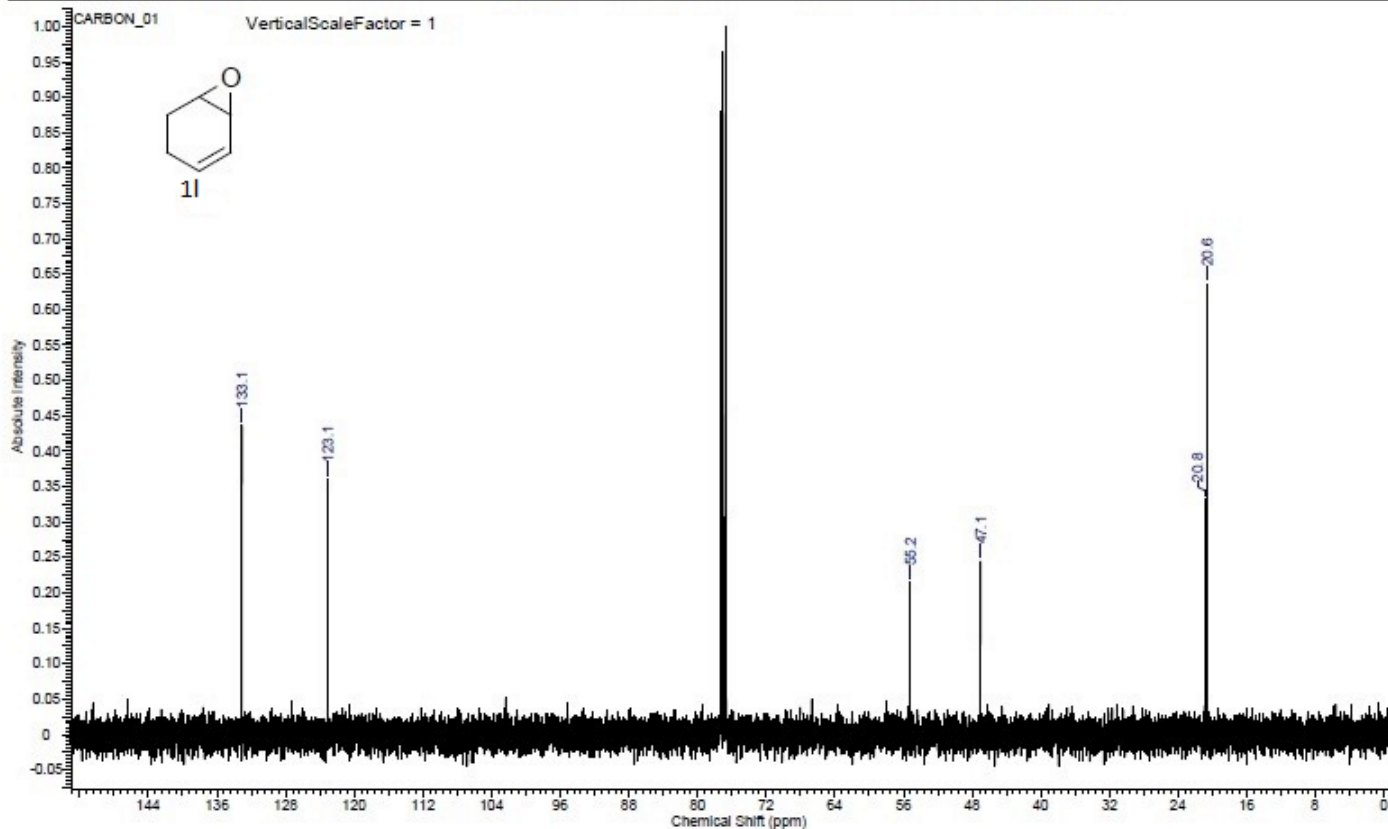
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



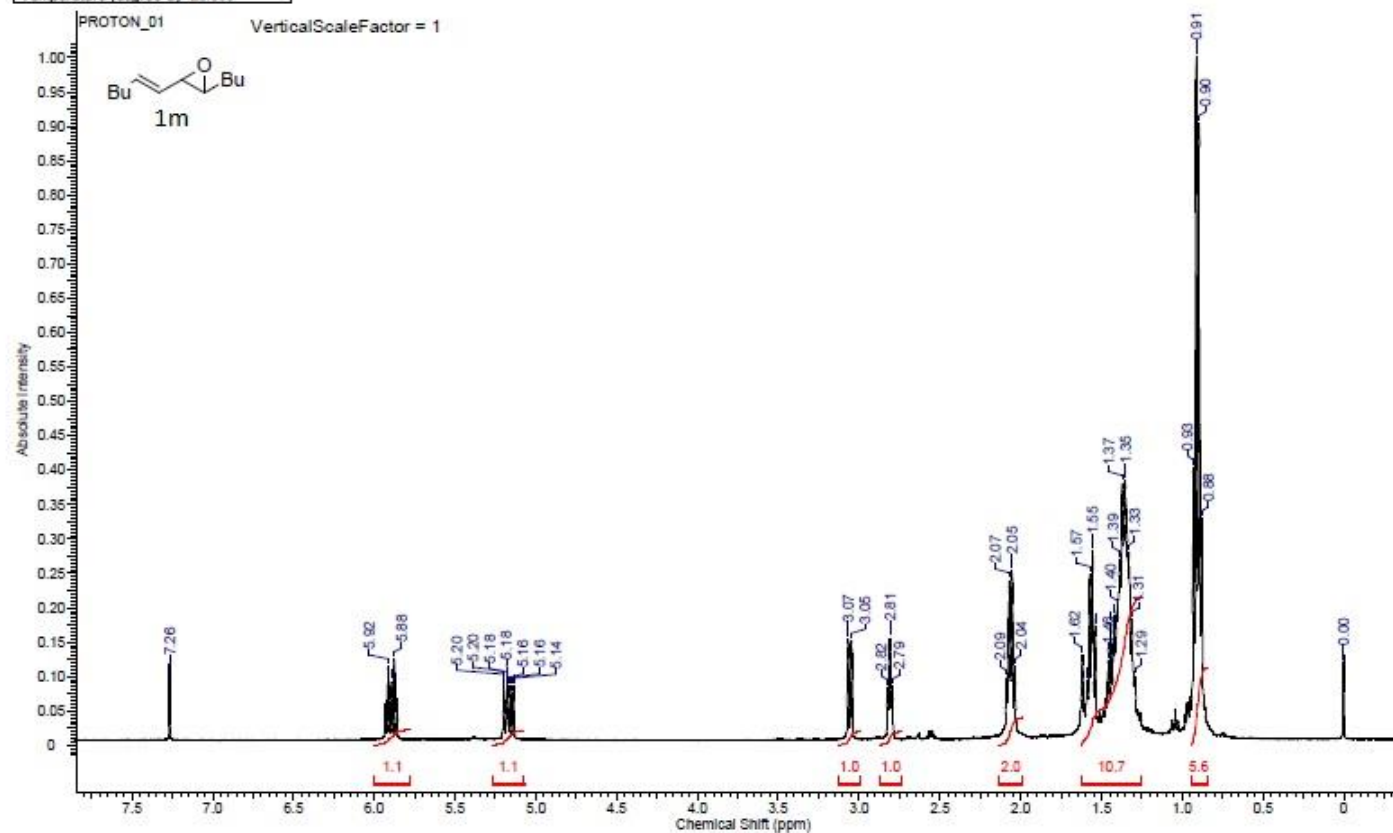
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Points Count	16384	Pulse Sequence	s2ou1	Receiver Gain	60.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Original Points Count	16384
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



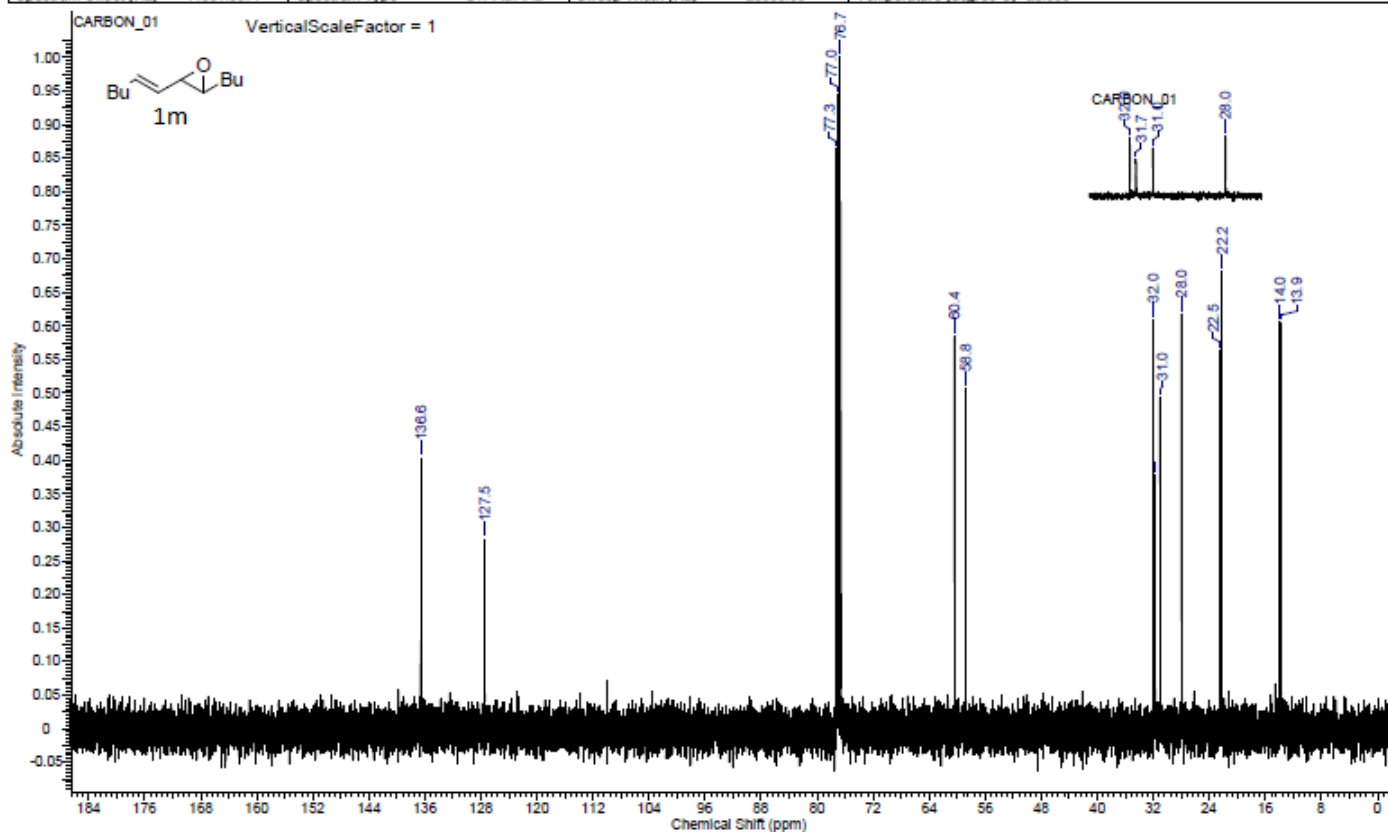
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



Acquisition Time (sec)	2.5559	Comment	ERM-STZ-BUENEPBU	Date	Jul 19 2016	Date Stamp	Jul 19 2016
File Name	F:\ERM-STZ-BUENEPBU_20160719_01\PROTON_01\FID\FID			Frequency (MHz)	399.92	Nucleus	¹ H
Number of Transients	8	Original Points Count	16384	Points Count	16384	Pulse Sequence	s2ou
Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	2403.4060	Spectrum Type	STANDARD	Receiver Gain	60.00
Temperature (degree C)	25.000					Sweep Width (Hz)	6410.26



Acquisition Time (sec)	1.3107	Comment	ERM-STZ-BUENEPBU		Date	Jul 19 2016	
Date Stamp	Jul 19 2016	File Name	F:\ERM-STZ-BUENEPBU_20160719_01\CARBON_01.FID\FID				
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2bul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



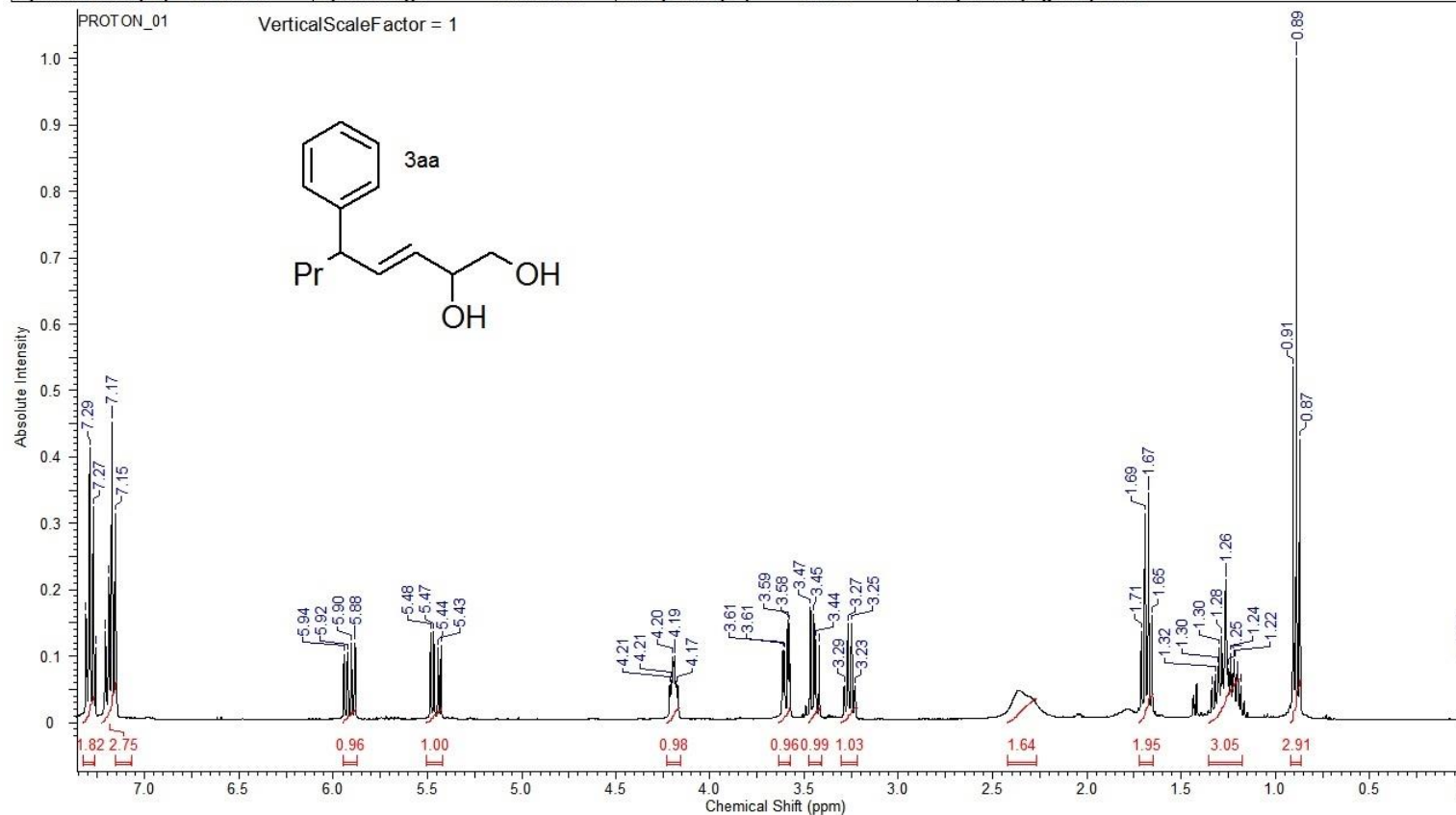
APPENDIX B

^1H NMR AND ^{13}C NMR SPECTRUMS OF PRODUCTS

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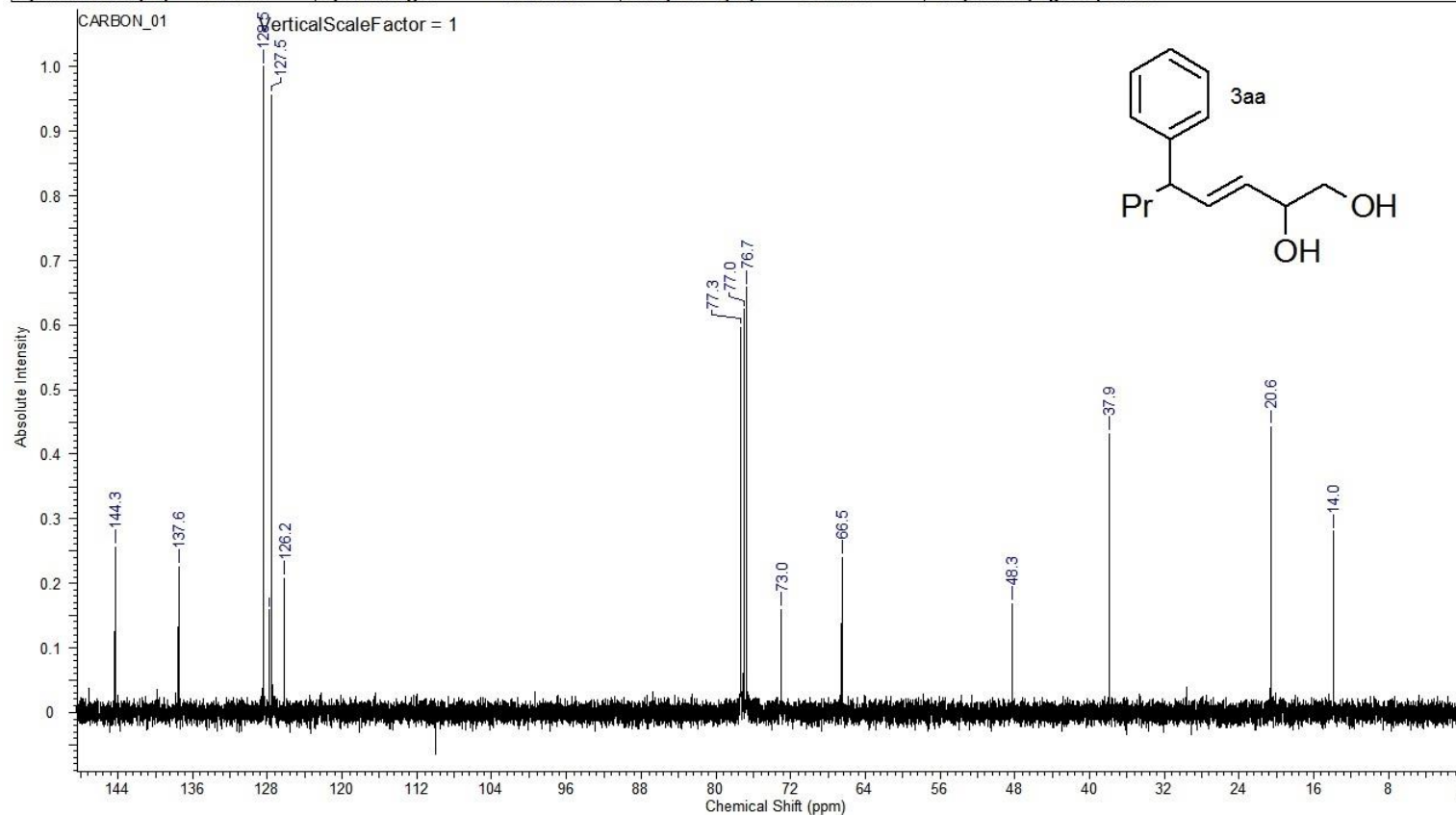
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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	32
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	54.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



C:\Users\Erman\Desktop\ERMAN\ERM-ENEP-138-1_3_20150904_02\PROTON_01

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Date Stamp	Sep 4 2015	File Name	C:\Users\Erman\Desktop\ERMAN\ERM-ENEP-138-1_3_20150904_02\CARBON_01.fid				
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Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000

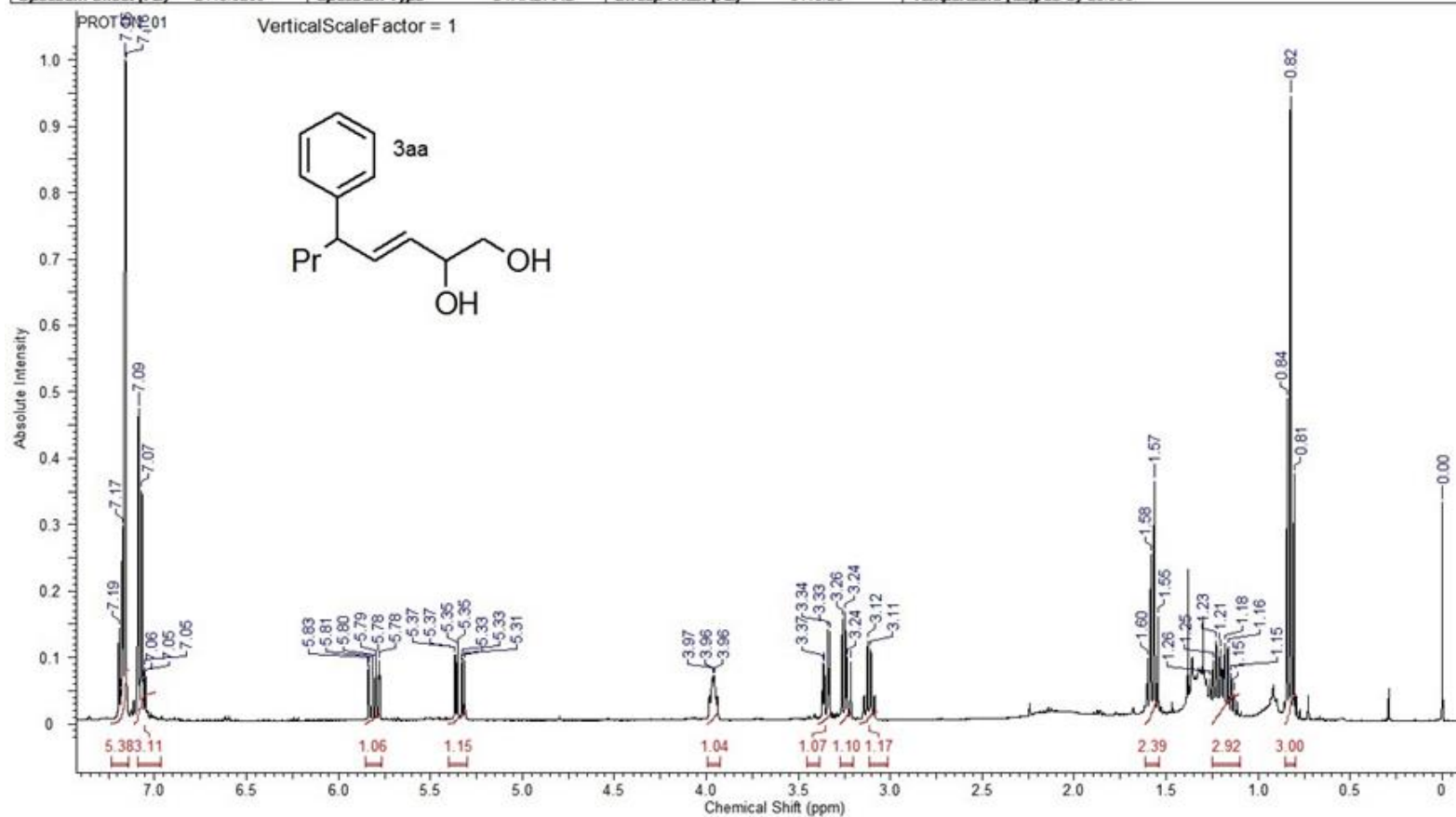


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10.10.2015 04:43:12

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Date Stamp	Oct 6 2015	File Name	C:\Users\Erman\Desktop\123\ERM-ENEP-155-1_3_20151006_02\PROTON_01.fid\fid		
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	32
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	58.00
Spectrum Offset (Hz)	2415.9268	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000

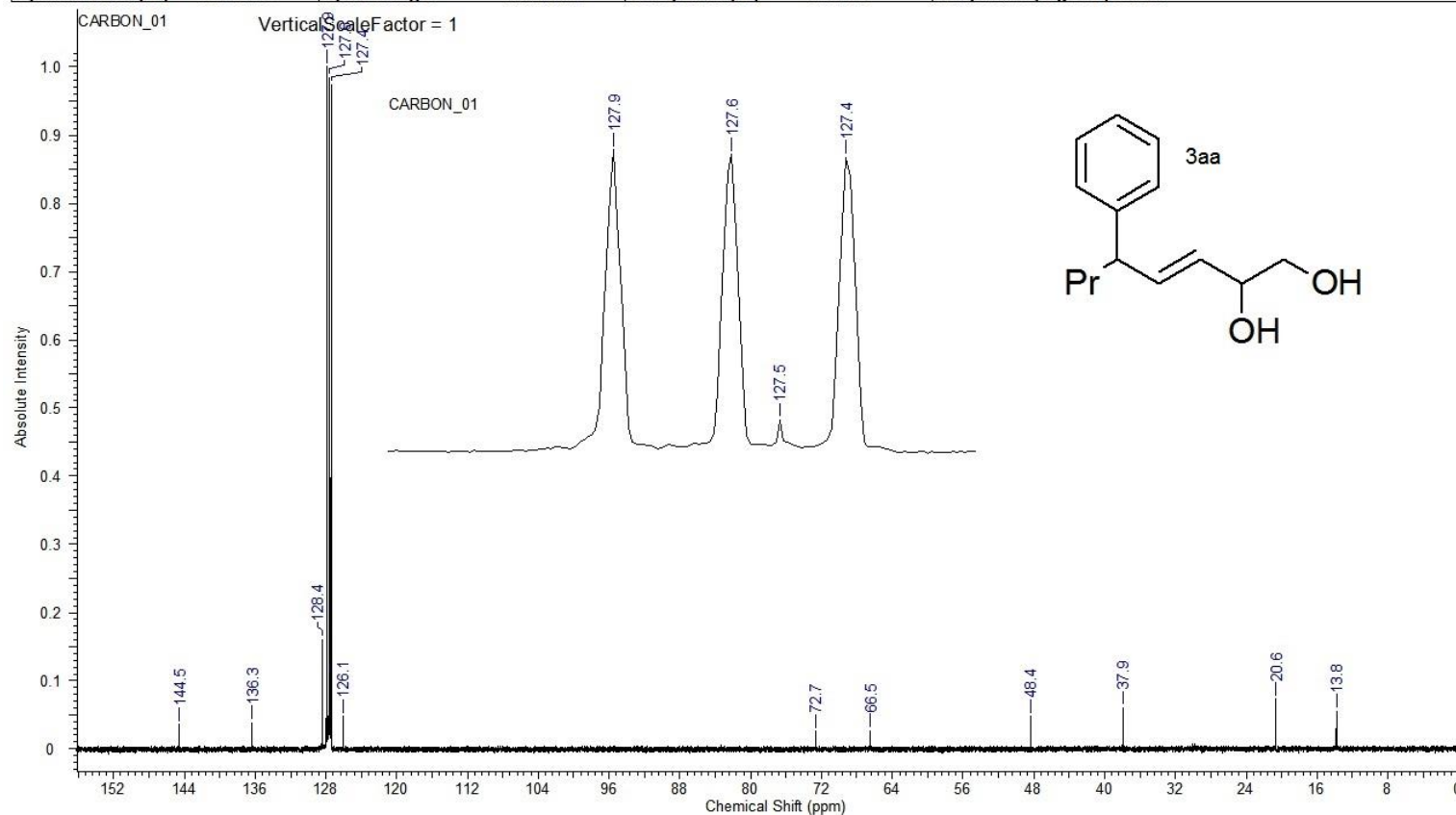


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10.10.2015 04:12:51

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Frequency (MHz)	100.57	Nucleus	13C	Number of Transients	512	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	BENZENE-d6
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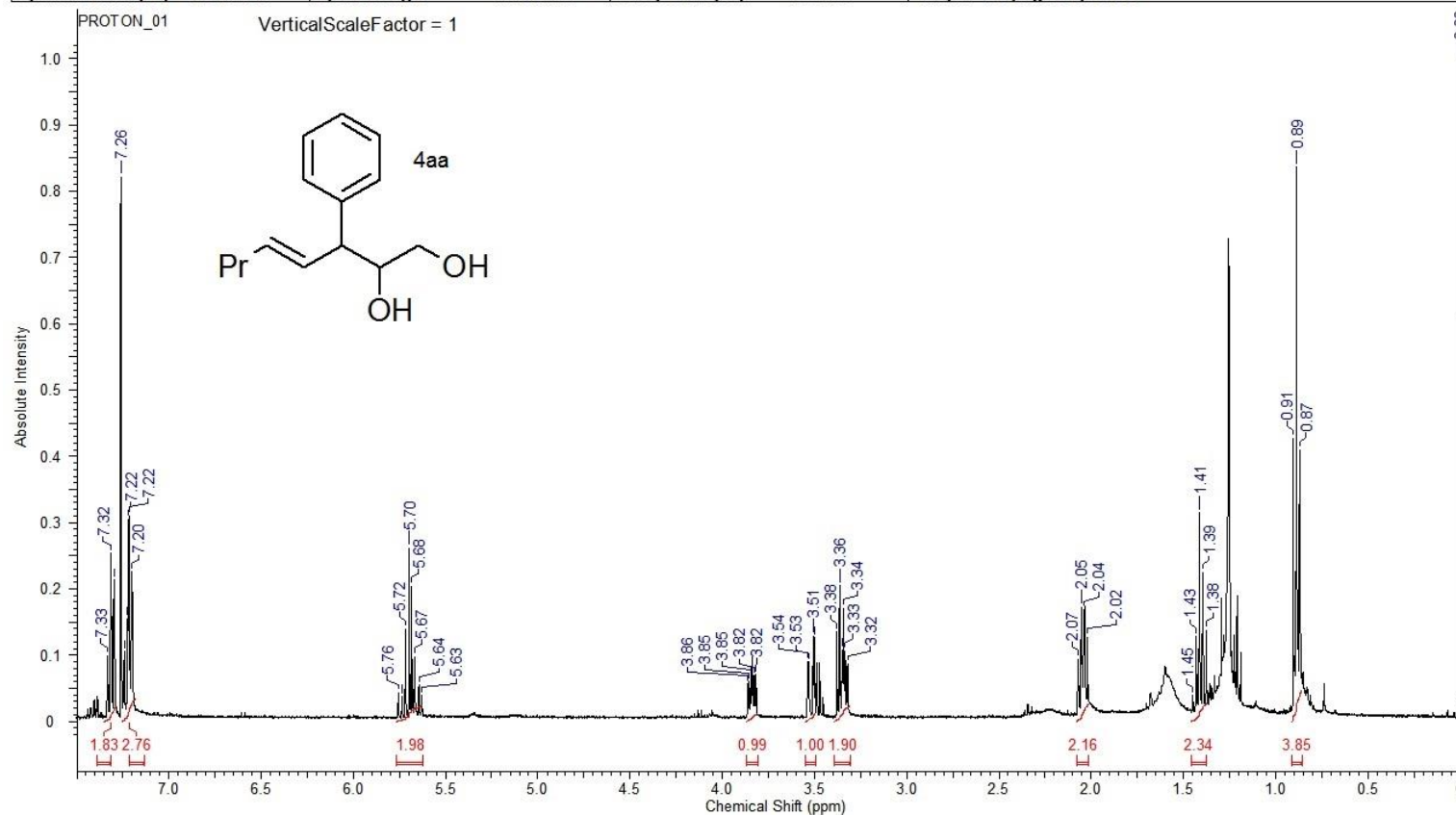


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This report was created by ACD/NMR Processor Academic Edition. For more information go to www.acdlabs.com/nmrproc/

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Date Stamp	Oct 5 2015	File Name	C:\Users\Erman\Desktop\123\ERM-ENEP-155-1_1_20151005_01\PROTON_01.fid.fid		
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Original Points Count	16384
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000

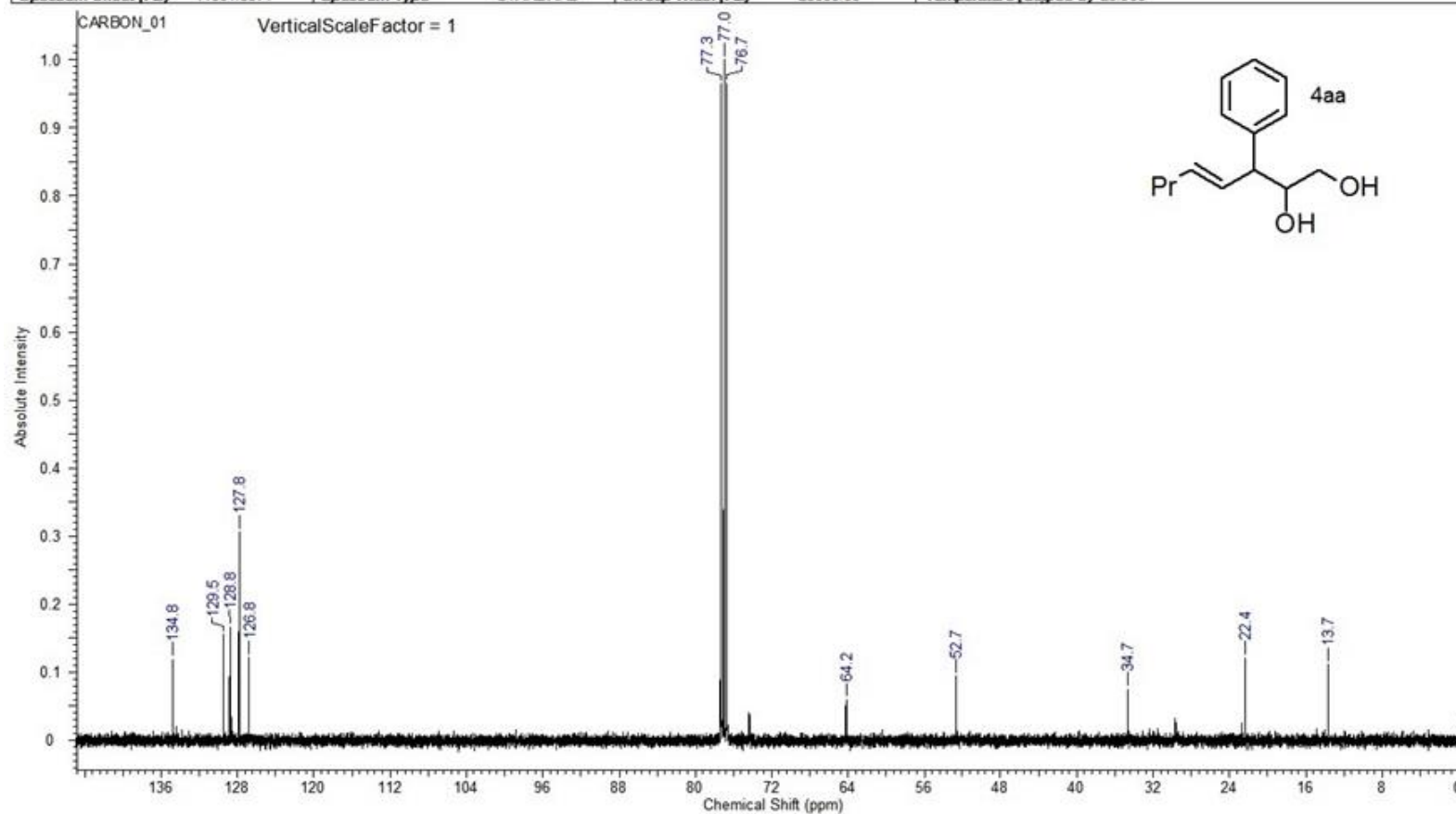


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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	2500	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000

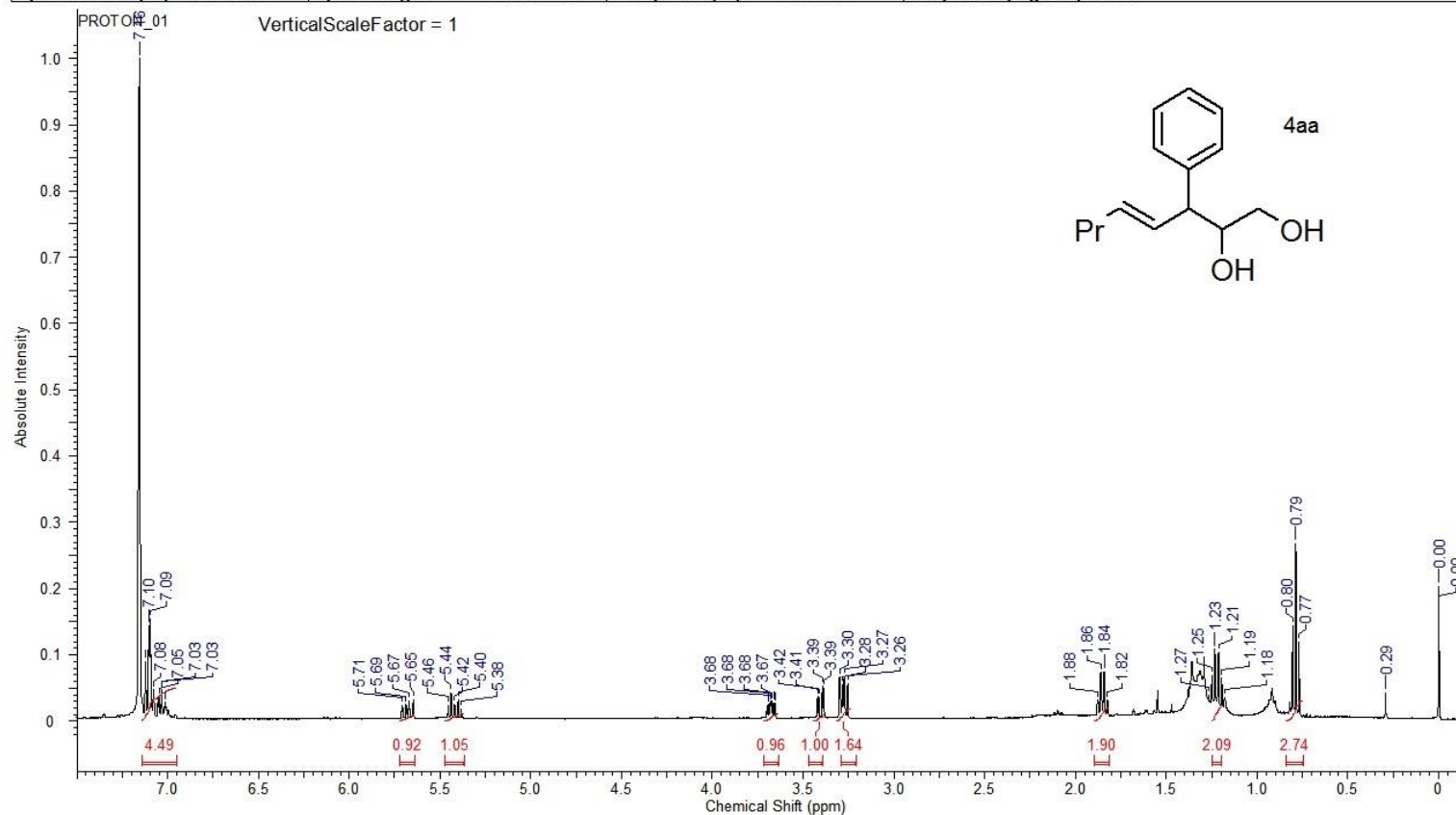


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Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	32
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00
Spectrum Offset (Hz)	2415.1443	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000

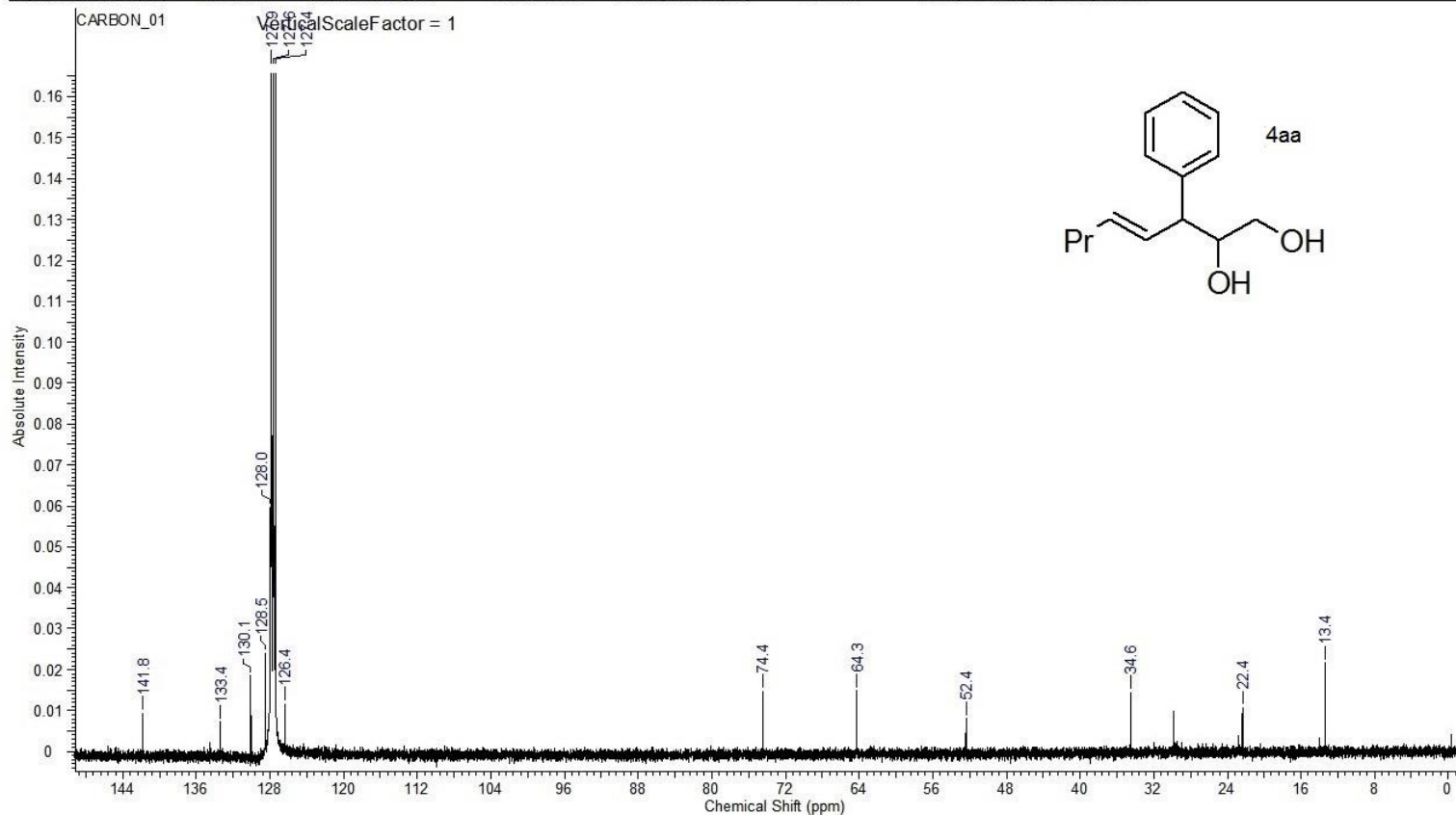


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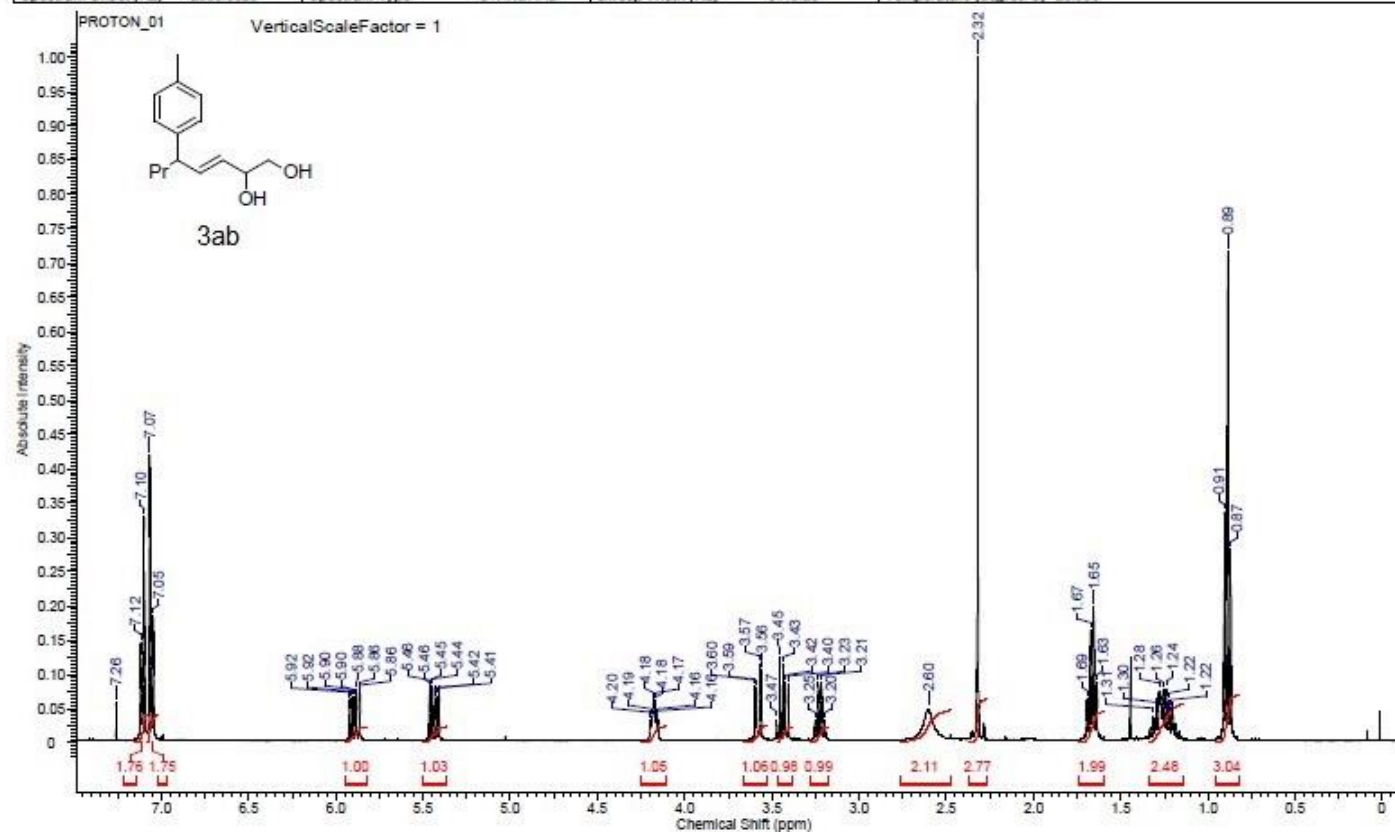
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	5000	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000

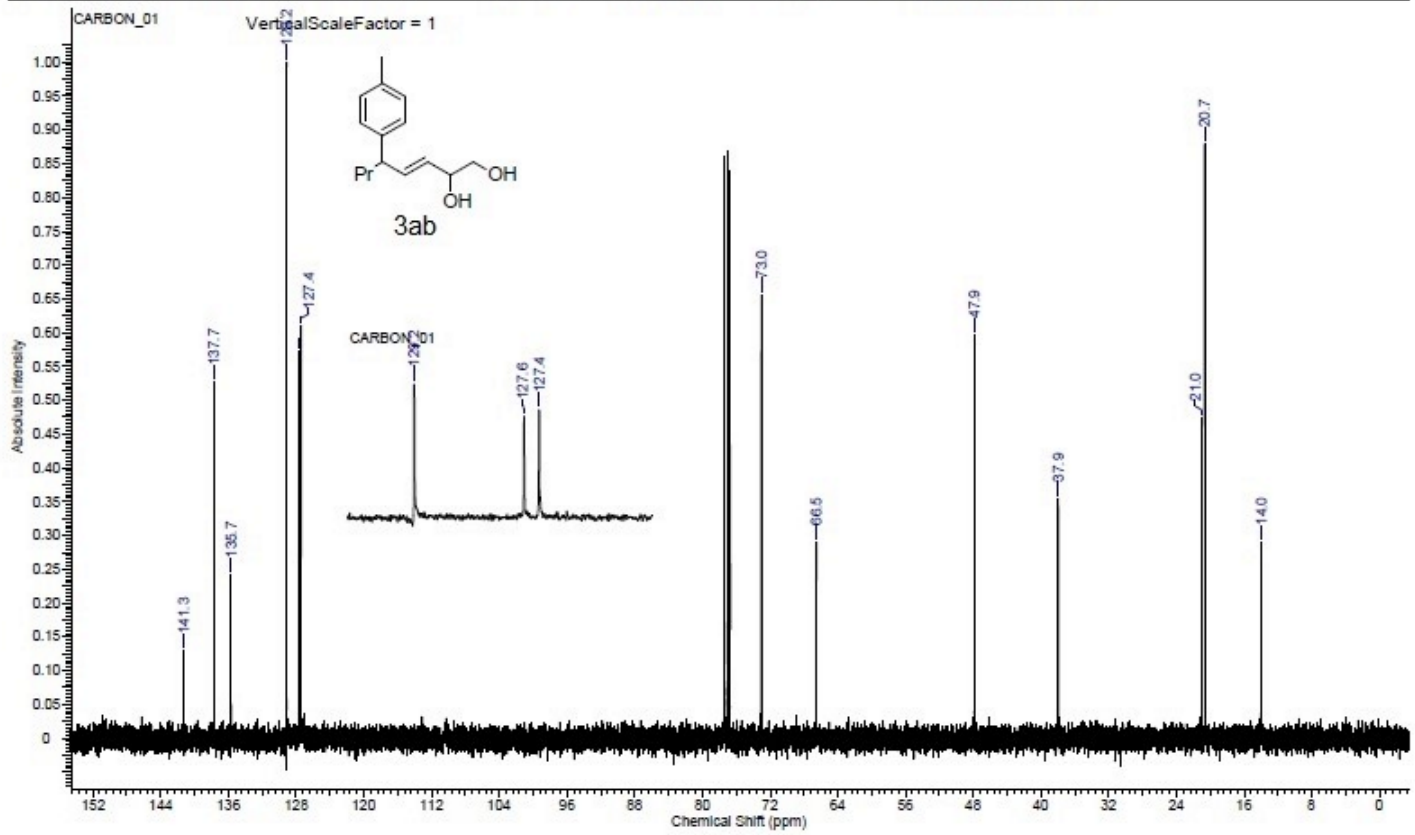


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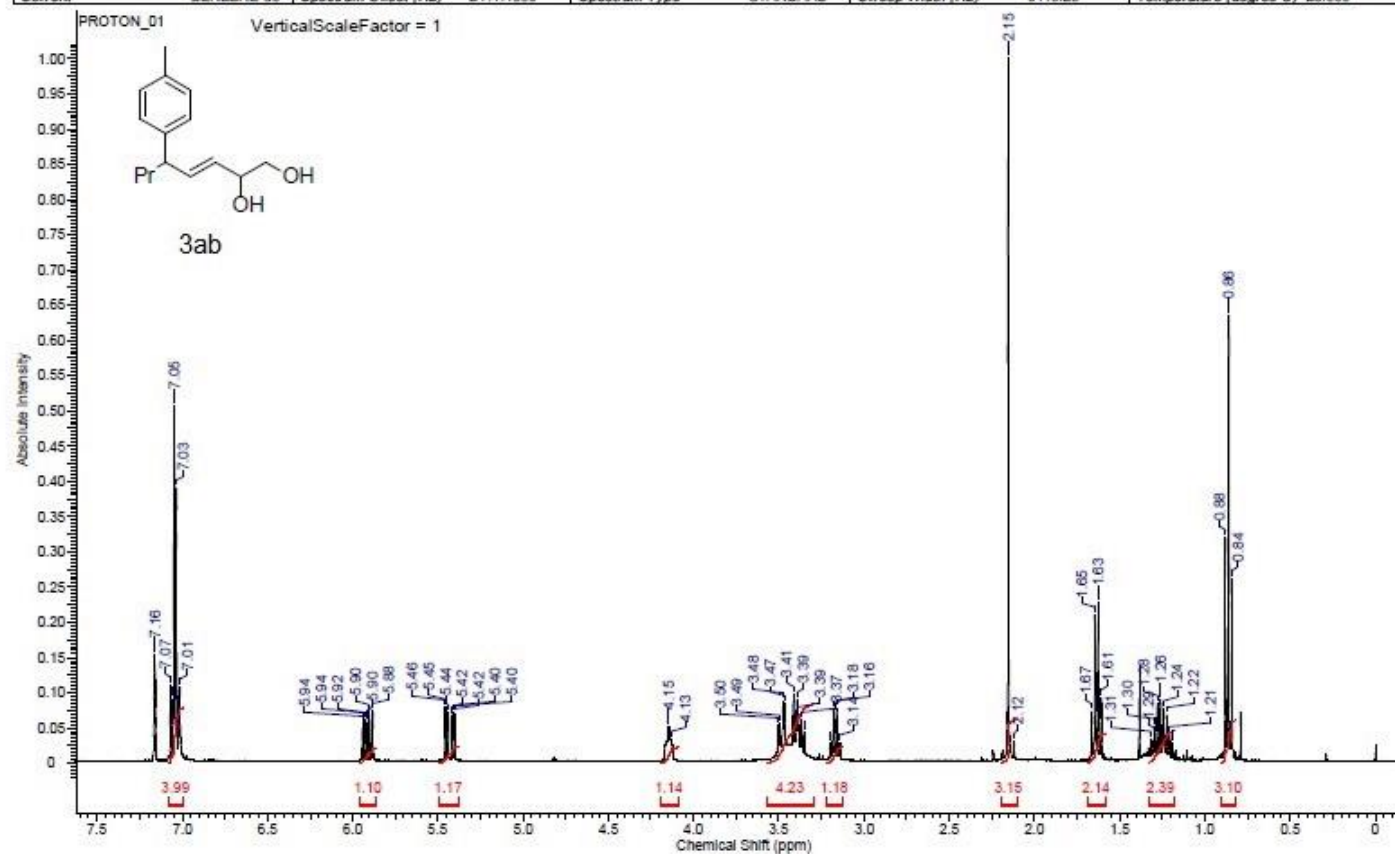
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Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	50.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



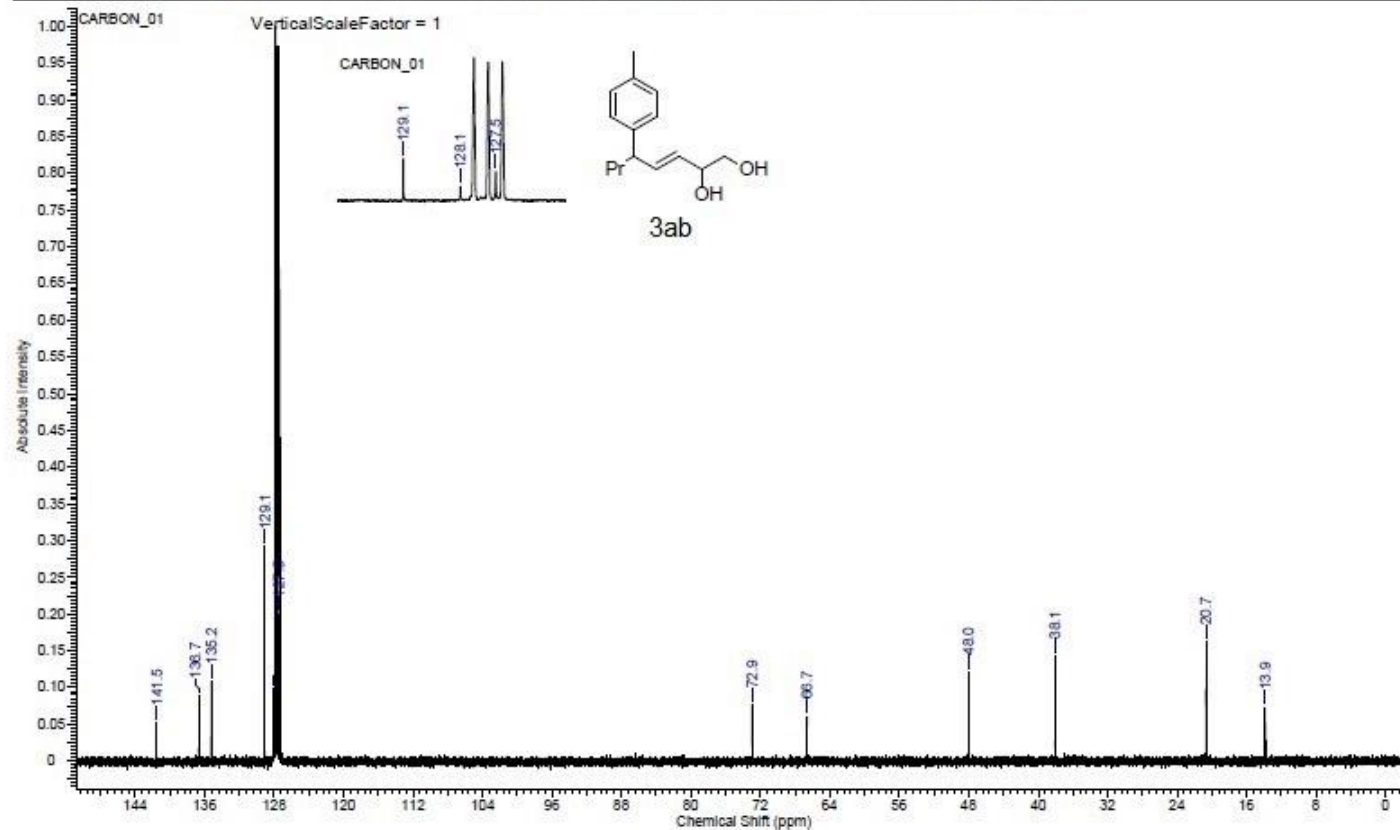
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Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
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				Temperature (degree C)	25.000



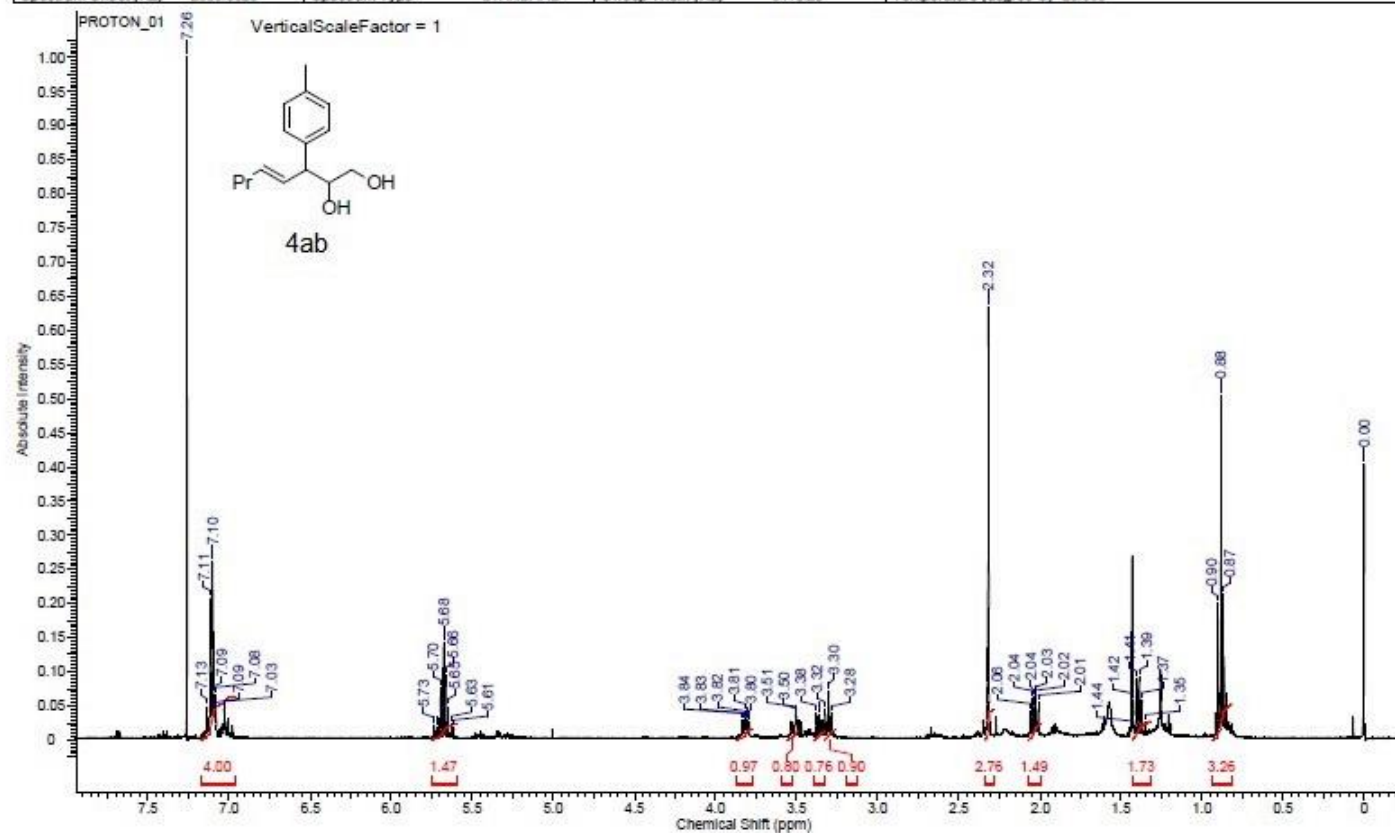
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File Name	D:\NMR\ENI\ERM-ENEP-207-1_3_20160405_01\PROTON_01.FID\FID				Frequency (MHz)	399.92	Nucleus	¹ H	
Number of Transients	8	Original Points Count	16384	Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	52.00
Solvent	BENZENE-d6	Spectrum Offset (Hz)	2417.1006	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



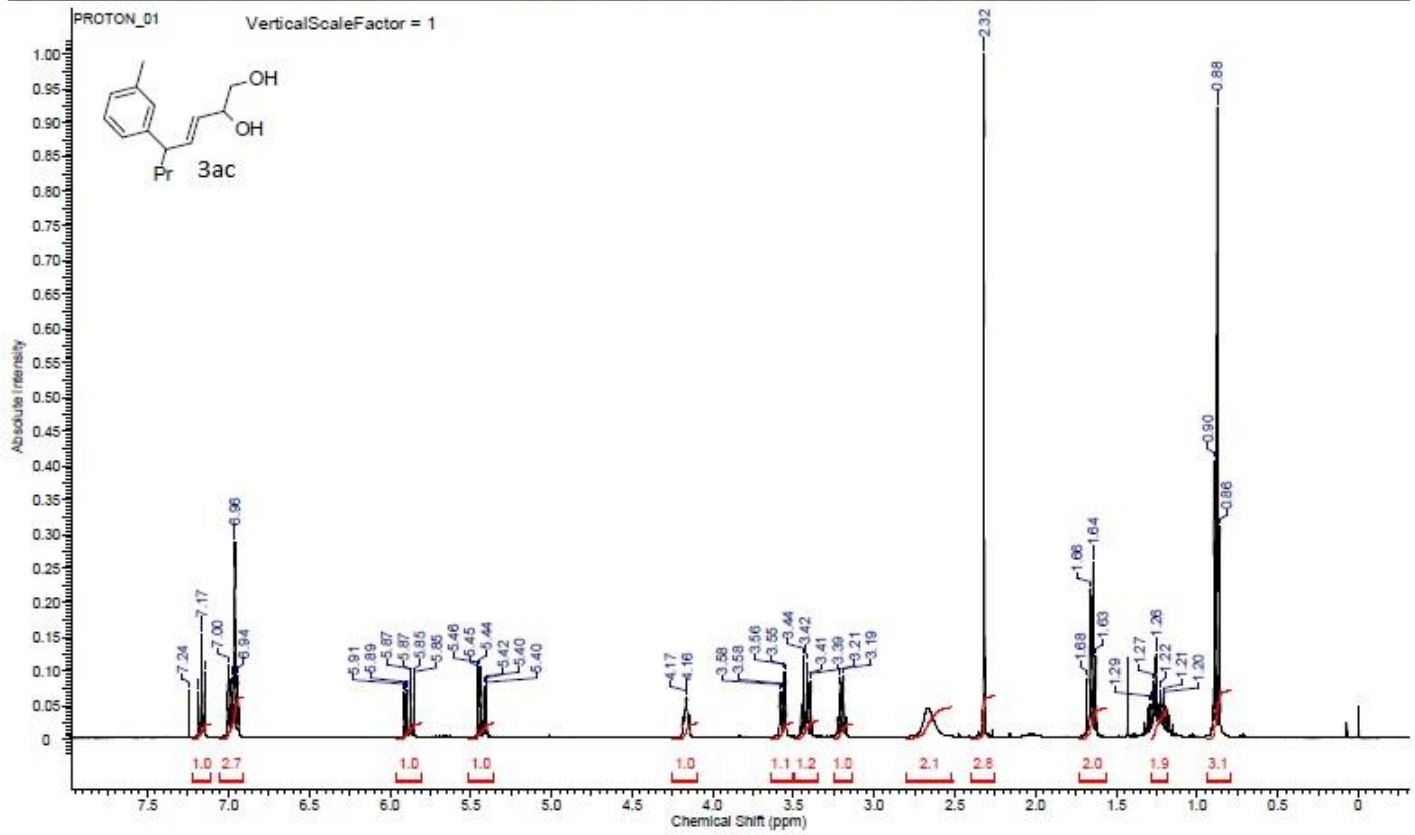
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
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				Temperature (degree C)	25.000



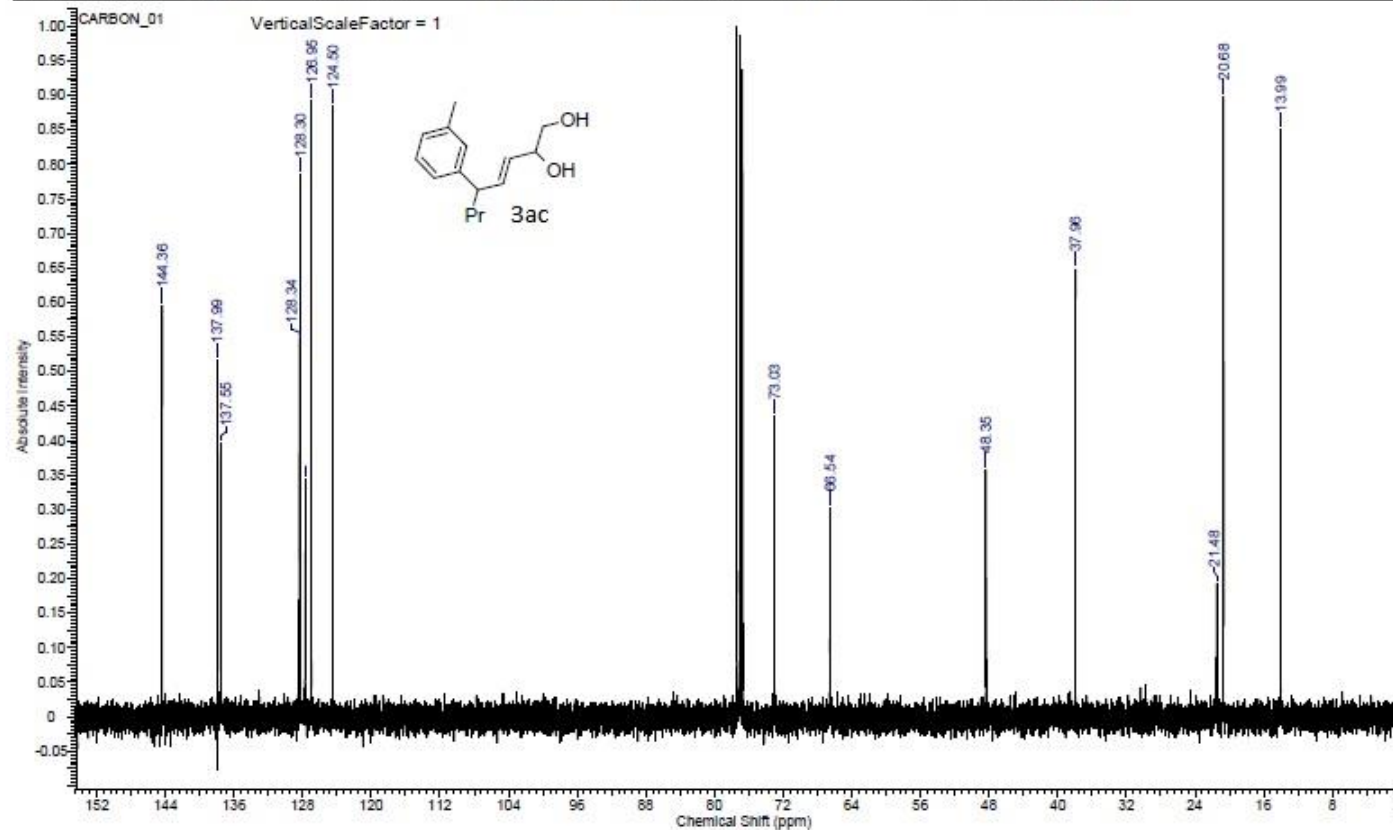
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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



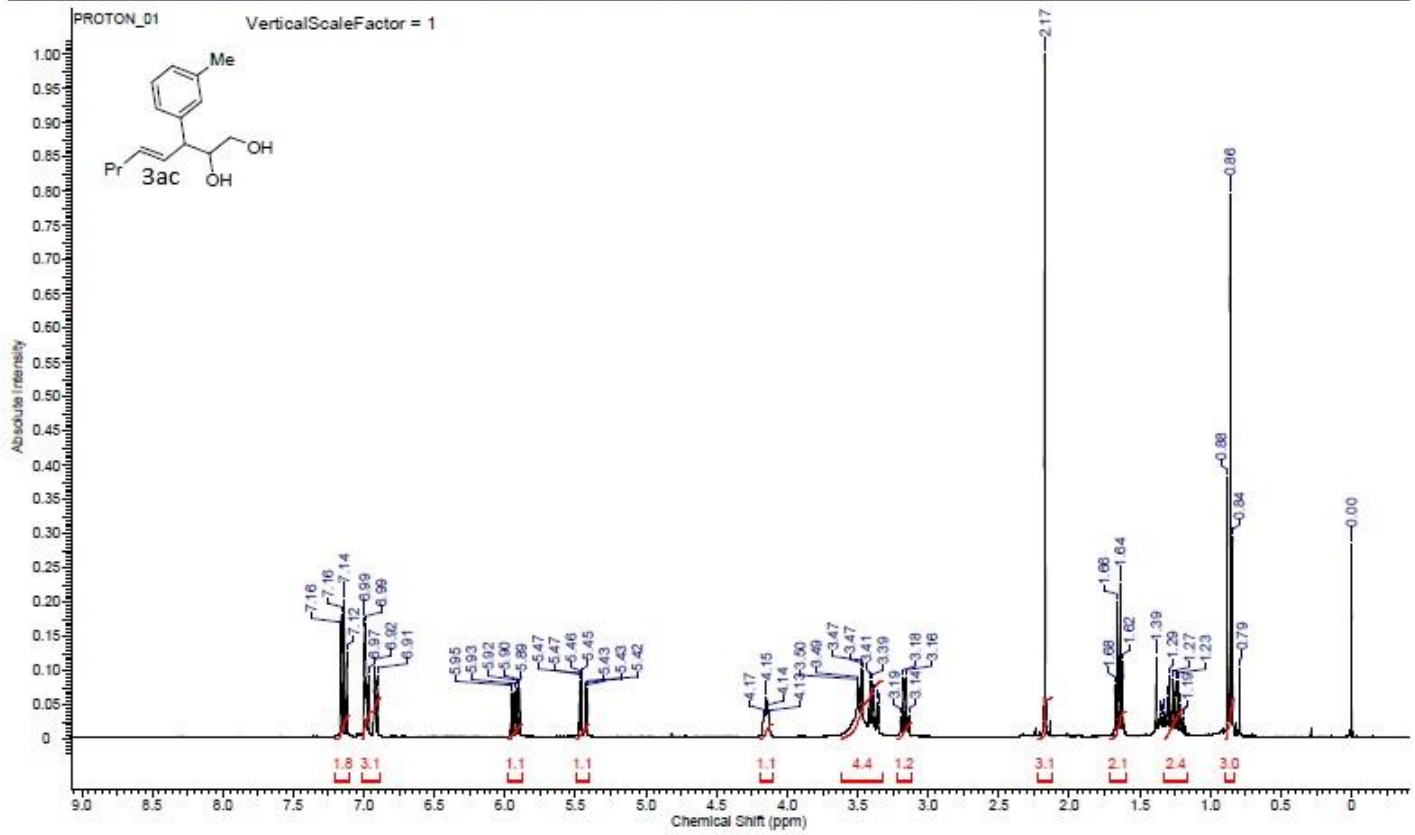
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Date Stamp	May 3 2016	File Name	F:\UNTITLED FOLDER\ERM-ENEP-212-1 3-CDCL3 20160503	01\PROTON_01.FID\FID	
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2nu1	Receiver Gain	50.00
Spectrum Offset (Hz)	2393.2329	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-212-1 3-CDCL3	Date	May 3 2016
Date Stamp	May 3 2016	File Name	F:\UNTITLED FOLDER\ERM-ENEP-212-1 3-CDCL3 20160503	01\CARBON_01.FID\FID	
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
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				Temperature (degree C)	25.000

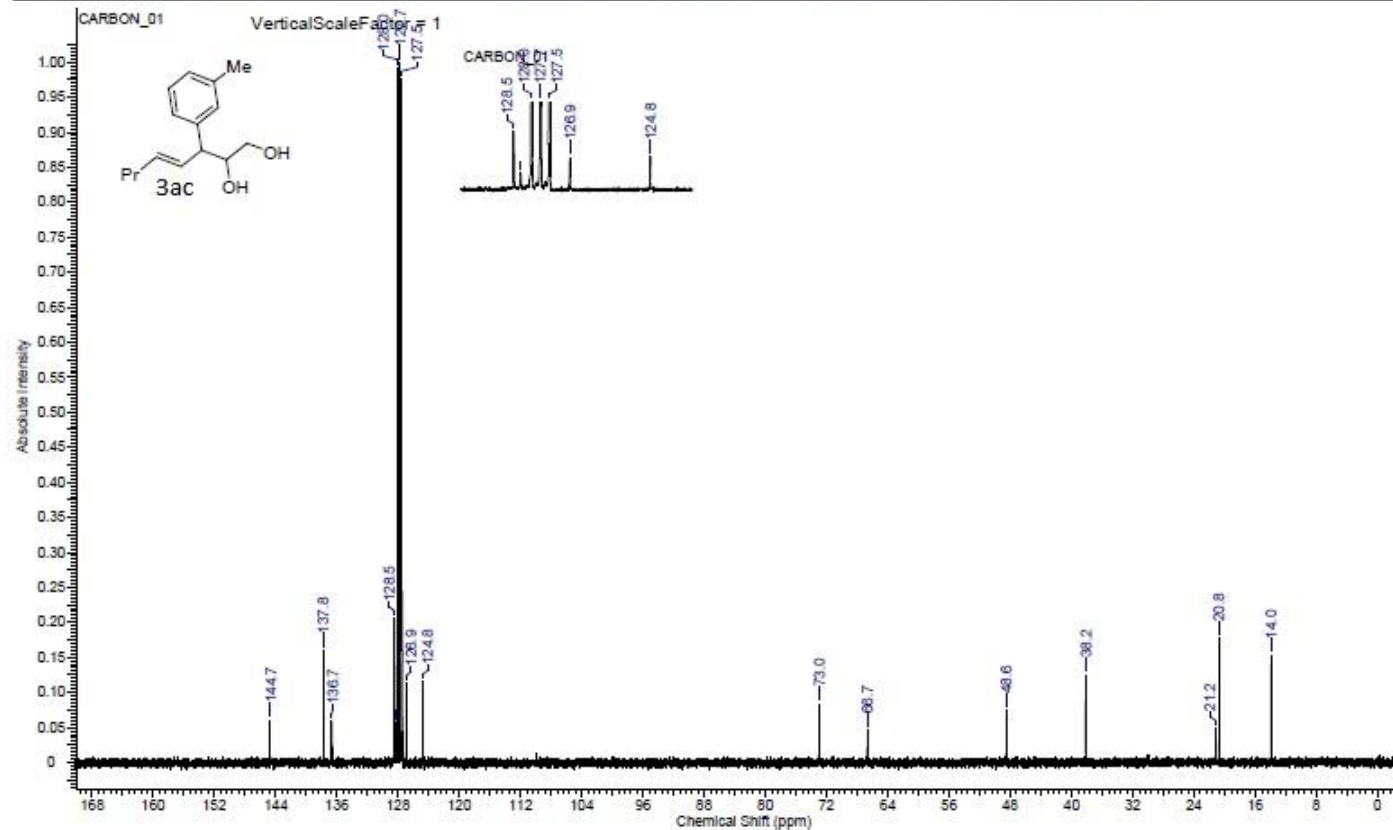


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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	50.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	2381.1033	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000

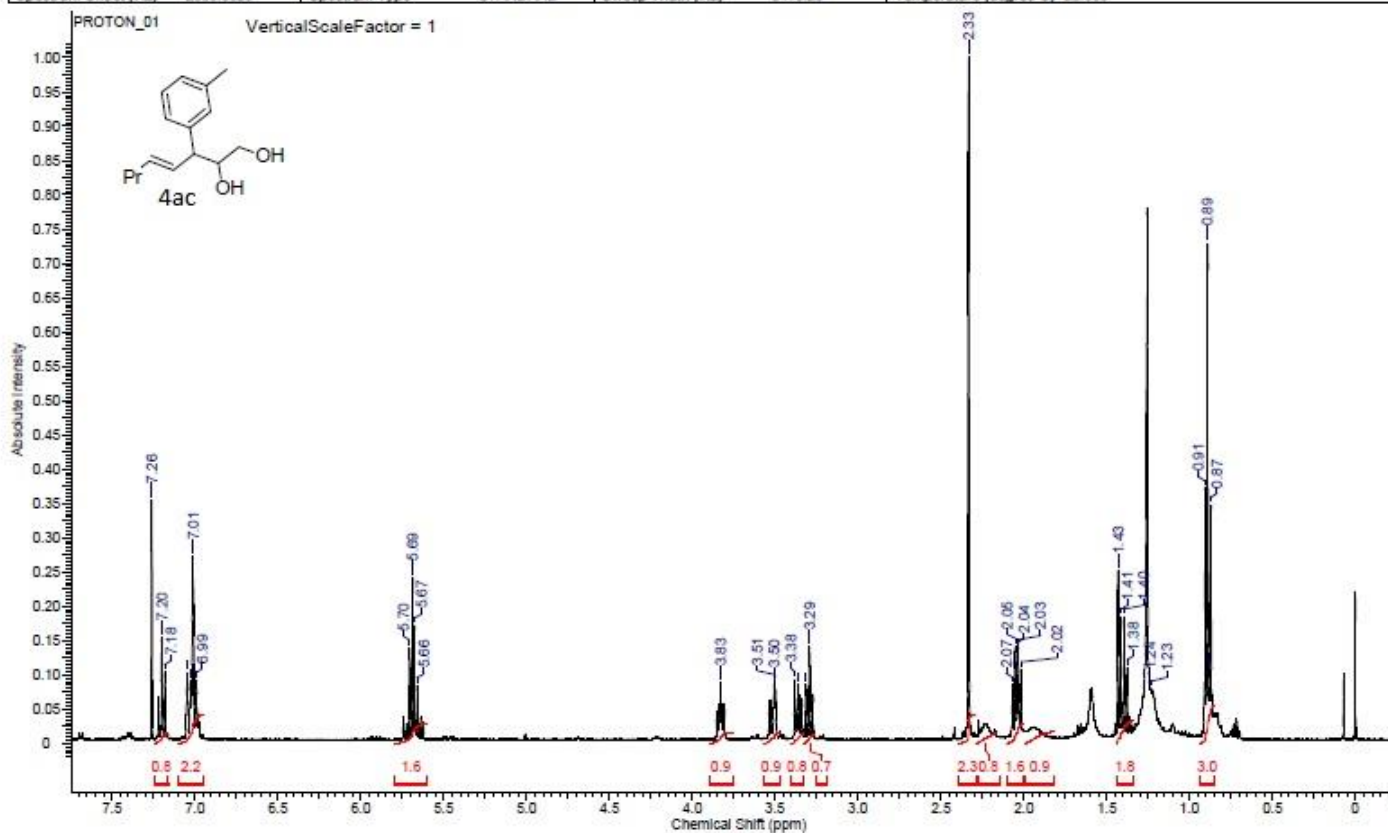


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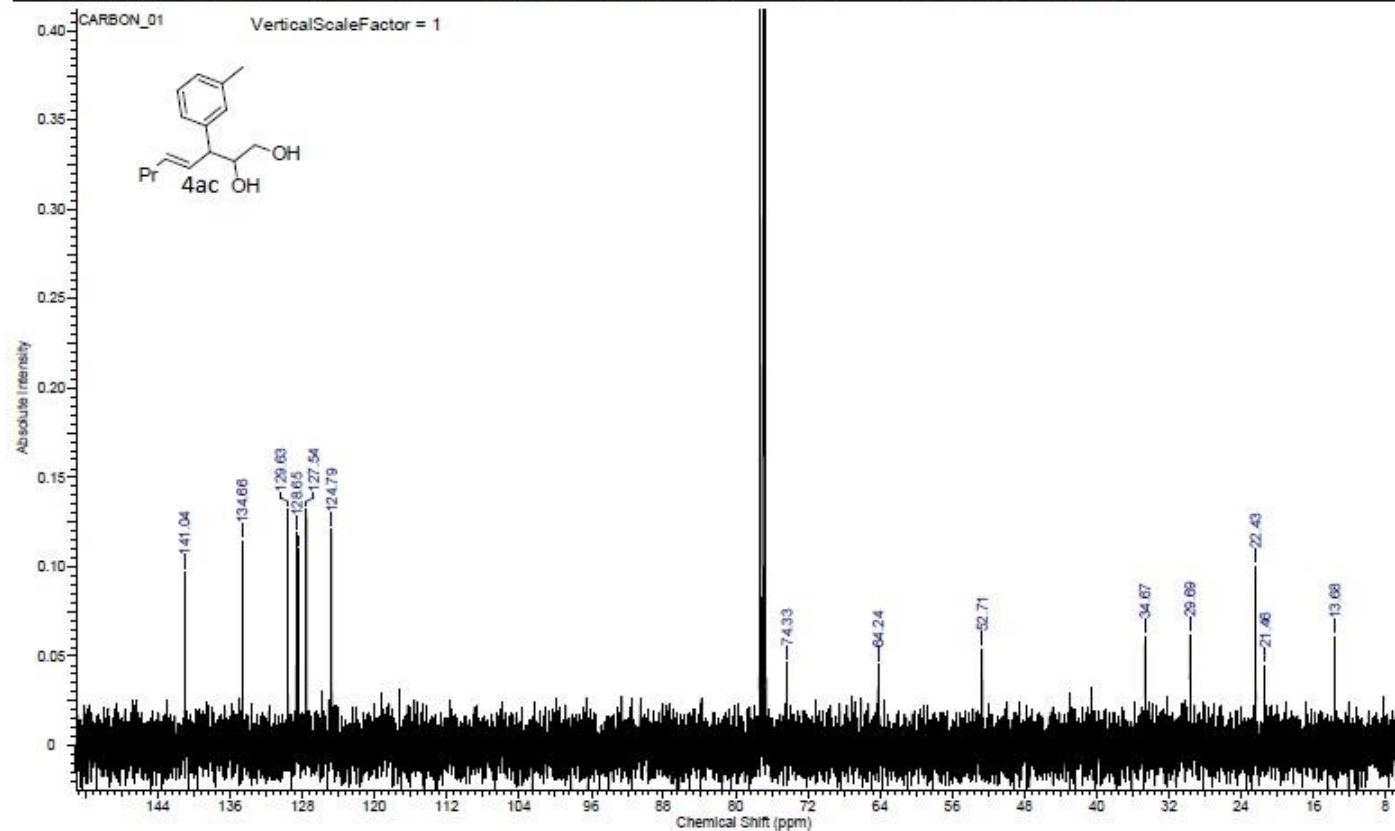
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Date Stamp	May 3 2016	File Name	C:\USERS\ERMAN\DESKTOP\UNTITLED FOLDER\ERM-ENEP-212-1-3-BNZ_20160503_01\CARBON_01.FID\FID		
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2nu1	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
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				Temperature (degree C)	25.000



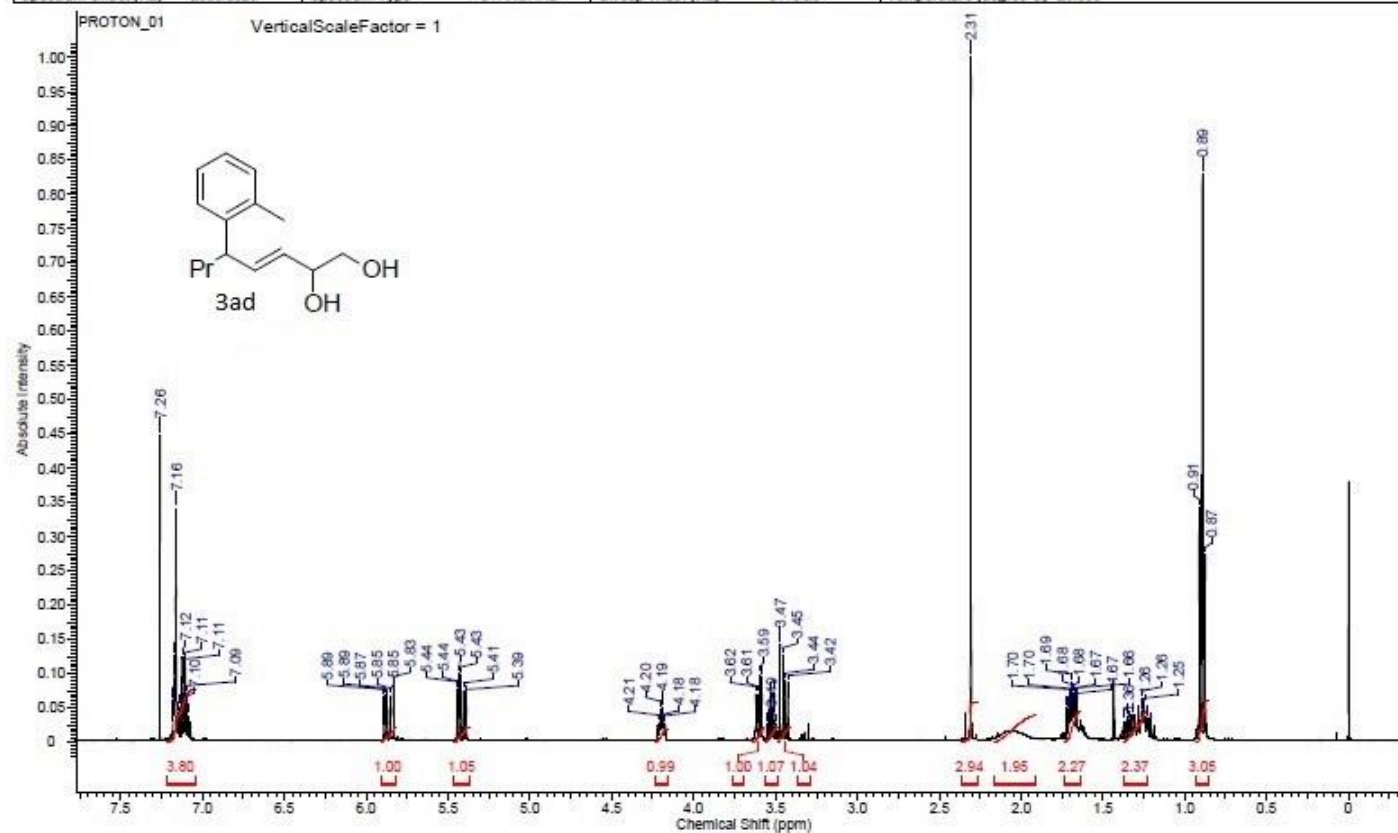
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Date Stamp	May 3 2016	File Name	F:\UNTITLED FOLDER\ERM-ENEP-212-1 1-CDCL3 20160503	01\PROTON_01.FID\FID	
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2pu1	Receiver Gain	60.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



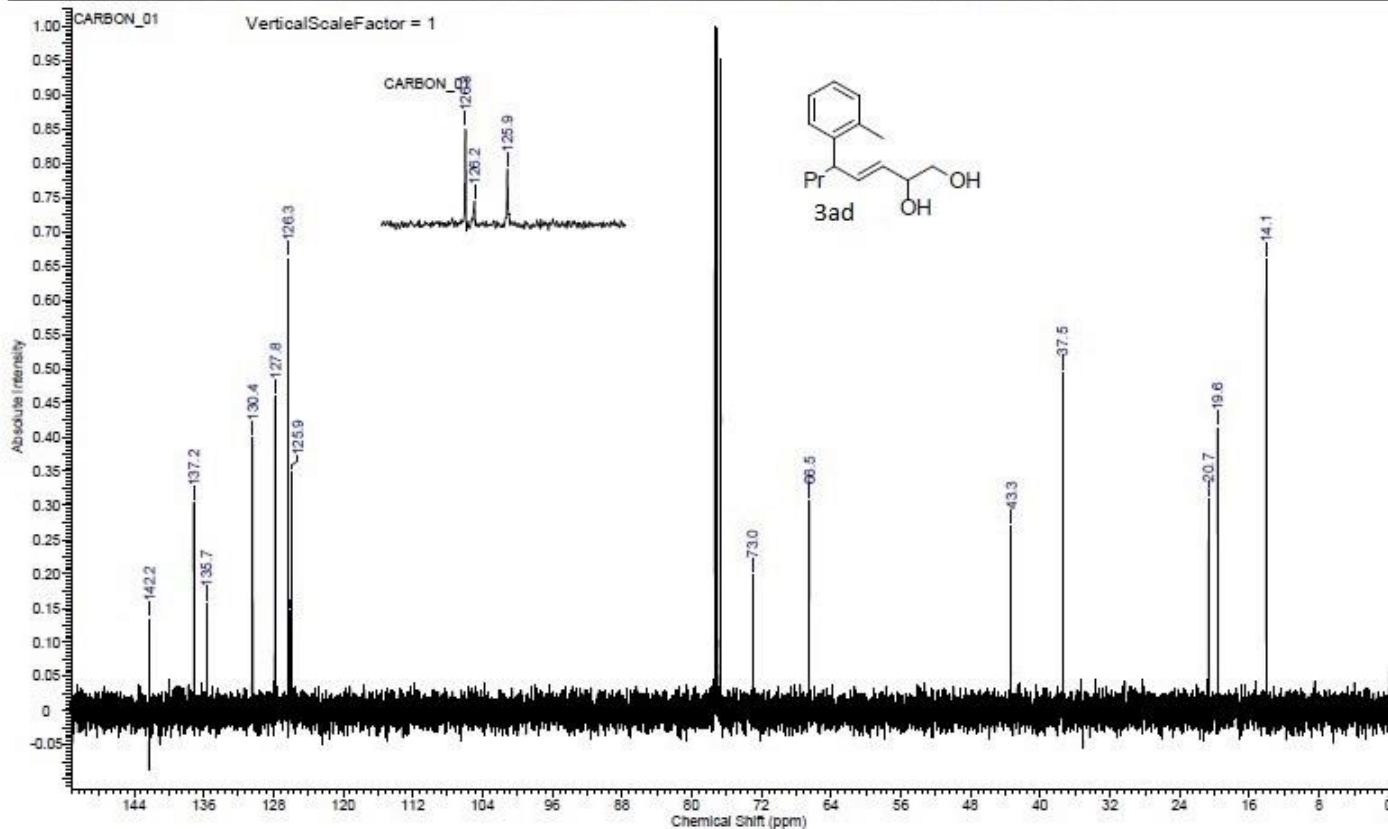
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Date Stamp	May 3 2016	File Name	F:\UNTITLED FOLDER\ERM-ENEP-212-1 1-CDCL3 20160503	01\CARBON_01.FID\FID	
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	512
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



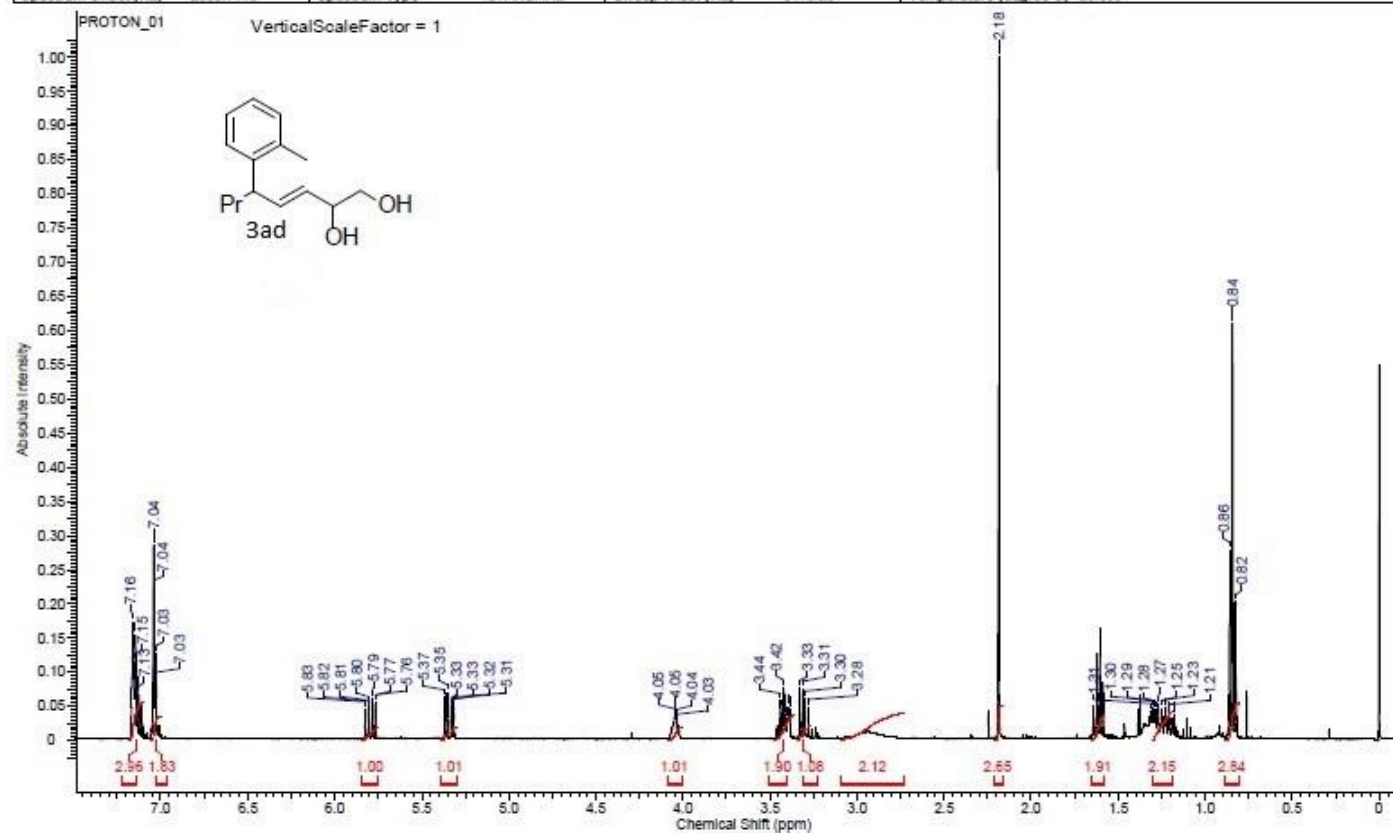
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-193-1 3-CDCL3		Date	Mar 30 2016	
Date Stamp	Mar 30 2016	File Name	D:\YENI NMR\ERM-ENEP-193-1 3-CDCL3 20160330_01\PROTON_01.FID\FID				
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



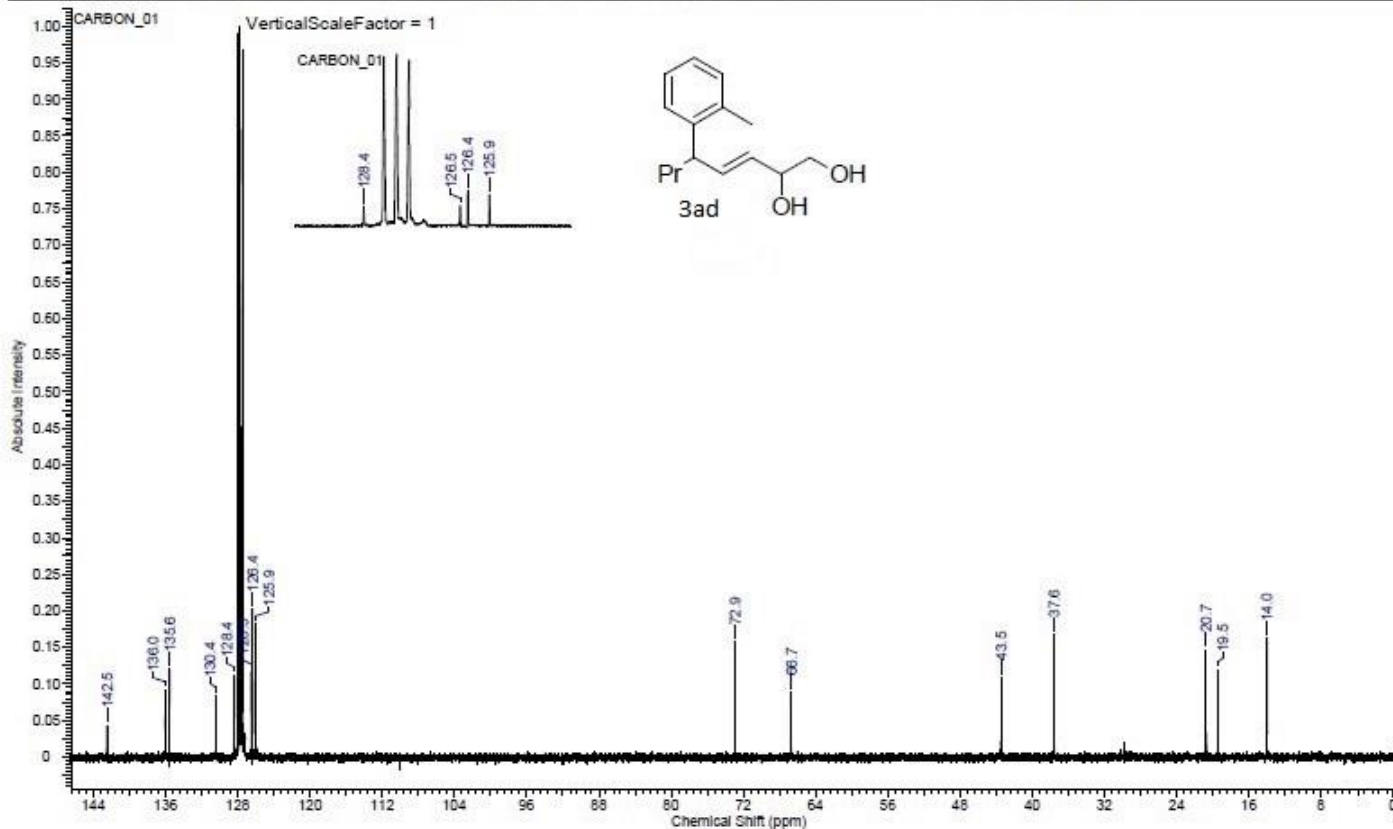
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Date Stamp	Mar 30 2016	File Name	D:\YENI NMR\ERM-ENEP-193-1 3-CDCL3 20160330 01\CARBON_01.FID\FID		
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2nuJ	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



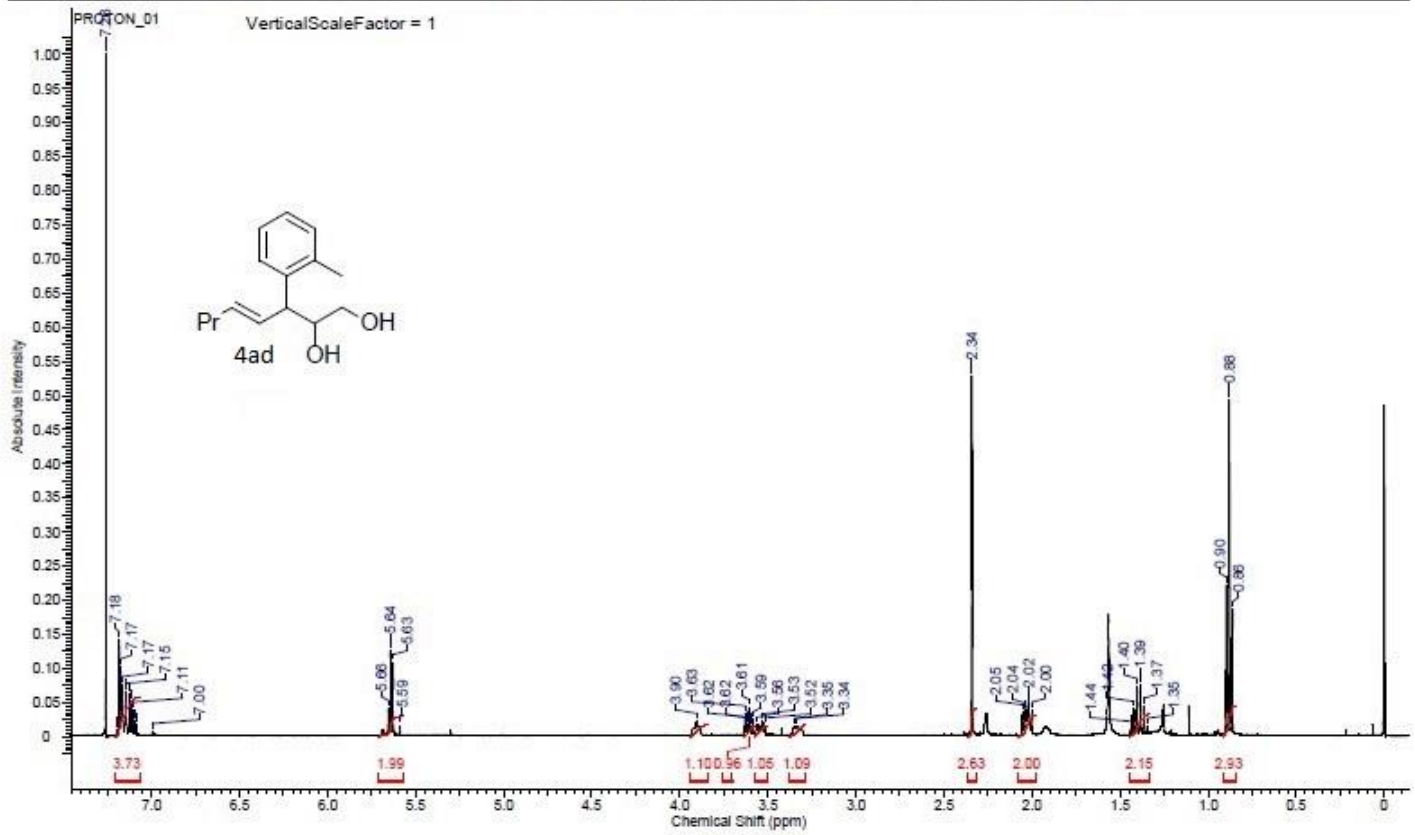
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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	50.00
Spectrum Offset (Hz)	2380.7119	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



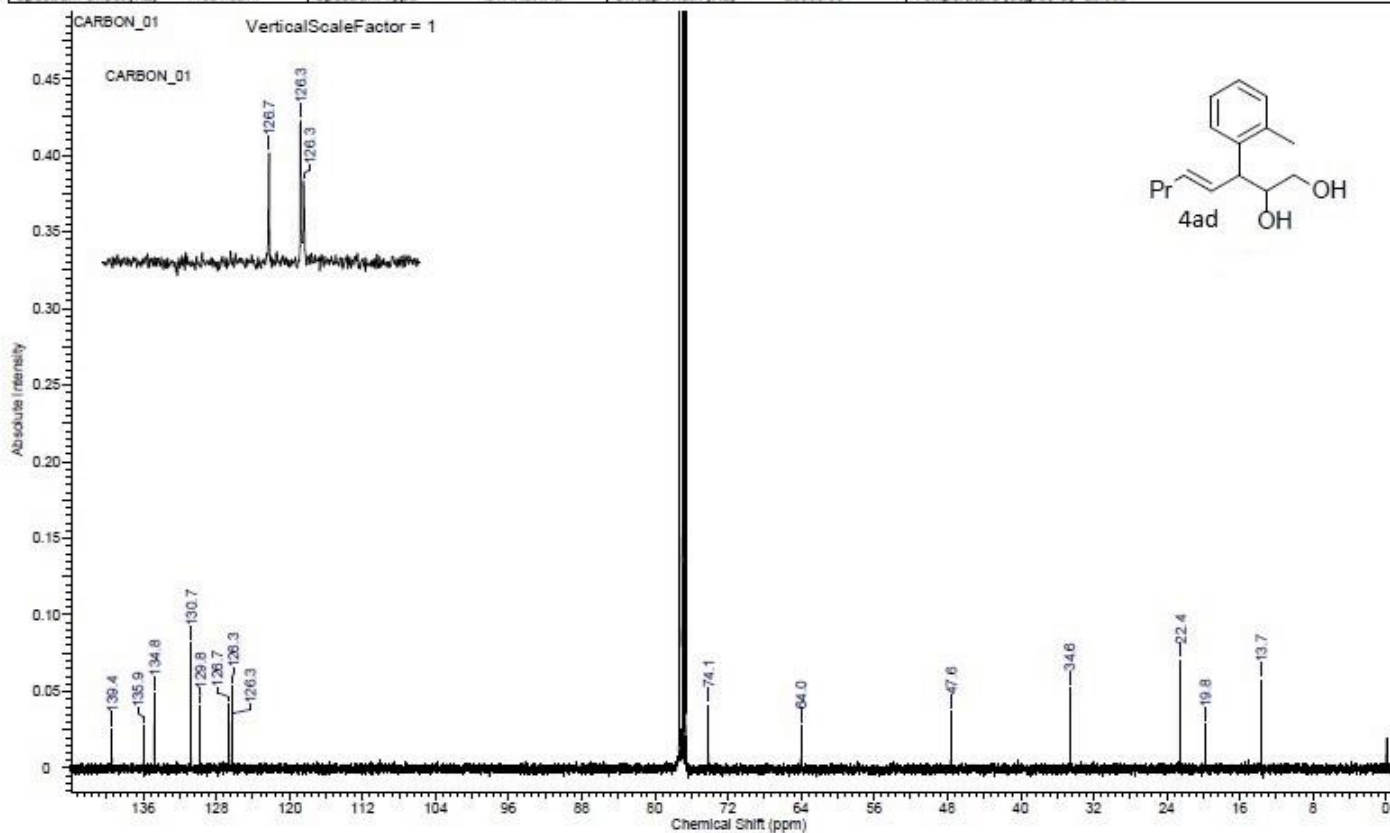
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



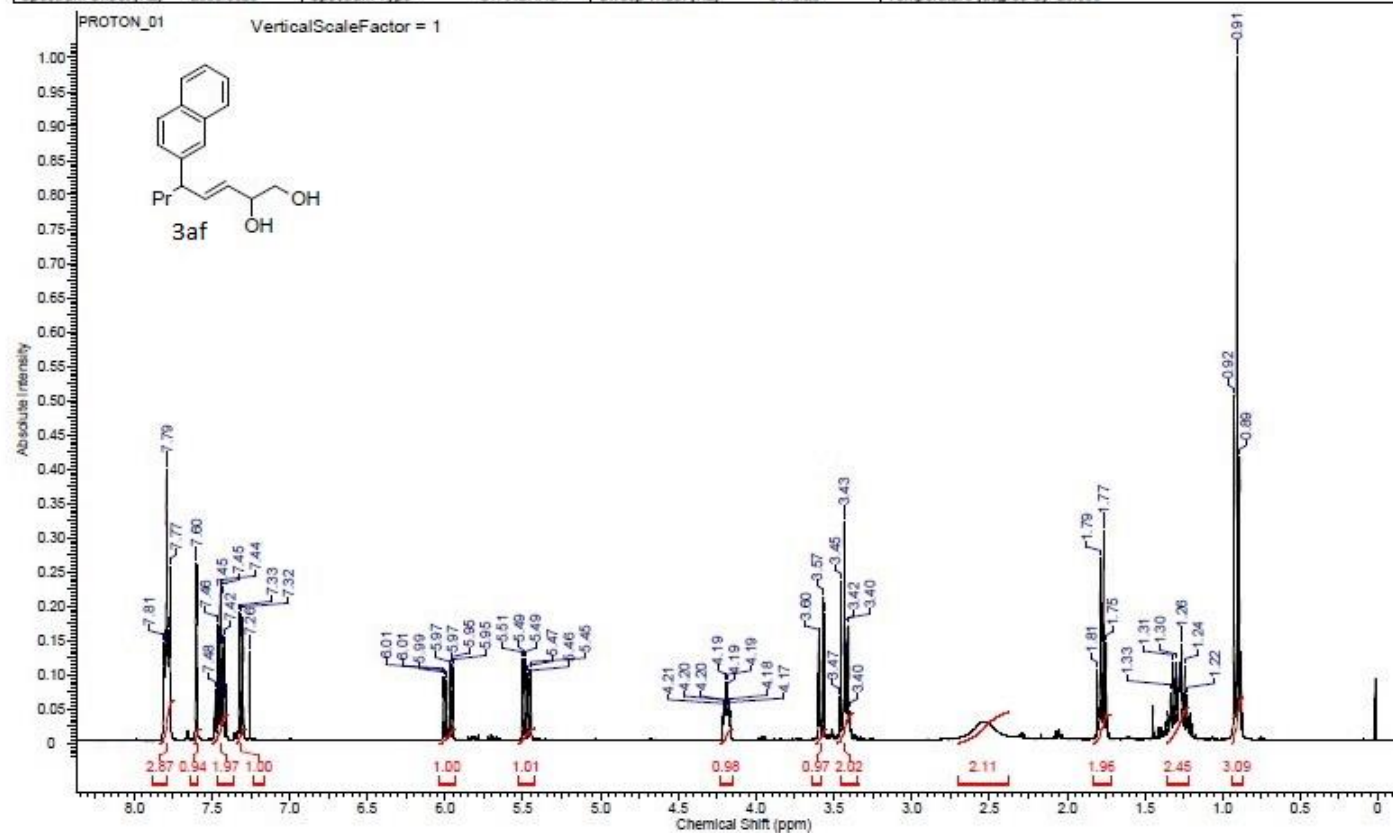
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Points Count	16384	Pulse Sequence	s2bul	Receiver Gain	60.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



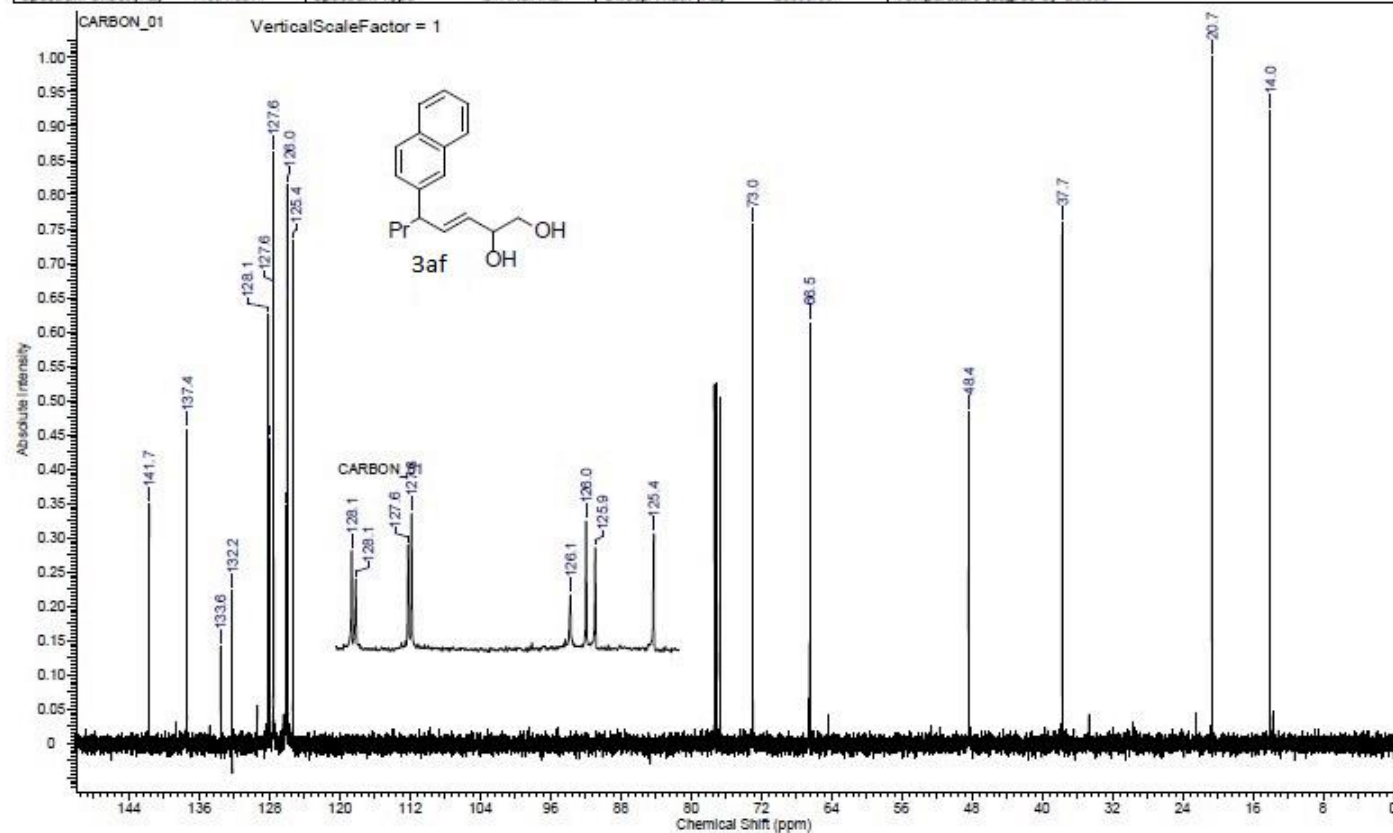
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	5000	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2p1	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



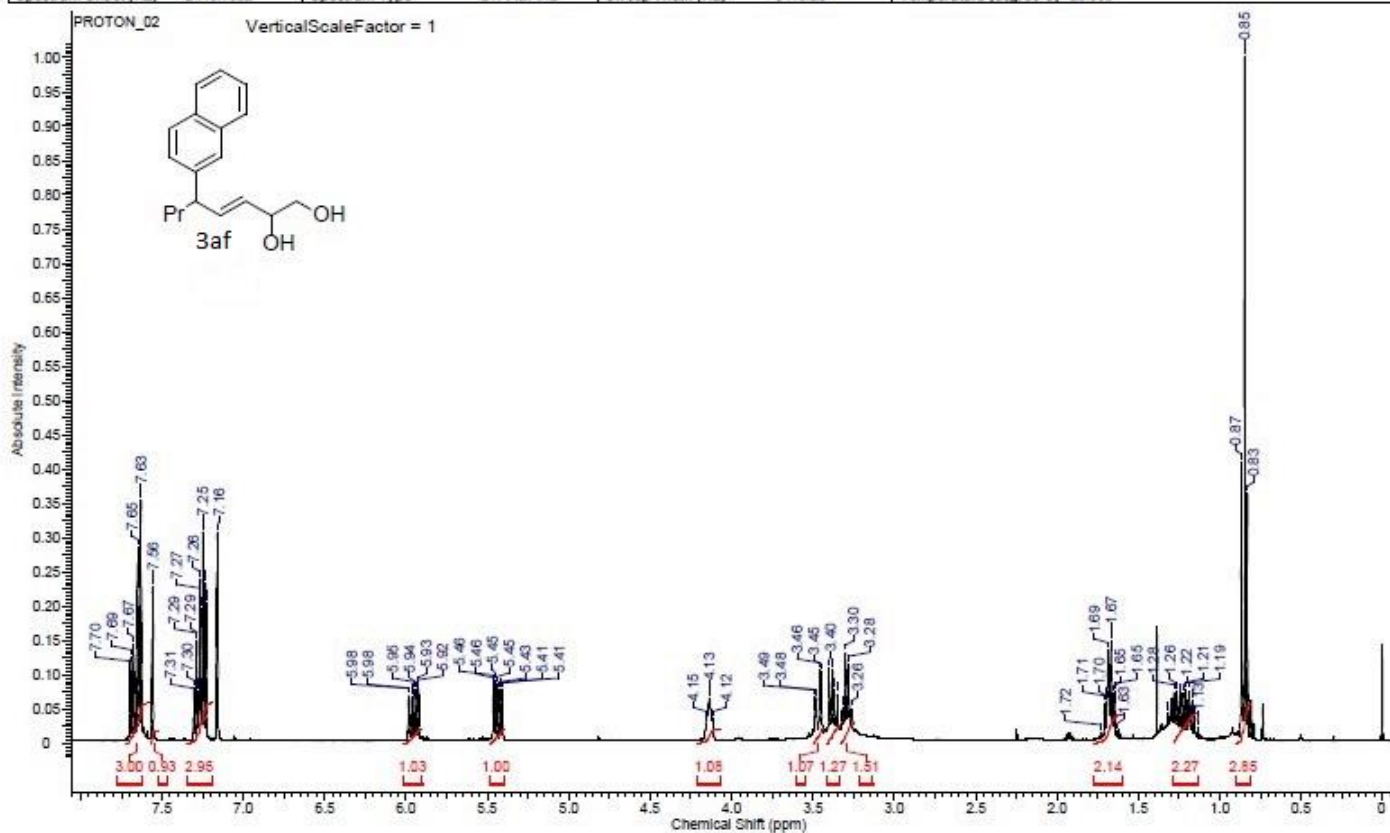
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Date Stamp	Mar 30 2016	File Name	D:\YENI NMR\ERM-ENEP-200-1 3-CDCL3 20160330 01\PROTON_01.FID\FID				
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	54.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



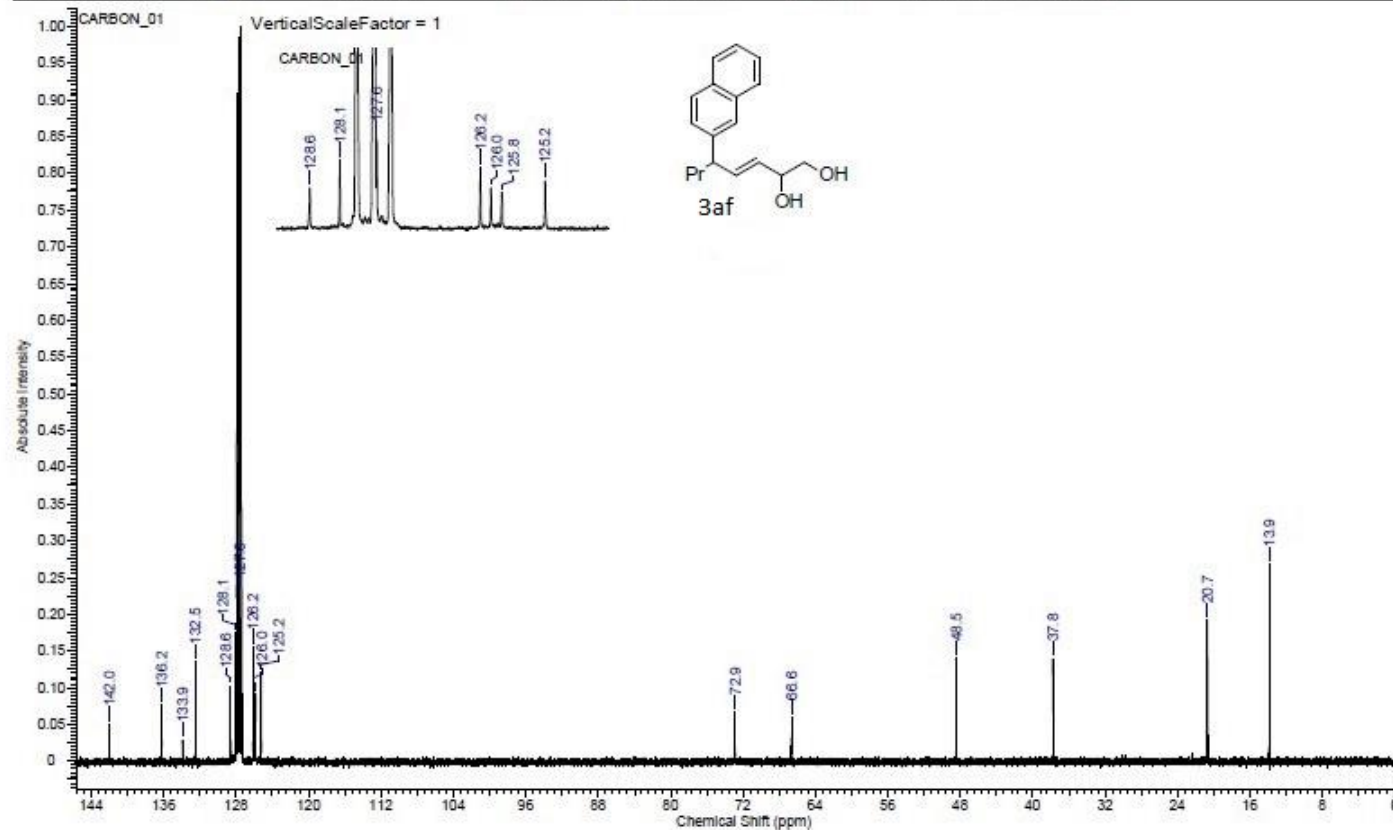
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2nuJ	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
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				Temperature (degree C)	25.000



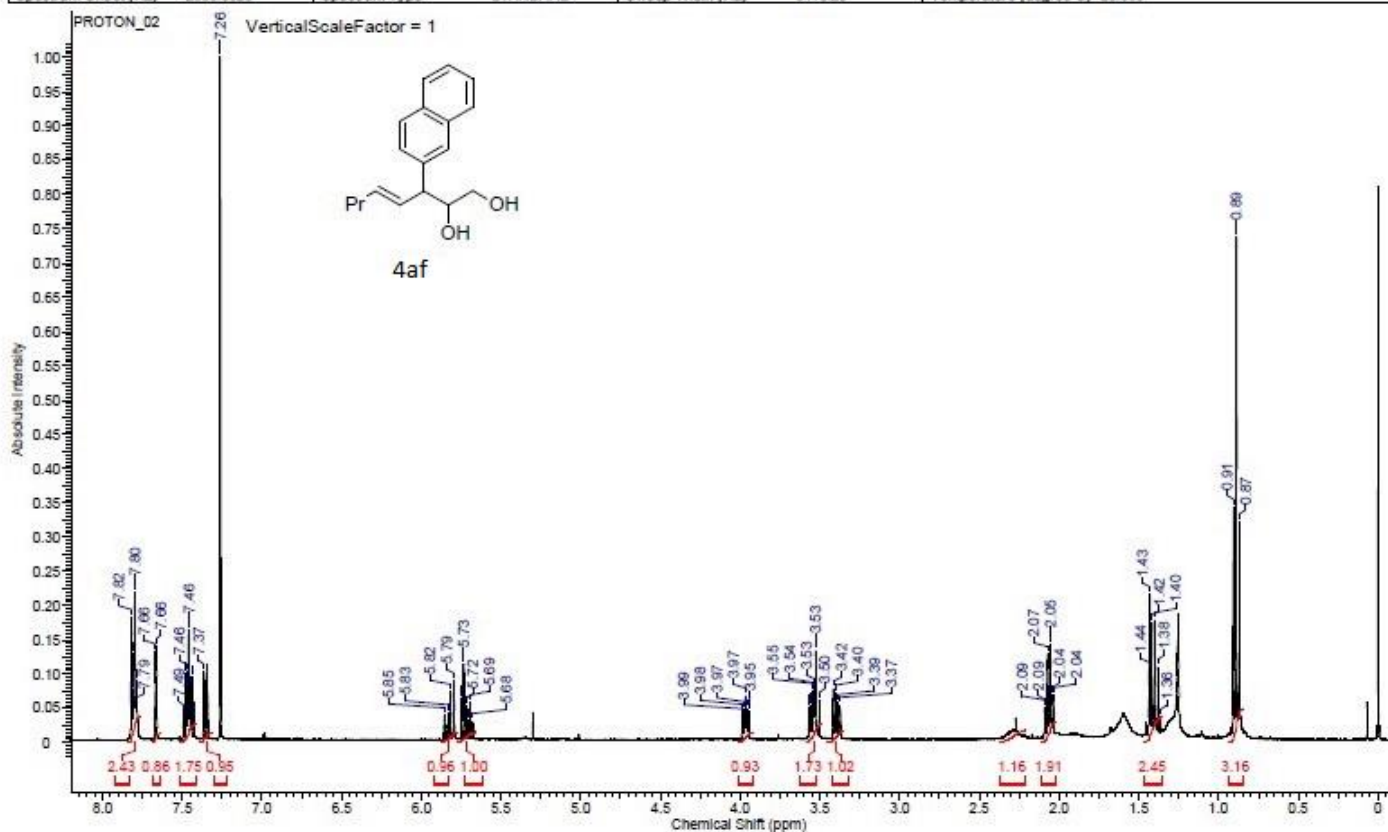
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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	64
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	52.00
Spectrum Offset (Hz)	2416.7092	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



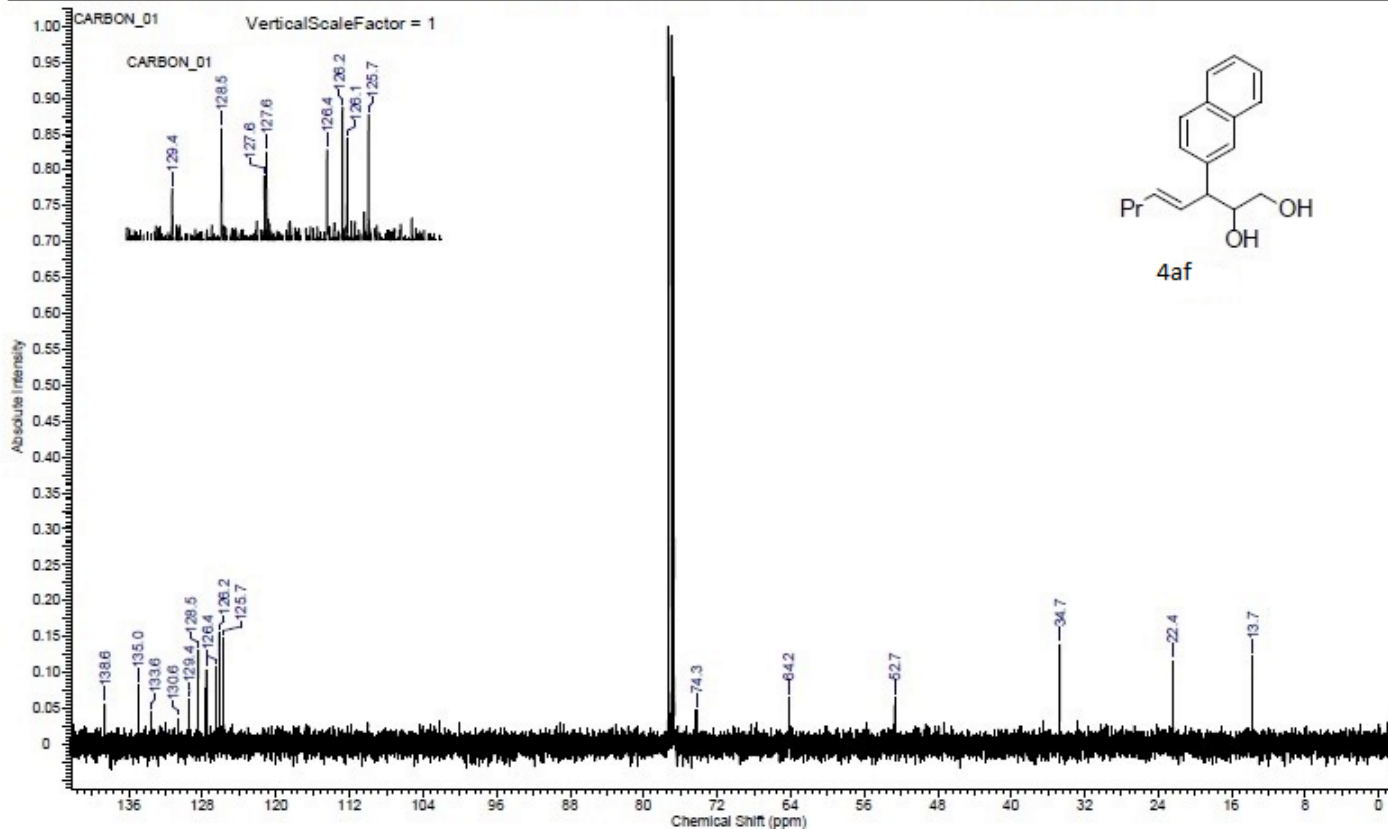
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



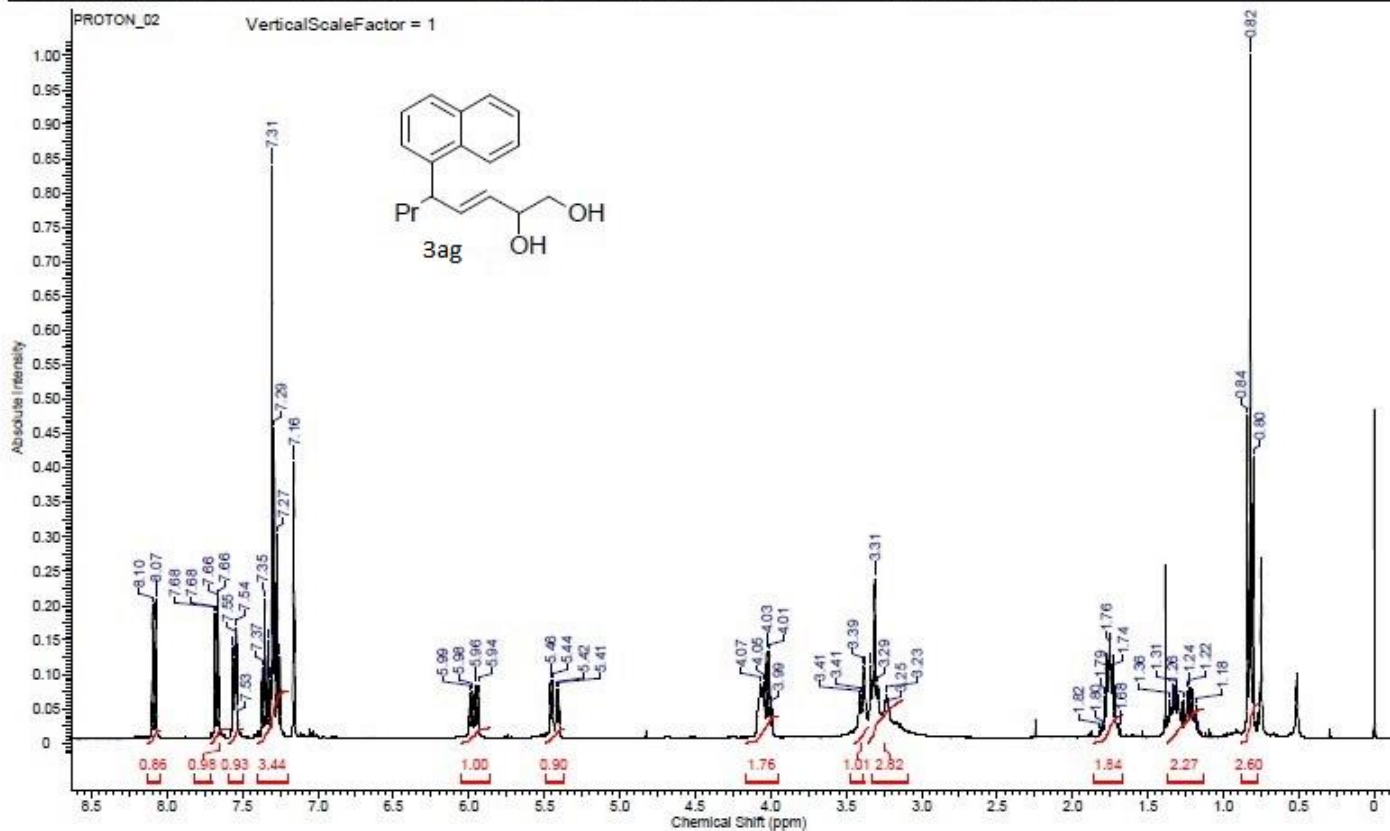
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Date Stamp	Feb 24 2016	File Name	C:\USERS\IERMAN\GOOGLE DRIVE\IERMAN\ERM-ENEP-200-1 1-DOGRU 20160224_01\PROTON_02.FID\FID	Original Points Count	16384
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	32
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Temperature (degree C)	25.00



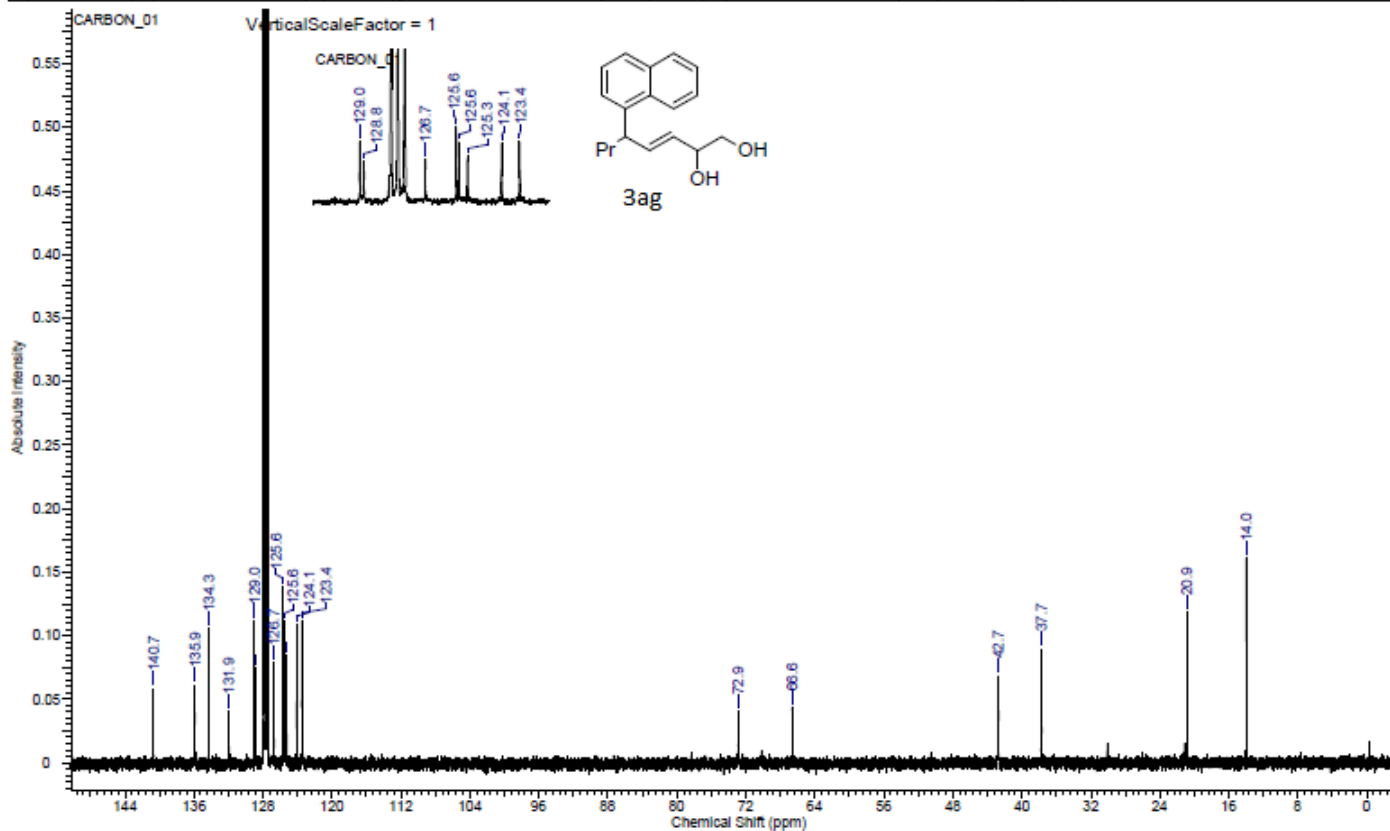
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	500	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



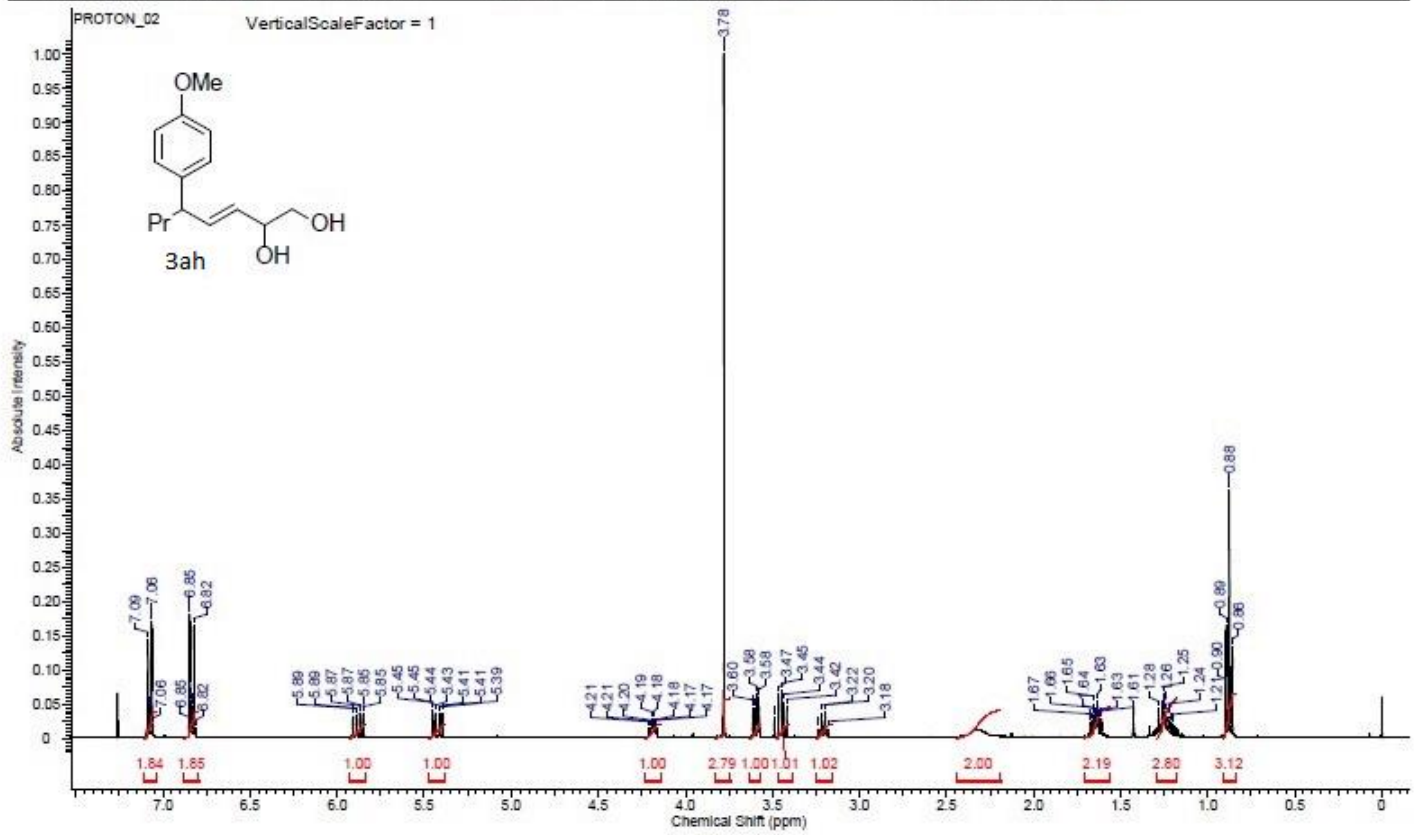
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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	32
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	52.00
Spectrum Offset (Hz)	2416.3181	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



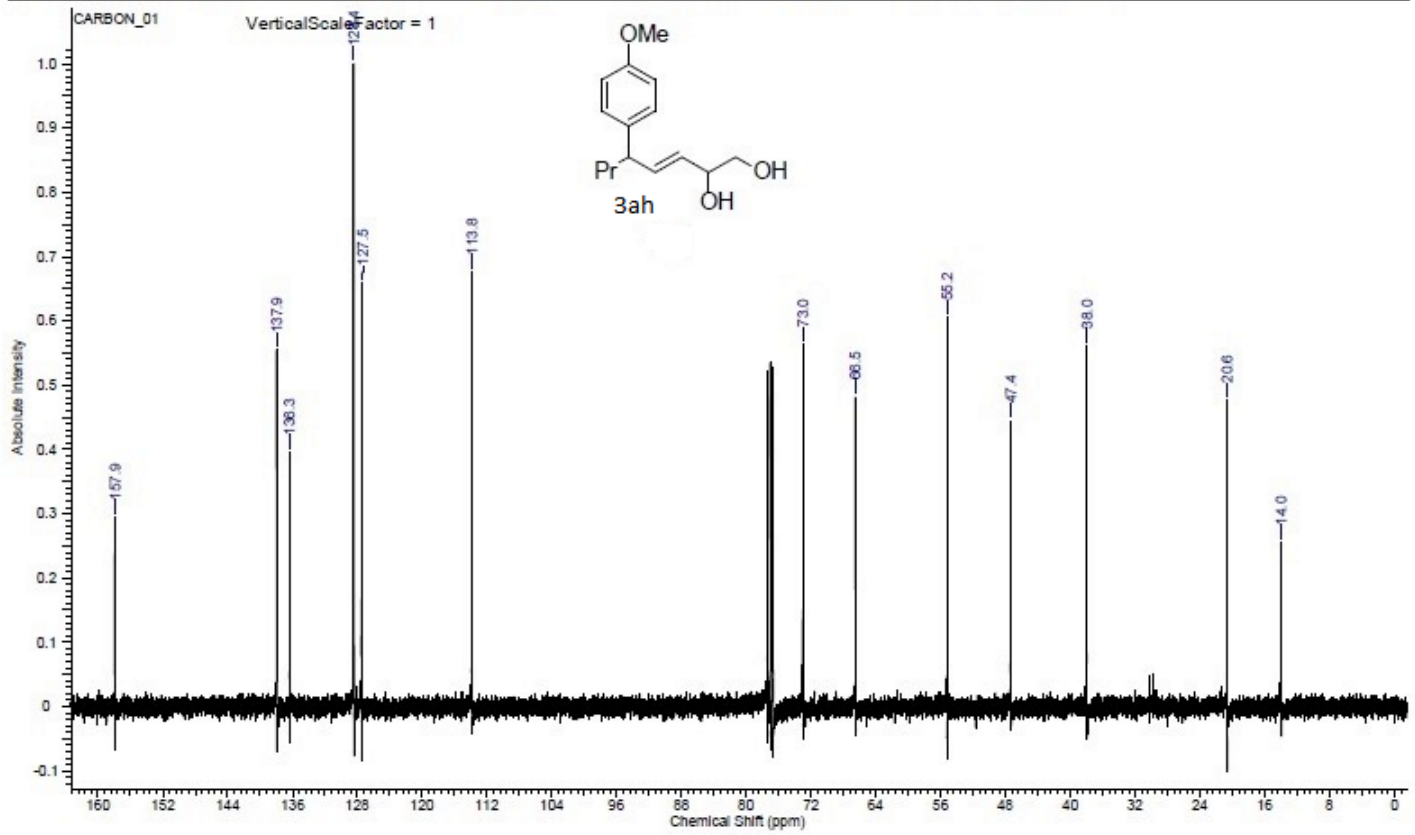
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



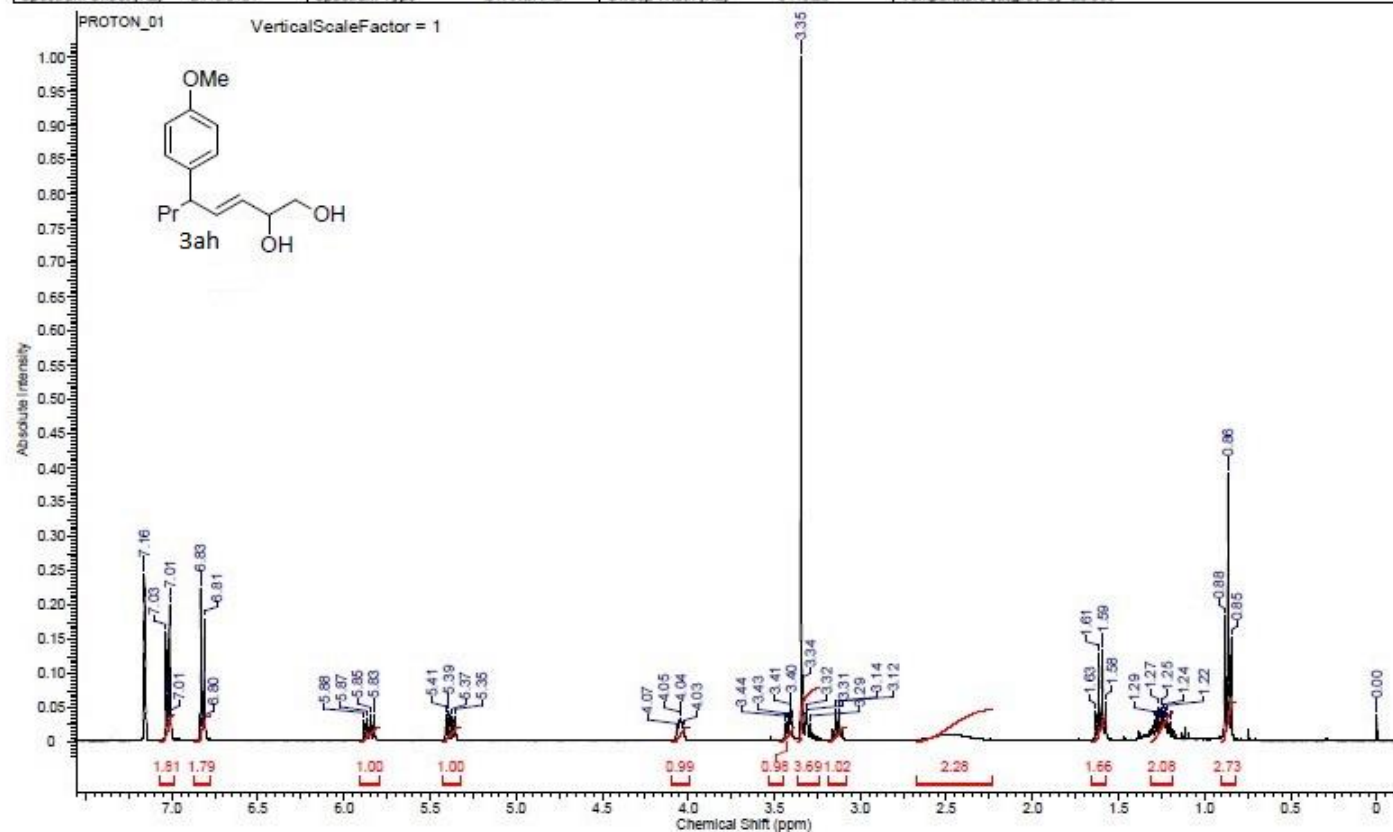
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Date Stamp	Feb 10 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-191-1	3.20160210_01\PROTON_02.FID\FID	
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2oul	Receiver Gain	54.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Temperature (degree C)	25.000



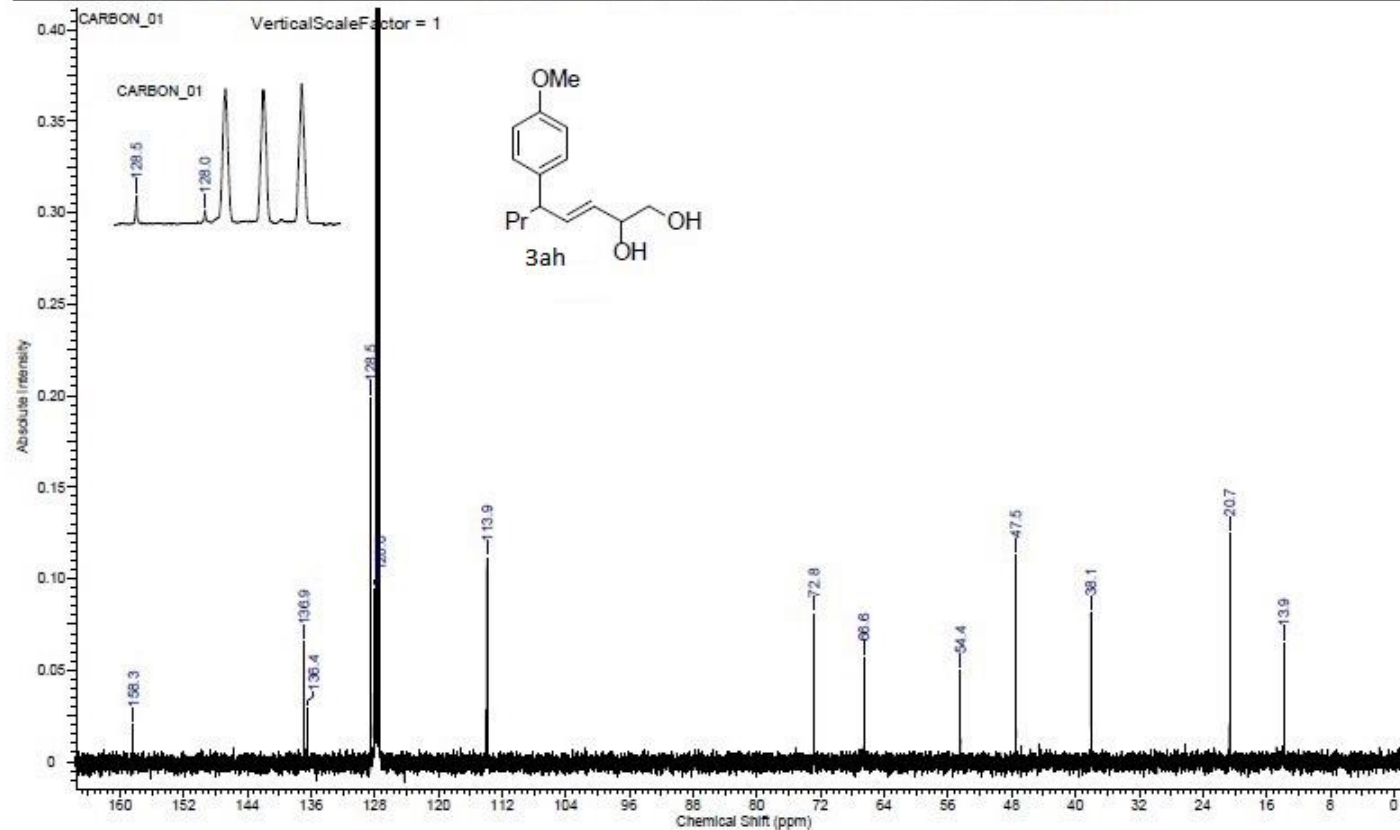
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



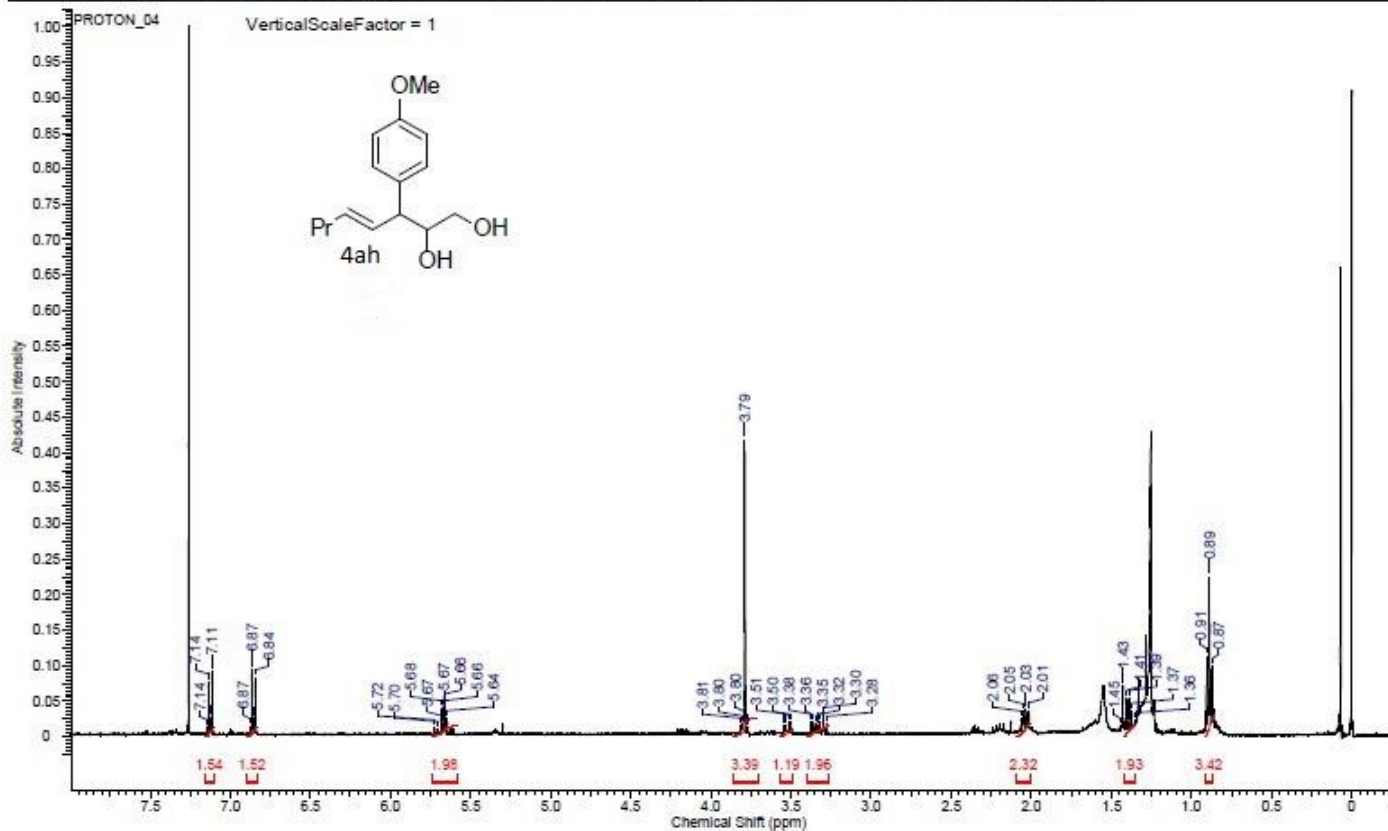
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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2p1d	Receiver Gain	56.00
Spectrum Offset (Hz)	2416.3181	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



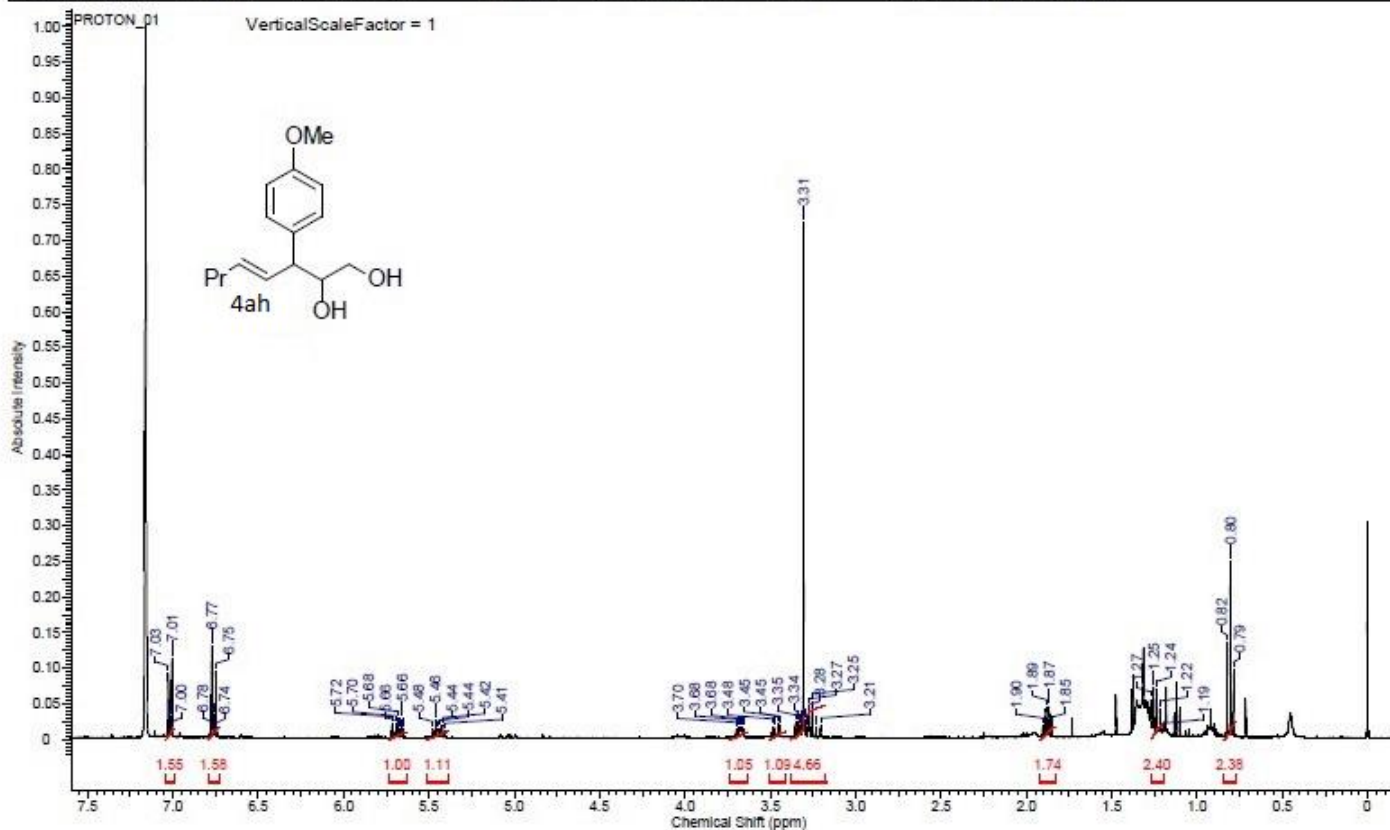
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



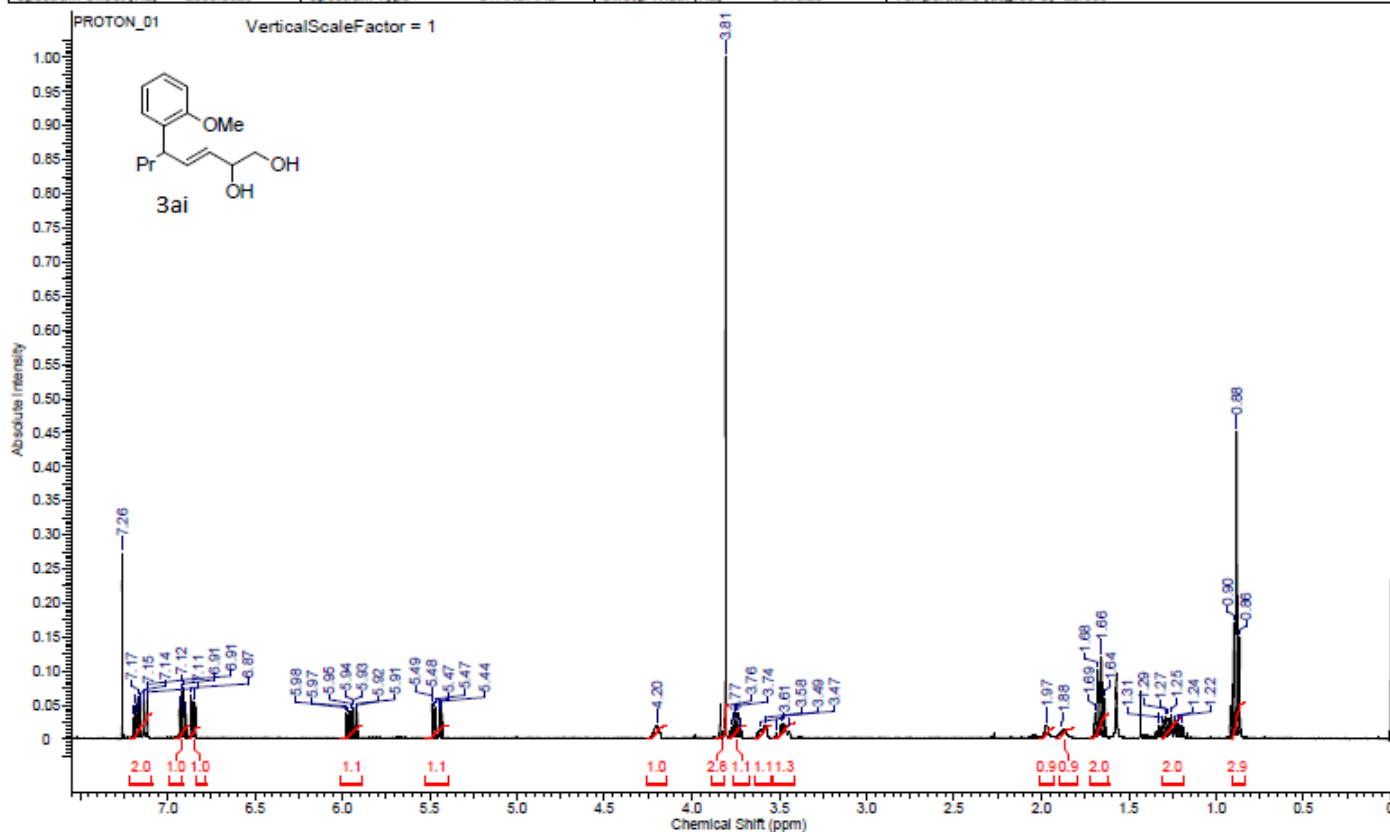
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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2oul	Receiver Gain	60.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-191-1_1		Date	Feb. 2.2016	
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Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	32	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	2416.7764	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000

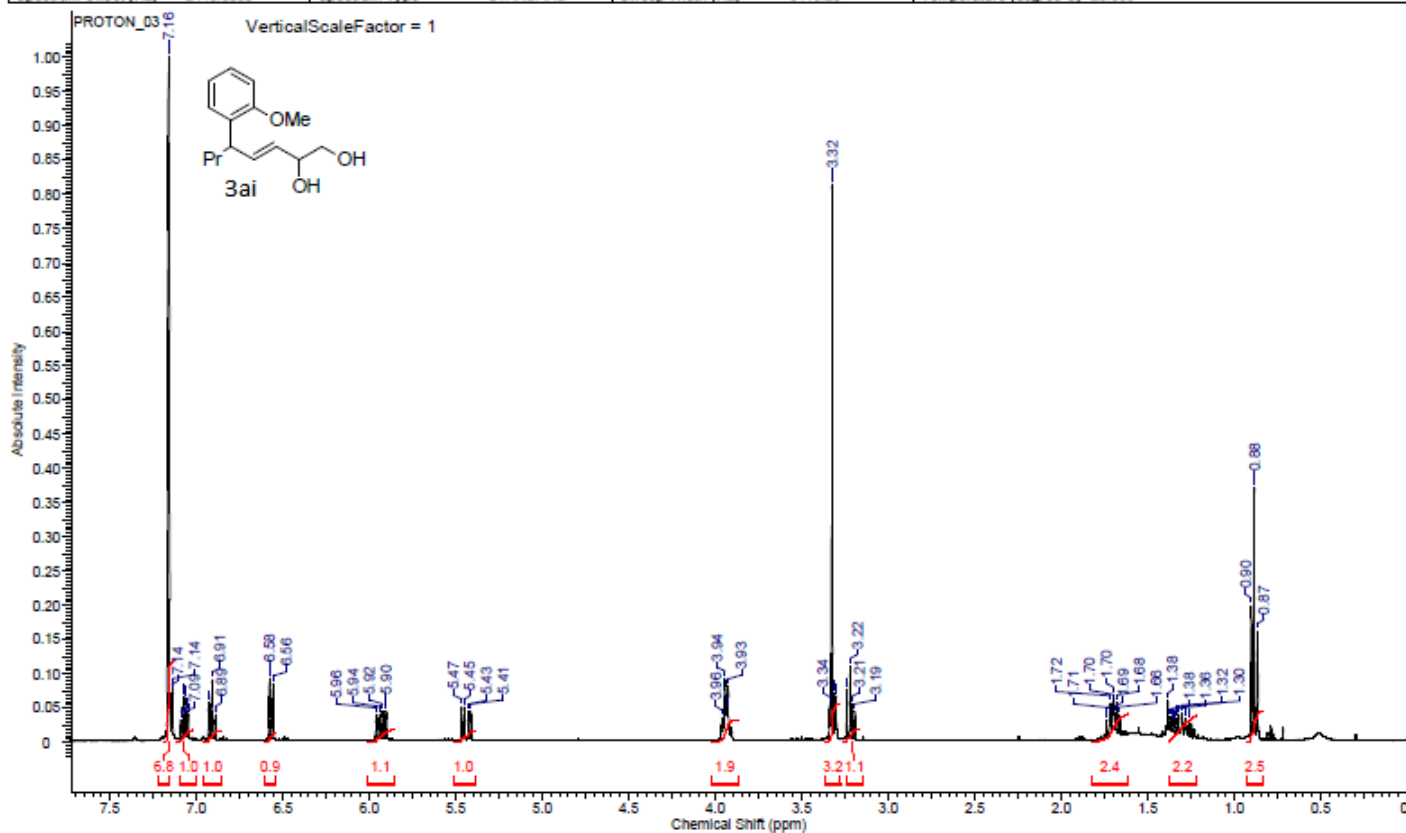


Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-210-1 3-CDCL3	Date	Apr 22 2016
Date Stamp	Apr 22 2016	File Name	F:\UNTITLED FOLDER\ERM-ENEP-210-1 3-CDCL3 20160422	01\PROTON_01.FID\FID	
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2nu1	Receiver Gain	60.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000

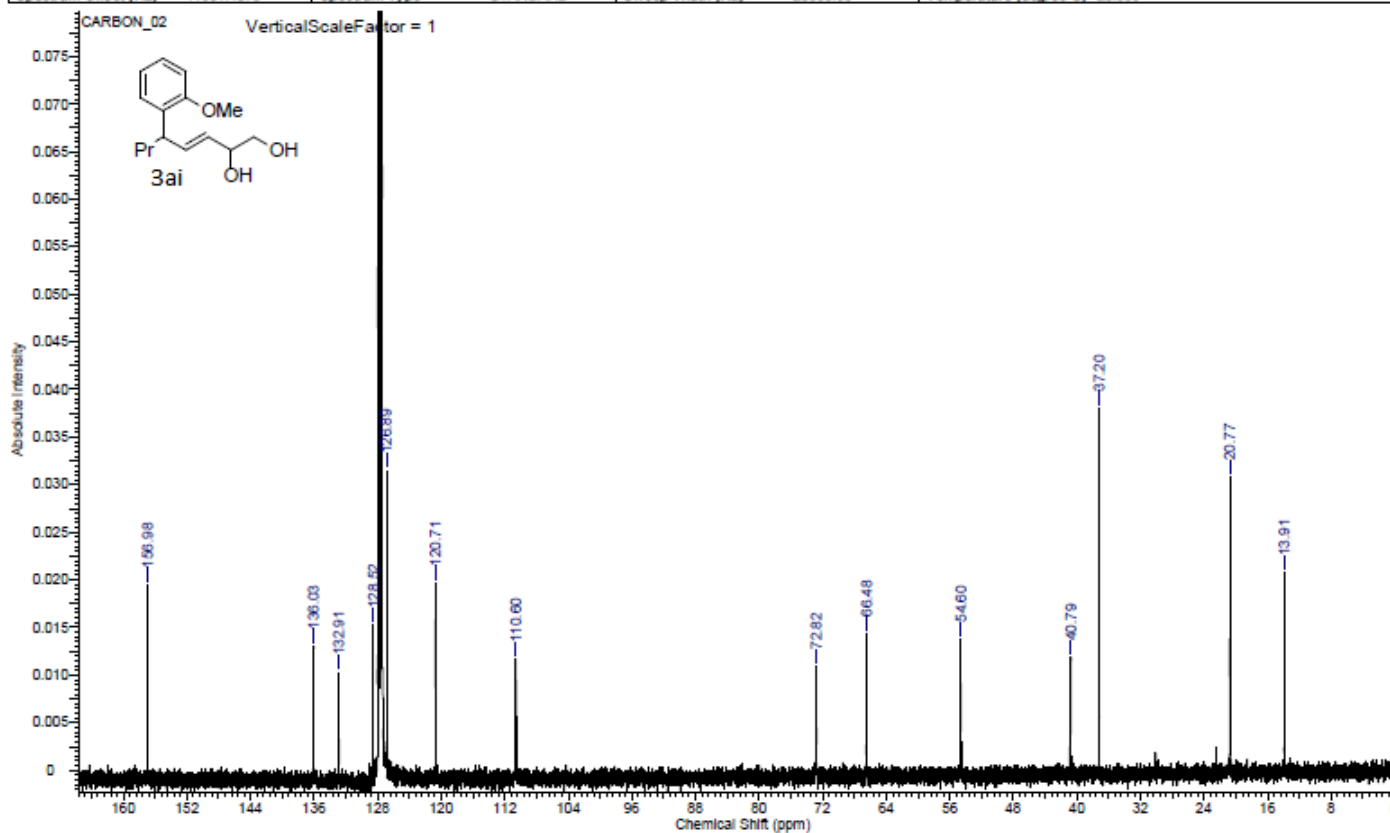


This report was created by ACD/NMR Processor Academic Edition. For more information go to www.acdlabs.com/nmrproc/

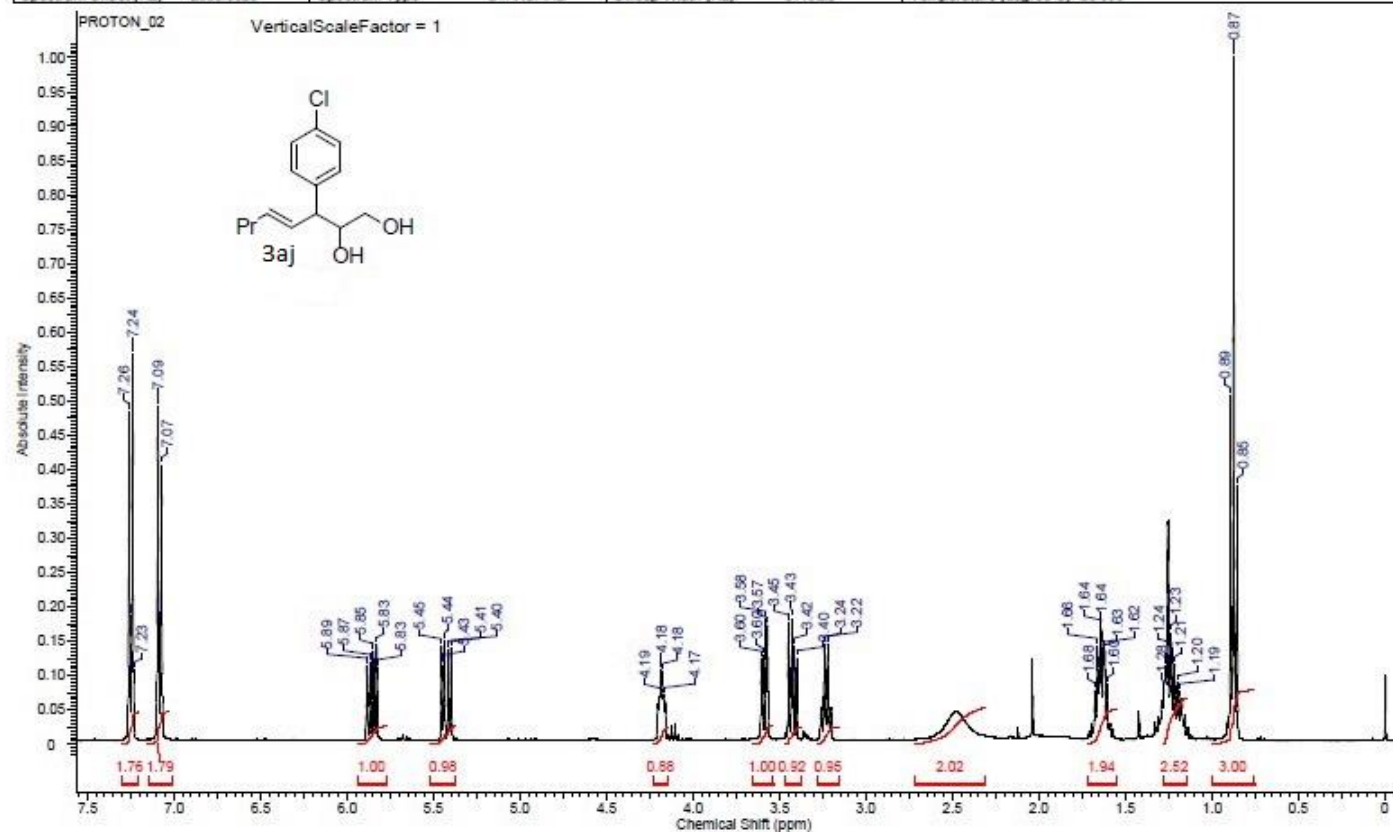
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-210-1 3-BNZ		Date	Apr 25 2016	
Date Stamp	Apr 25 2016	File Name	C:\USERS\IERMAN\DESKTOP\UNTITLED FOLDER\ERM-ENEP-210-1 3-BNZ 20160425 01\PROTON_03.FID\FID		Original Points Count	16384	
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	64		
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	2416.3853	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



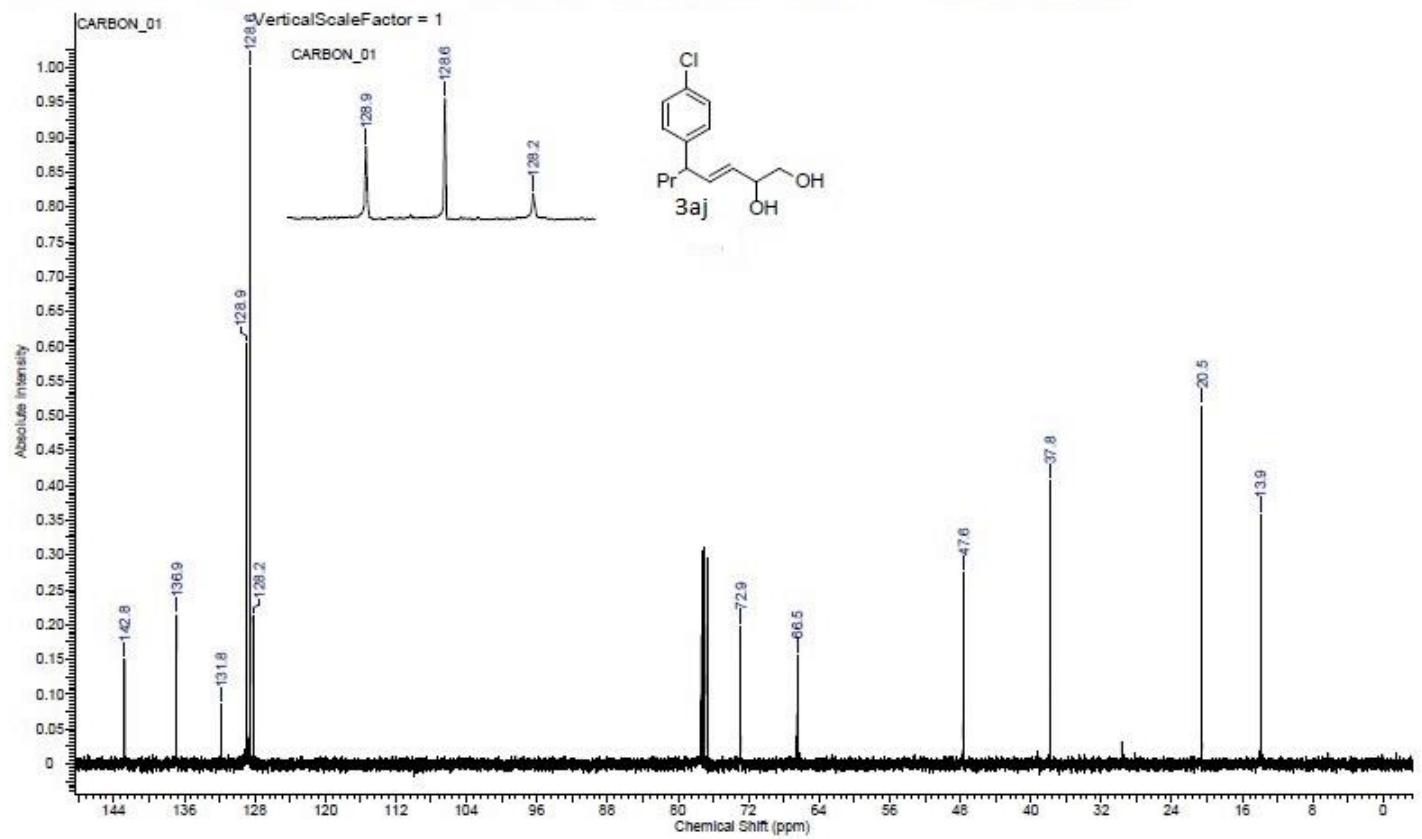
Acquisition Time (sec)	1.3107	Comment	erm-eneq-210-1 3-brz	Date	Apr 27 2016
Date Stamp	Apr 27 2016	File Name	C:\USERS\IERMAN\DESKTOP\UNTITLED FOLDER\ERM-ENEP-210-1 3-BNZ_20160426_01\CARBON_02.FID\FID		
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	12000
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



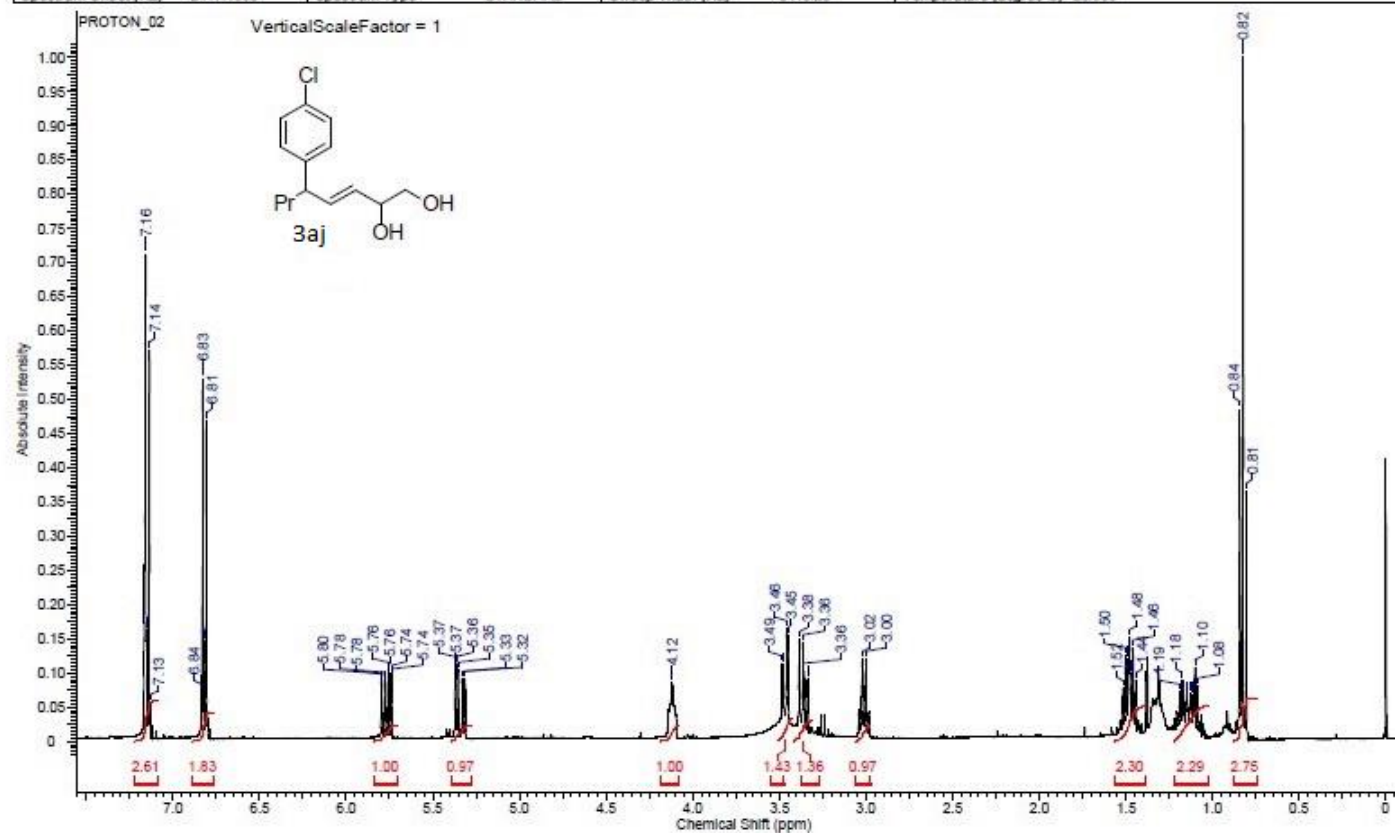
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-195-1_3		Date	Feb 15 2016	
Date Stamp	Feb 15 2016	File Name	C:\USERS\ERM\GOOGLE DRIVE\ERMAN\ERM-ENEP-195-1_3_20160215_01\PROTON_02\FID\FID				
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	32	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2bul	Receiver Gain	52.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



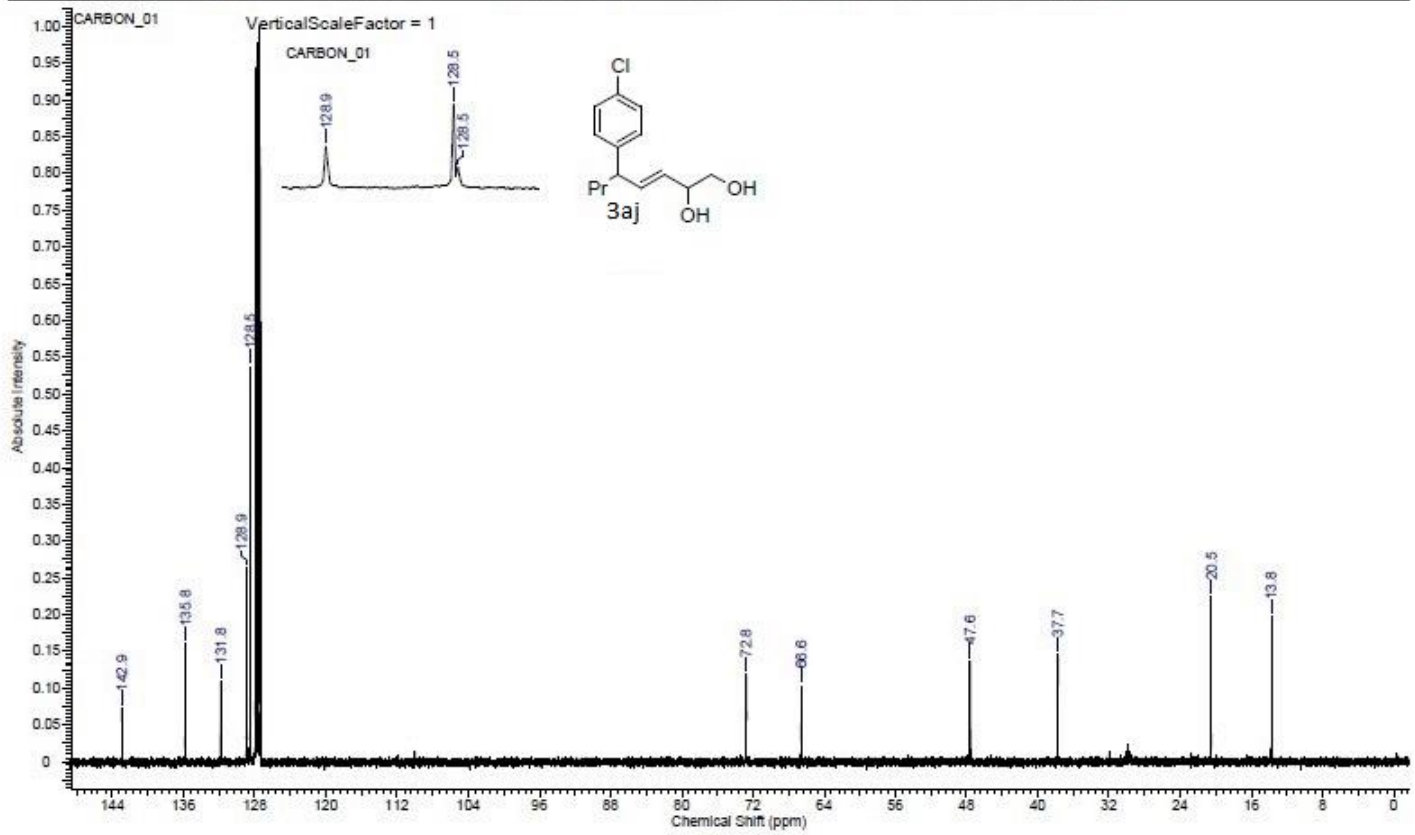
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Date Stamp	Feb 15 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-195-1_3	20160215_01\CARBON_01.FID\FID	
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



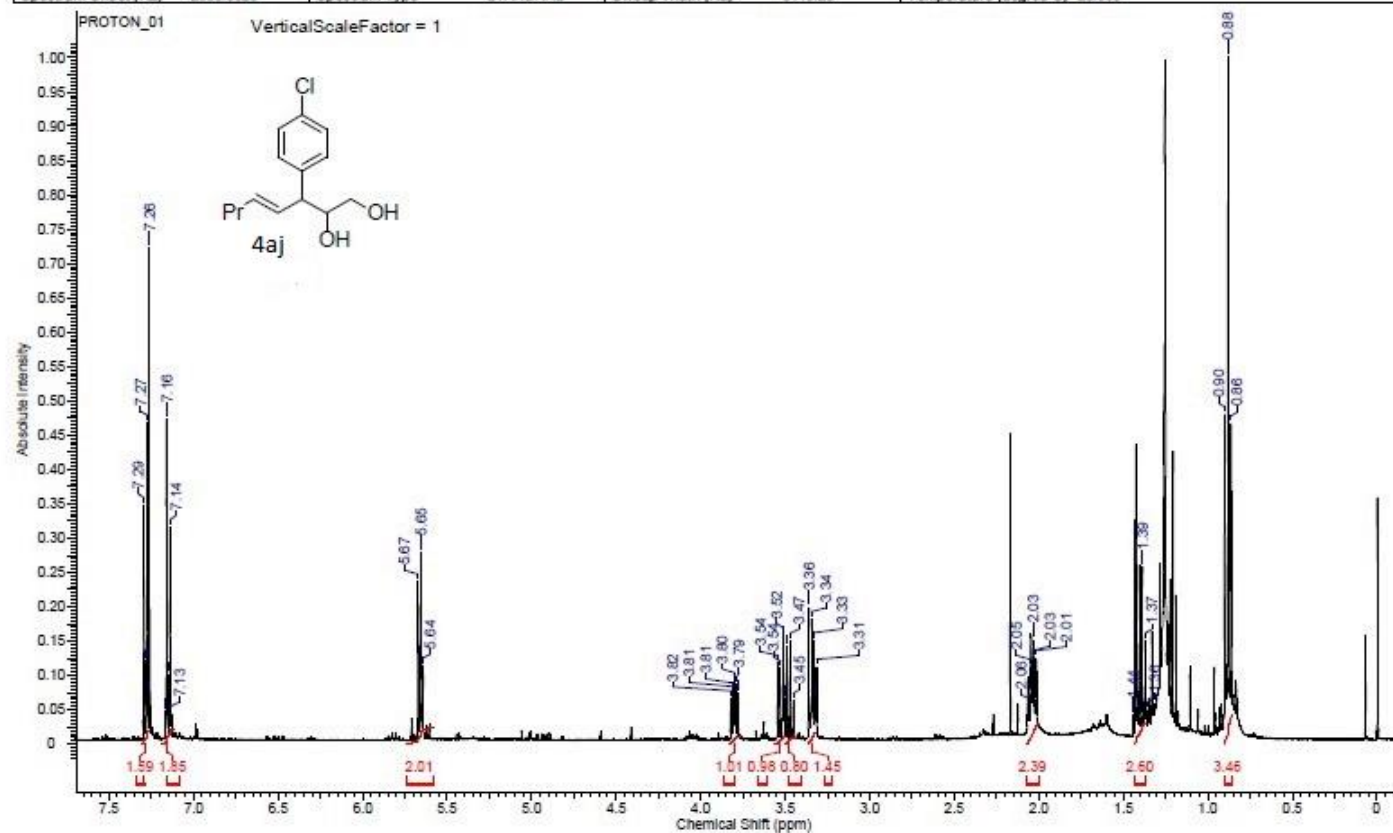
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-195-1_3	Date	Feb 12 2016
Date Stamp	Feb 12 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-195-1_3_20160212_01\PROTON_02.FID\FID		
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	50.00
Spectrum Offset (Hz)	2417.1006	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



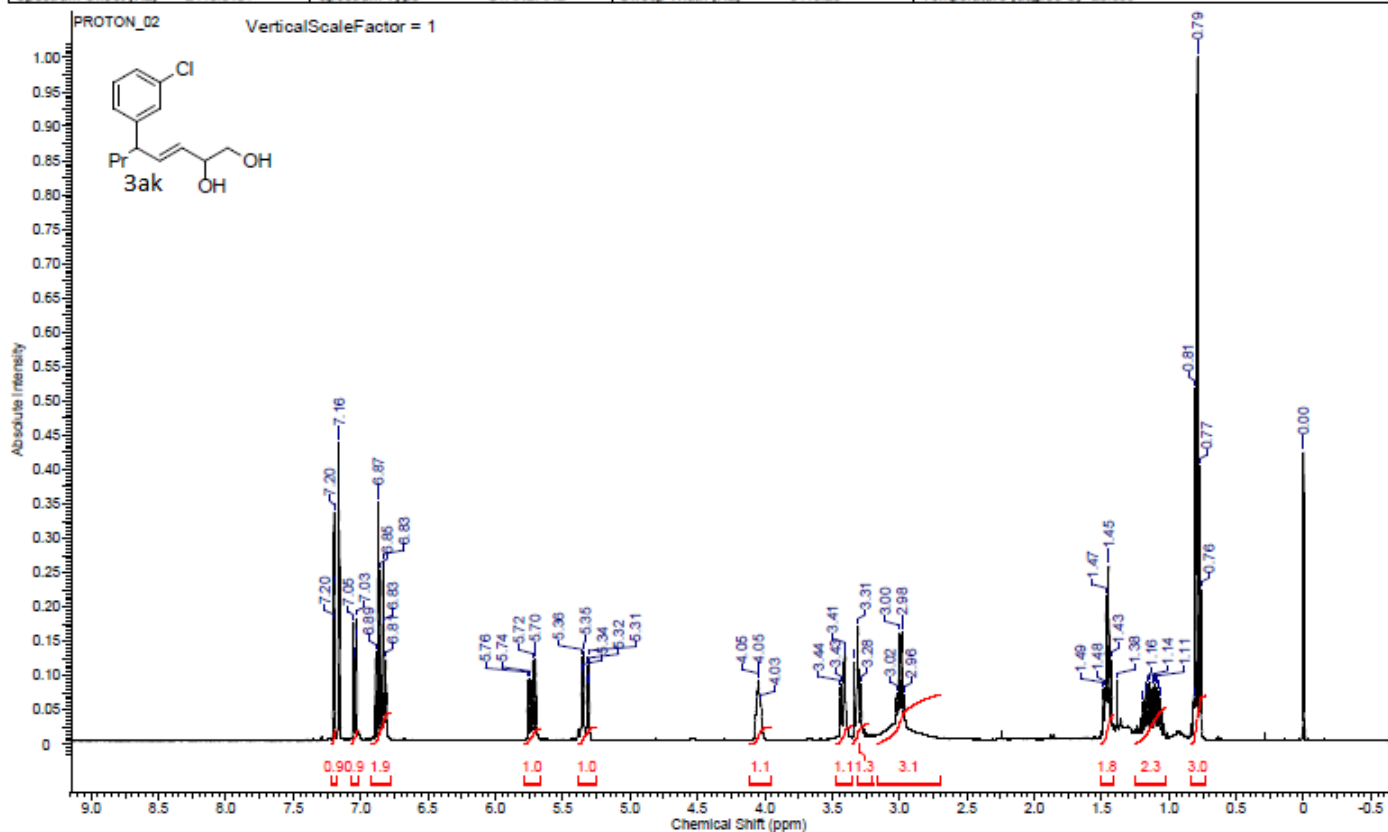
Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-195-1_3		Date	Feb 12 2016	
Date Stamp	Feb 12 2016	File Name	C:\USERS\ERM\GOOGLE DRIVE\ERMAN\ERM-ENEP-195-1_3_20160212_02\CARBON_01.FID\FID				
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-195-1-1	Date	Feb 16 2016
Date Stamp	Feb 16 2016	File Name	C:\USERS\IERM\GOOGLE DRIVE\IERM\ERM-ENEP-195-1-1_20160216_01\PROTON_01.FID\FID		
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2ou1	Receiver Gain	60.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000

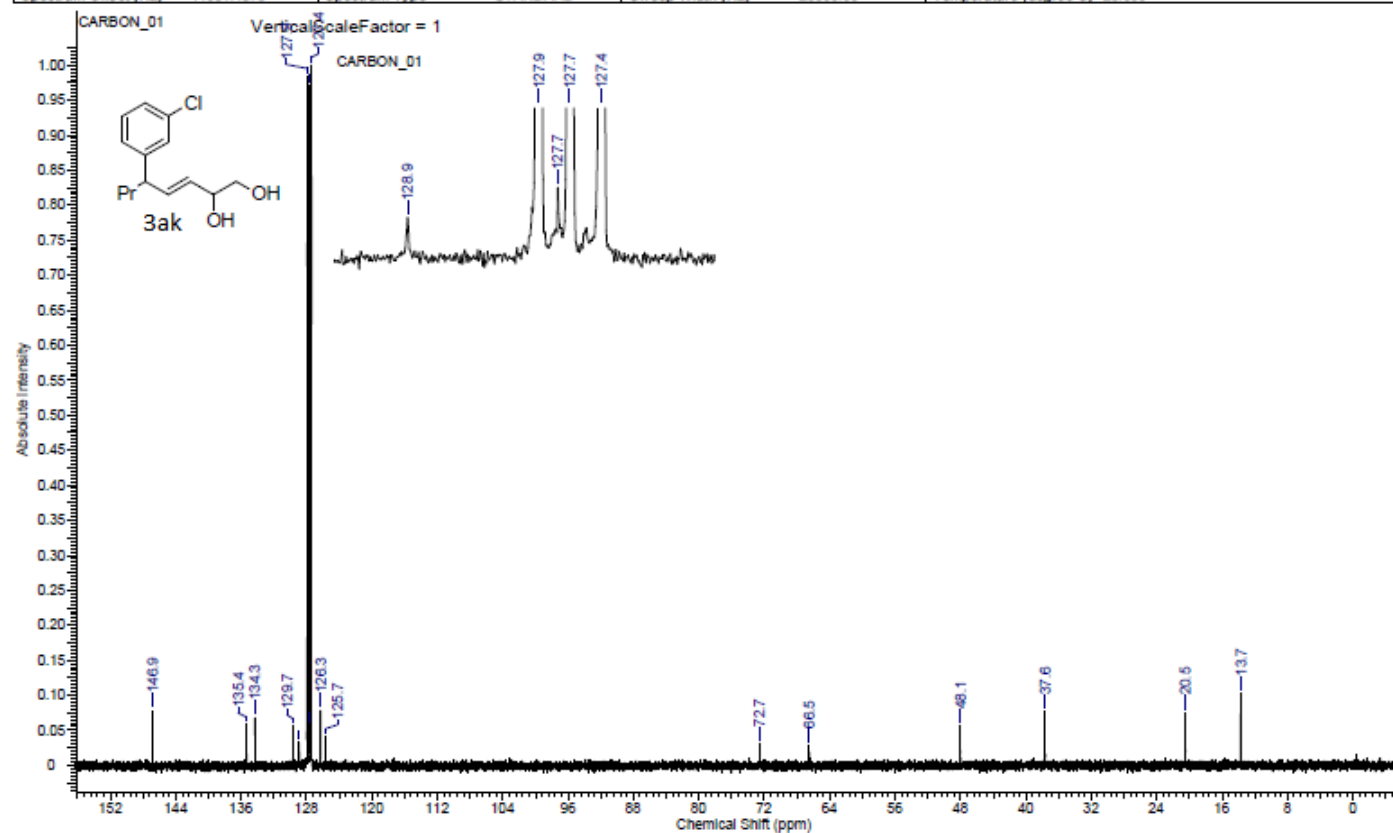


Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-213-1 3-BNZ		Date	May 4 2016	
Date Stamp	May 4 2016	File Name	C:\USERS\IERMAN\DESKTOP\UNTITLED FOLDER\ERM-ENEP-213-1 3-BNZ 20160504 01\PROTON_02.FID\FID				
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	32	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	54.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	2416.3181	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000

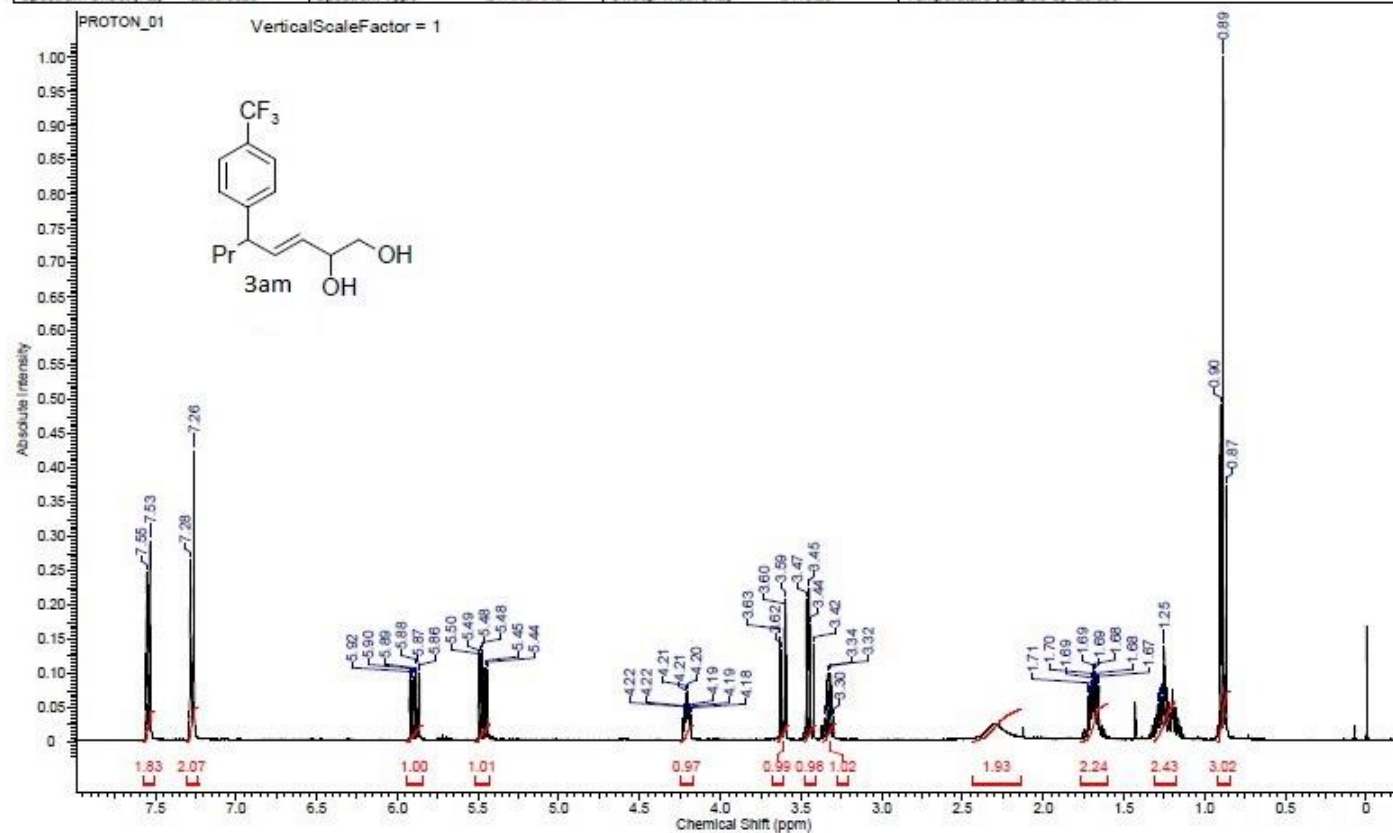


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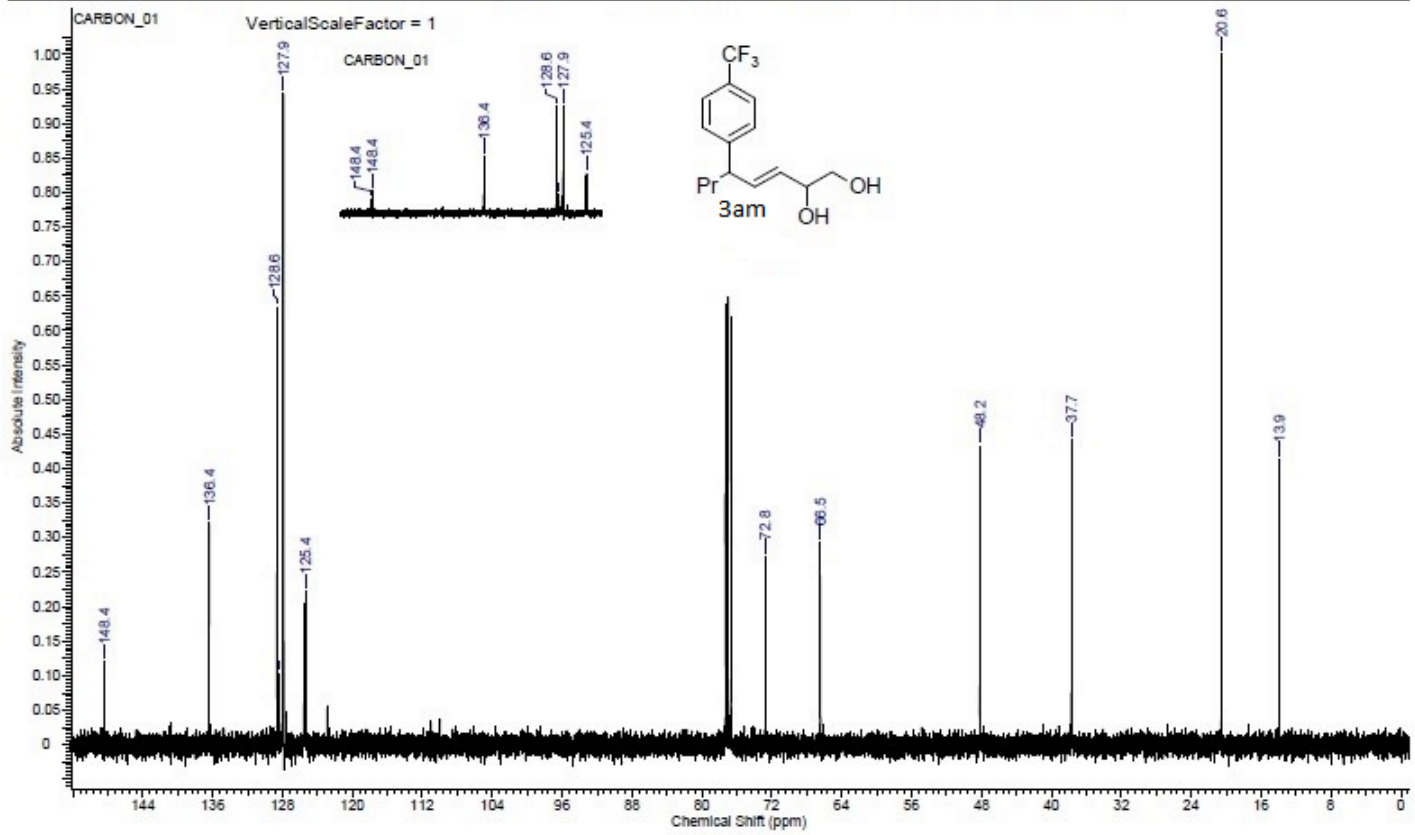
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Date Stamp	May 4 2016	File Name	C:\USERS\IERMAN\DESKTOP\UNTITLED FOLDER\ERM-ENEP-213-1 3-BNZ_20160504_01\CARBON_01.FID\FID				
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



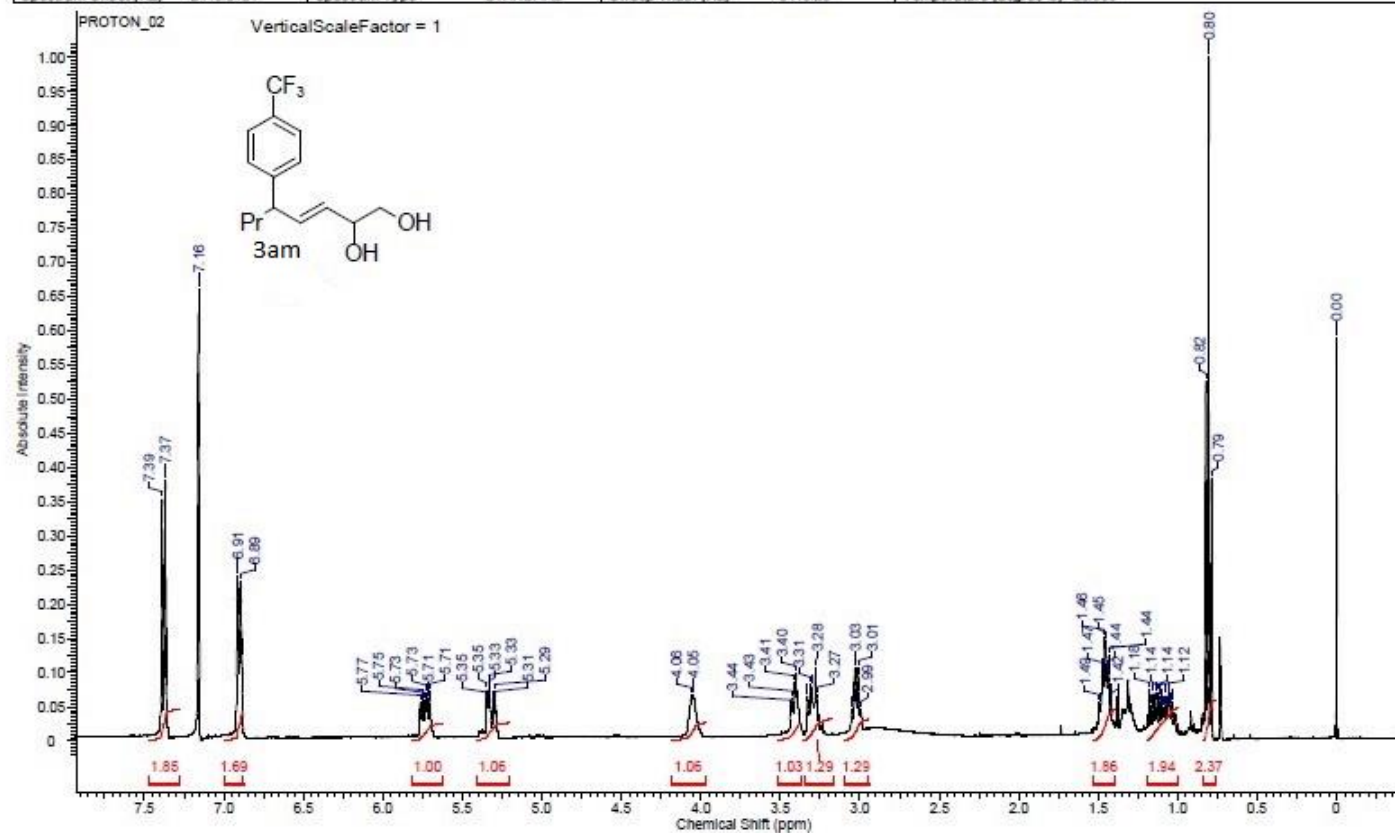
Acquisition Time (sec)	2.5559	Comment	erm-enep-192-1_3	Date	Feb 10 2016
Date Stamp	Feb 10 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-192-1	3_20160210_02\PROTON_01.FID\FID	
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2oul	Receiver Gain	56.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Temperature (degree C)	25.000



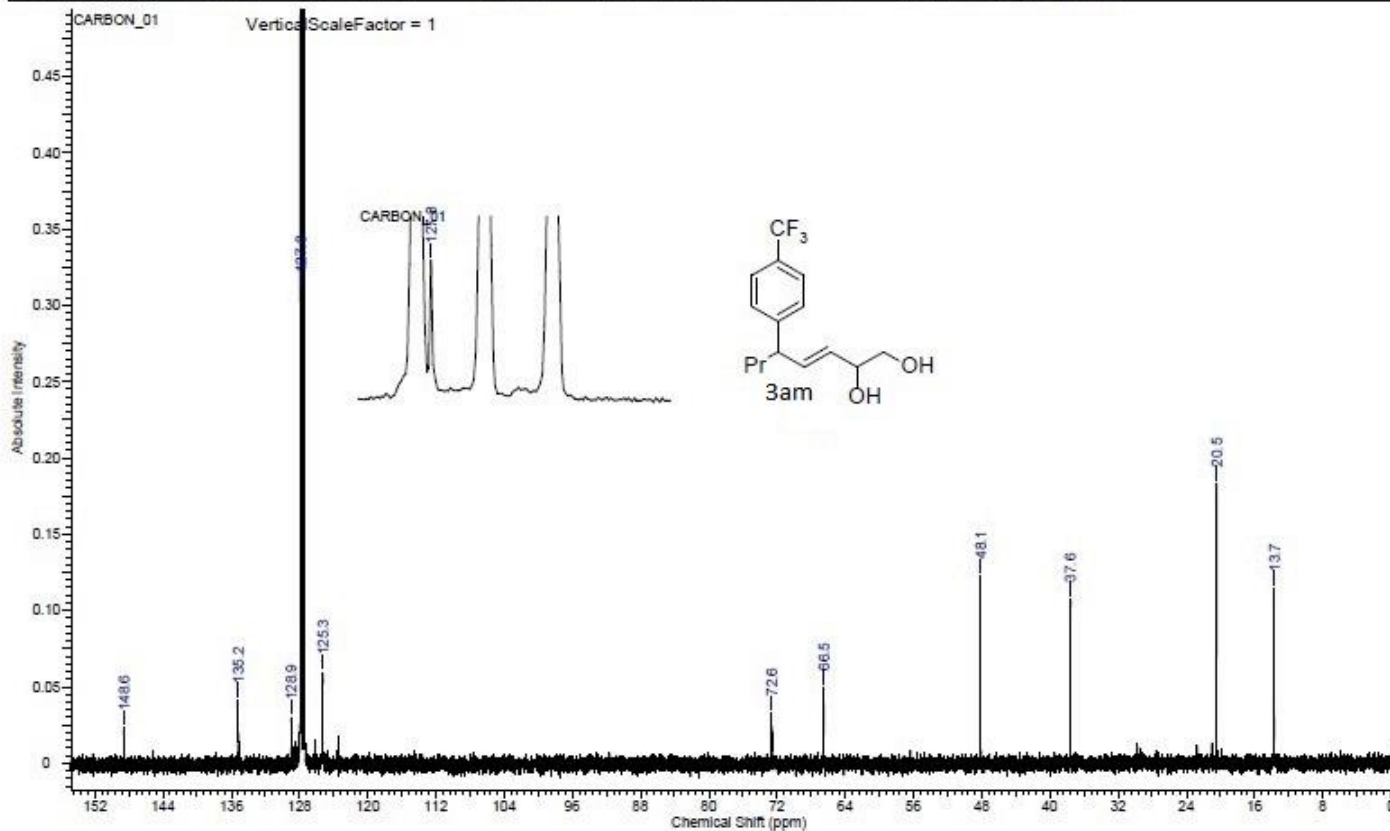
Acquisition Time (sec)	1.3107	Comment	erm-enep-192-1_3	Date	Feb 10 2016
Date Stamp	Feb 10 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-192-1	3_20160210_02\CARBON_01.FID\FID	
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



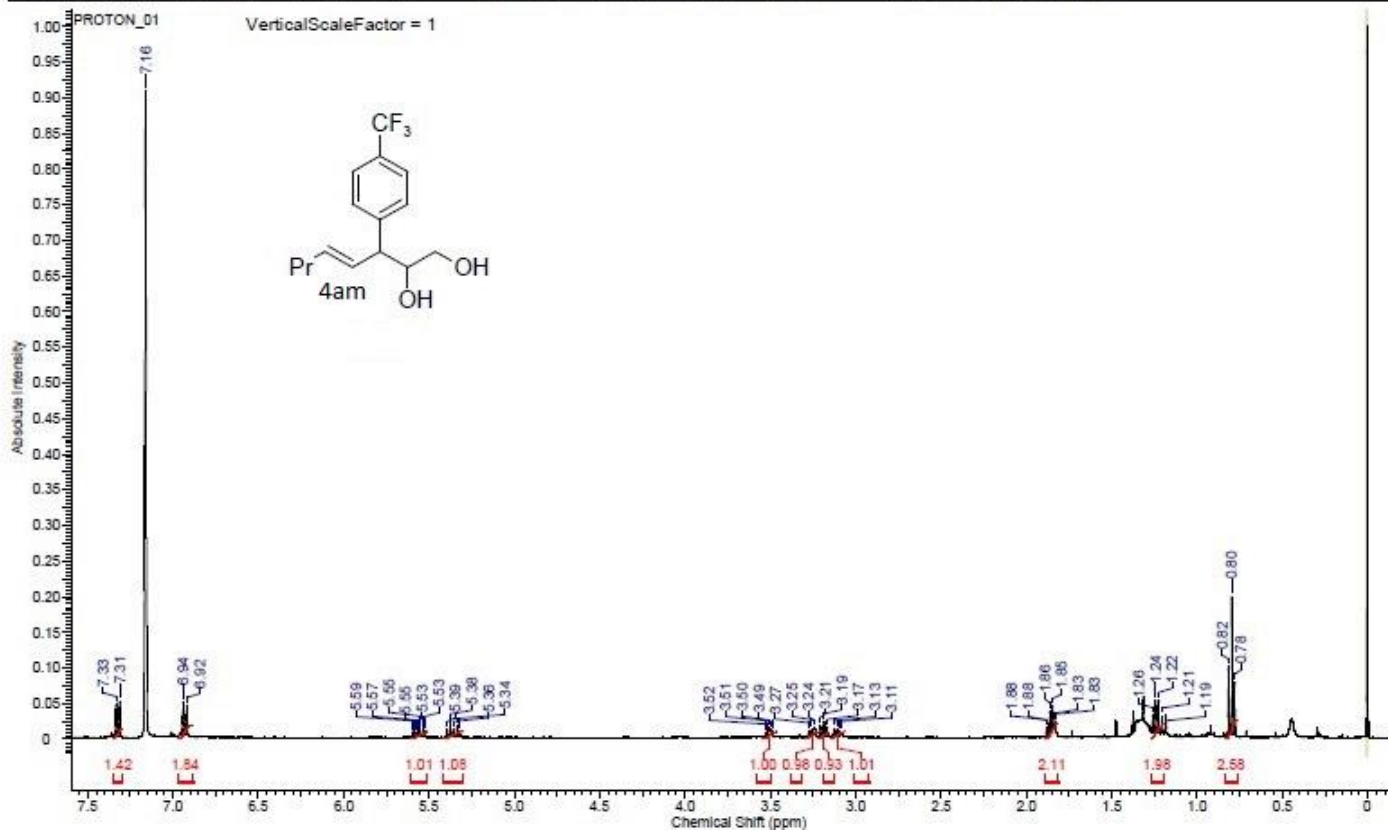
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-192-1_3	Date	Feb 2 2016
Date Stamp	Feb 2 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-192-1_3_20160202_01\PROTON_02.FID\FID		
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	16
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	54.00
Spectrum Offset (Hz)	2416.3181	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



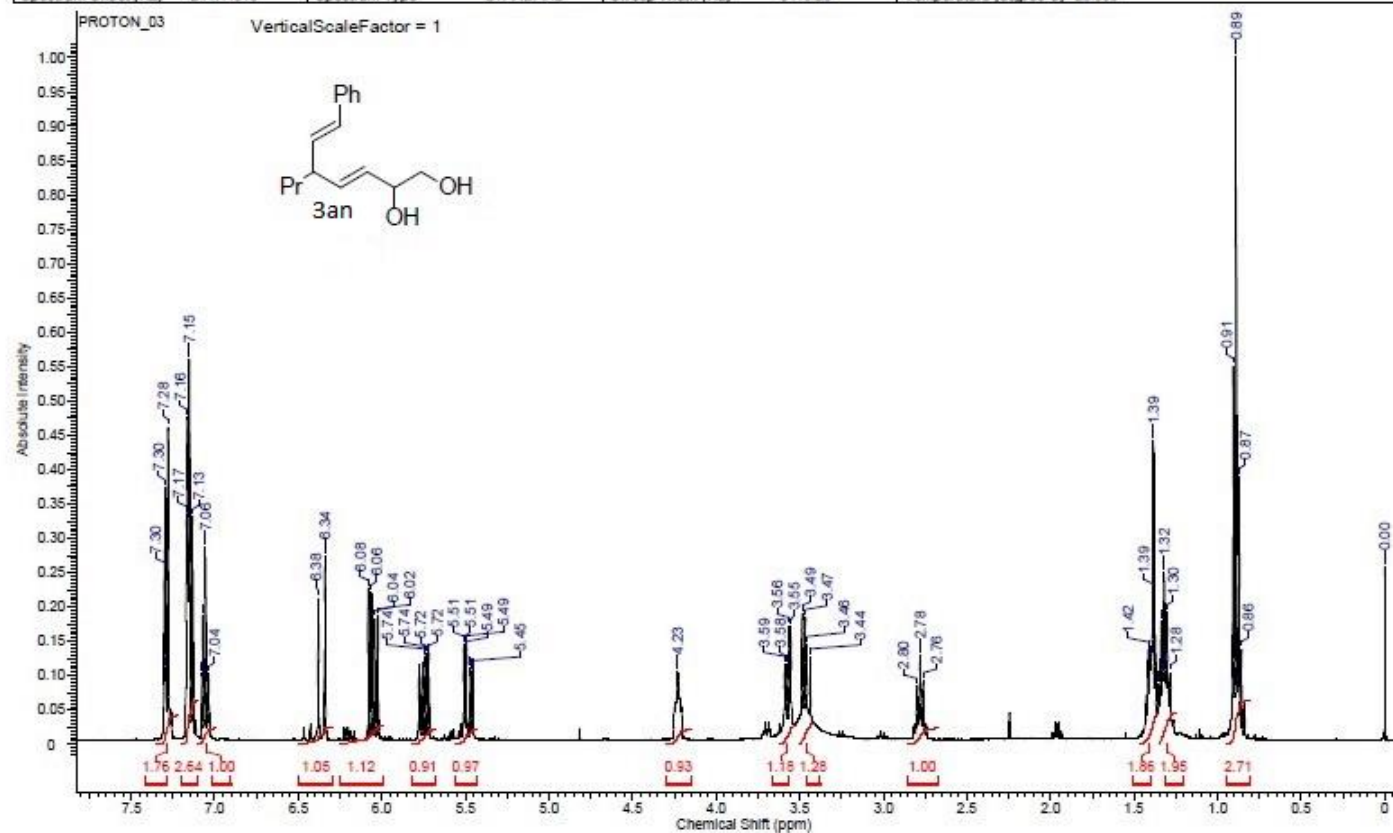
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Date Stamp	Feb 2 2016	File Name	C:\USERS\ERM\GOOGLE DRIVE\ERMAN\ERM-ENEP-192-1_3_20160202_01\CARBON_01.FID\FID	Original Points Count	32768
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



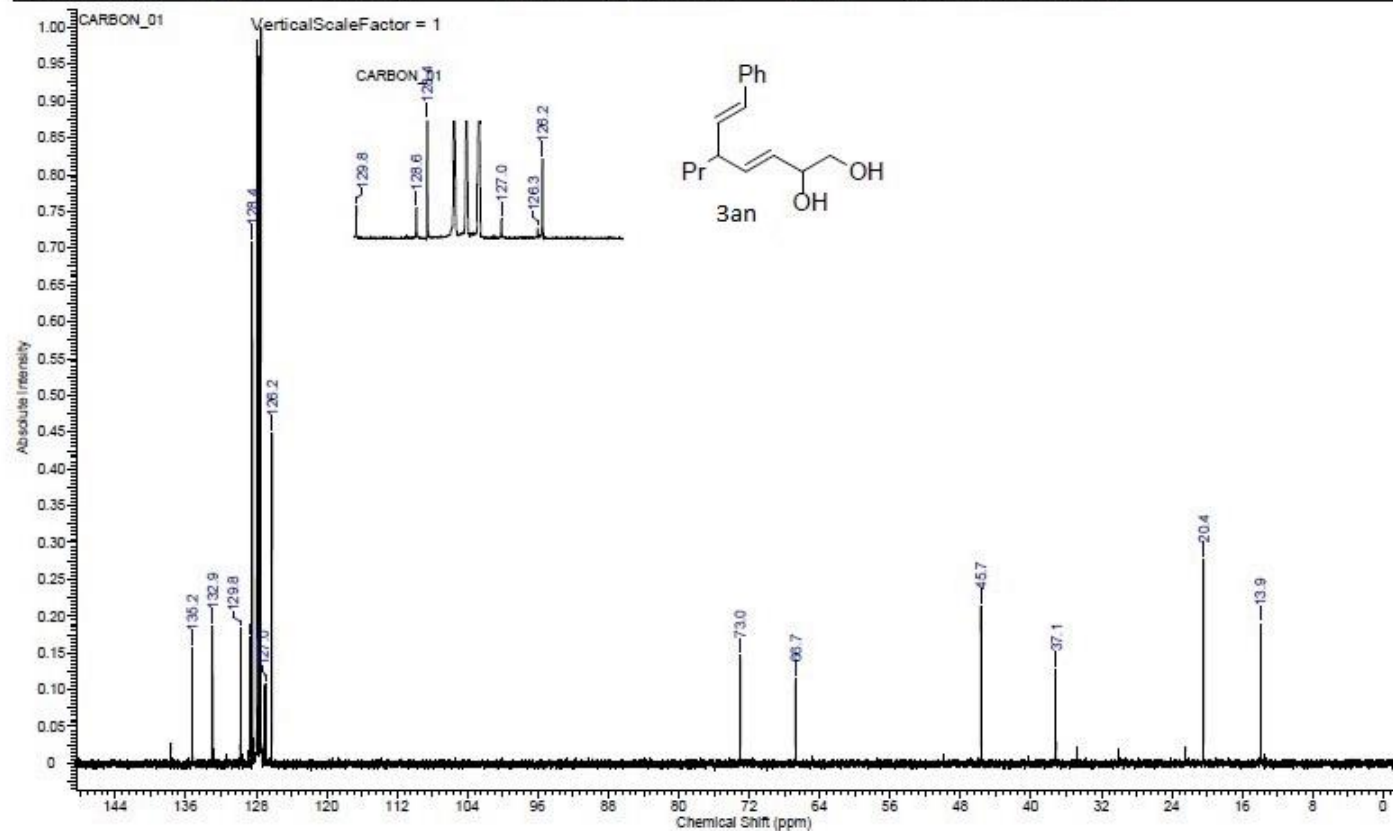
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Date Stamp	Feb. 2, 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-192-1_1_20160202_01\PROTON_01.FID\FID		
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	32
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00
Spectrum Offset (Hz)	2416.7764	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



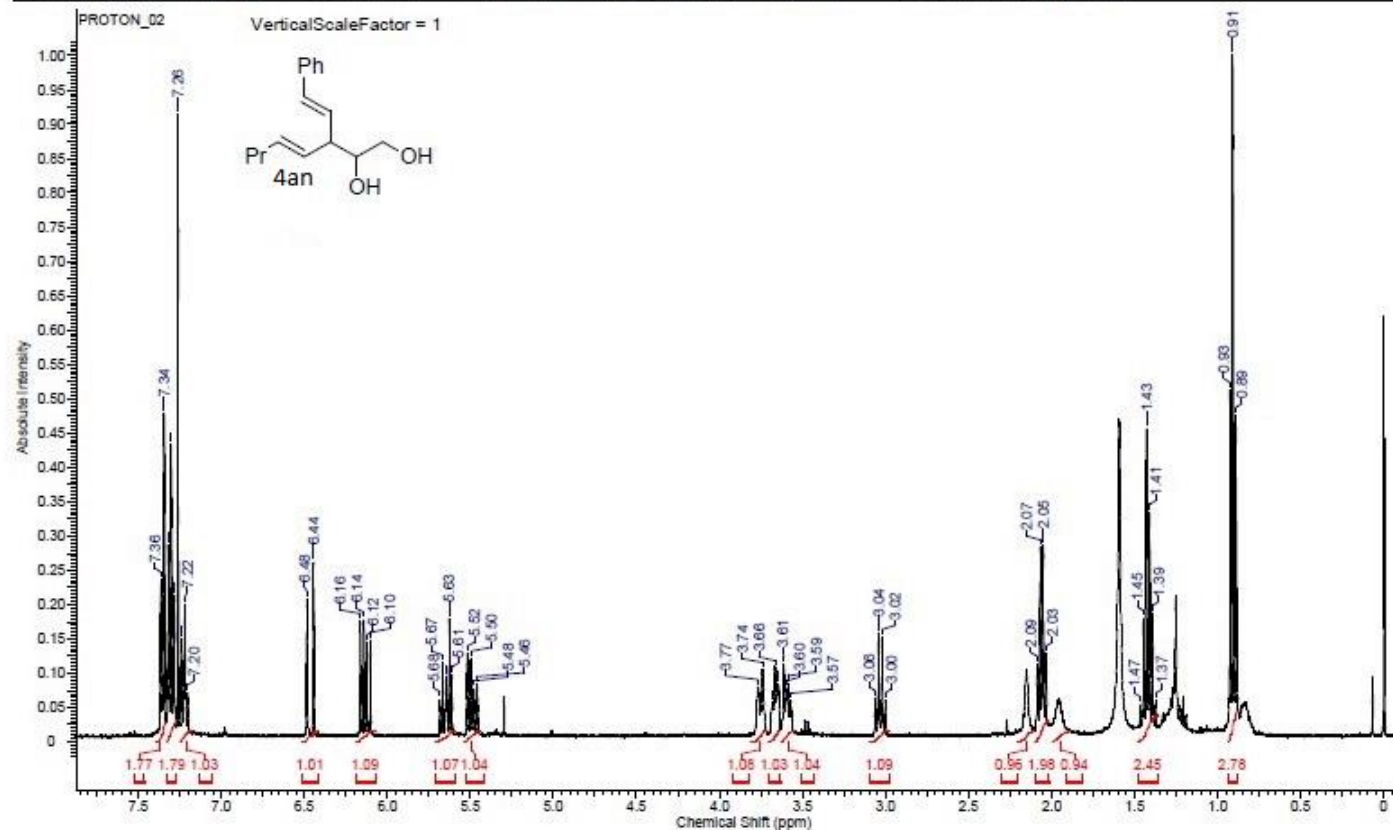
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-202-1 3	Date	Feb 26 2016
Date Stamp	Feb 26 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-202-1 3 20160226_01\PROTON_03.FID\FID		
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	32
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	50.00
Spectrum Offset (Hz)	2417.4919	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



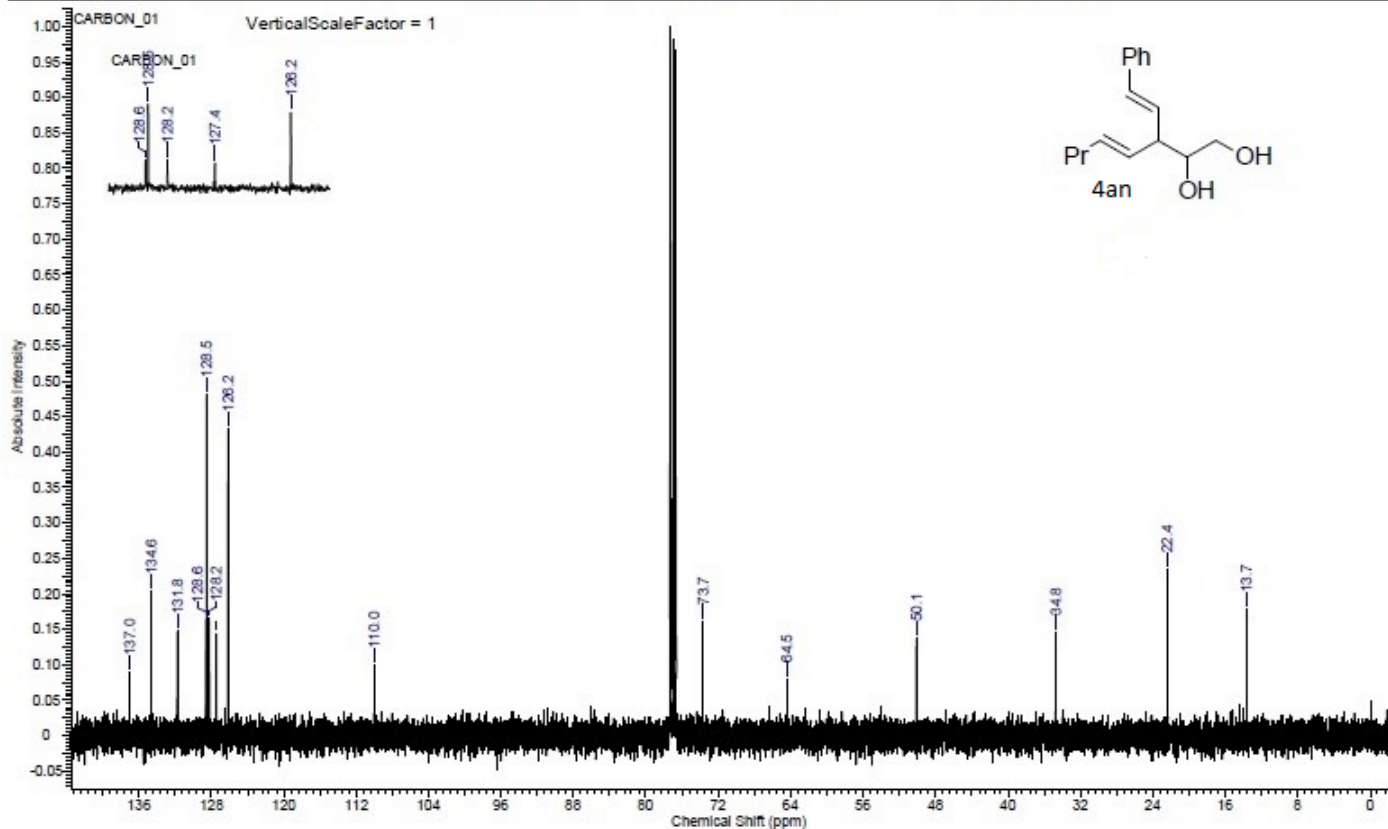
Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-202-1 3		Date	Feb 26 2016	
Date Stamp	Feb 26 2016	File Name	C:\USERS\IERM\GOOGLE DRIVE\IERMAN\ERM-ENEP-202-1 3		20160226_01\CARBON_01.FID\FID		
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



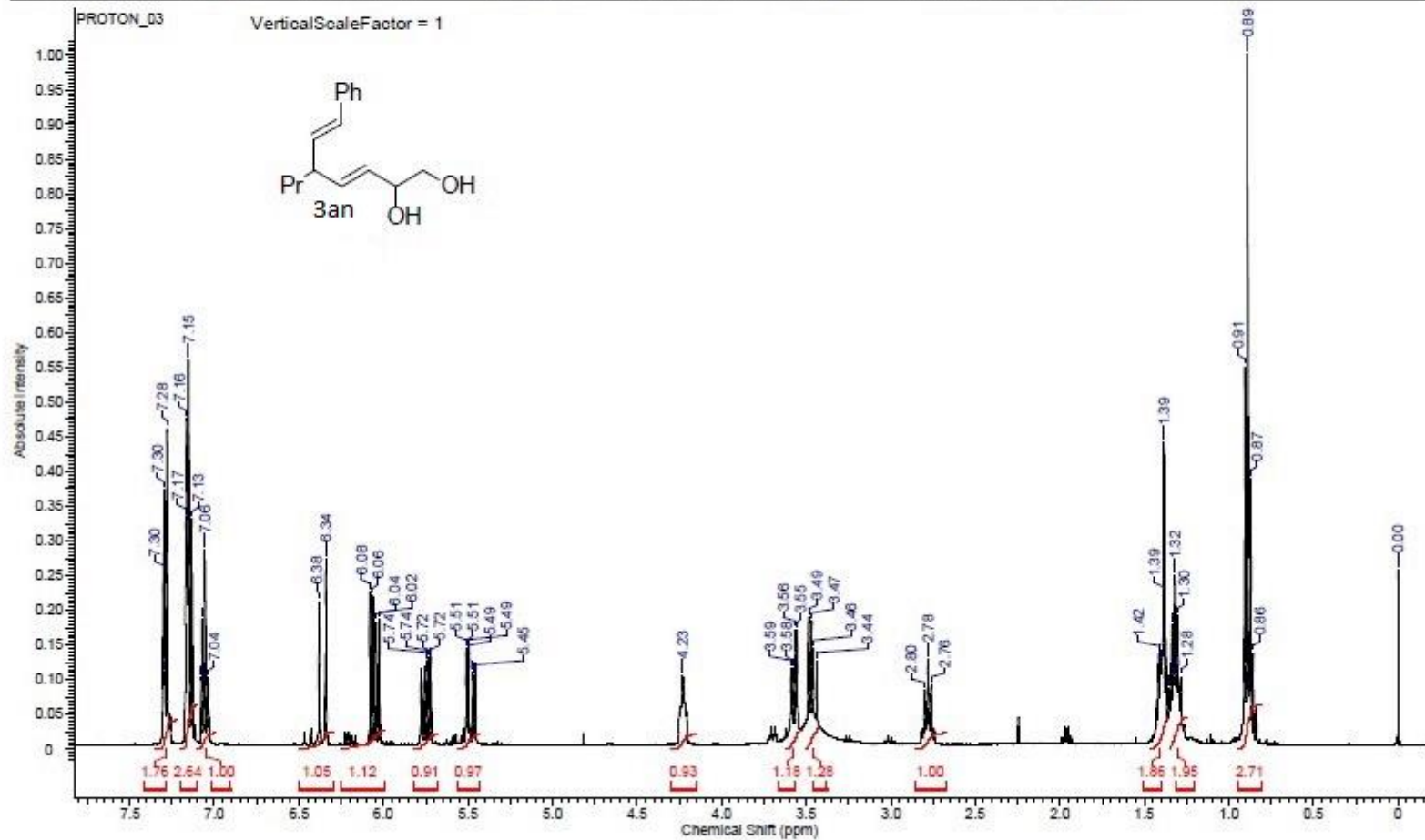
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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



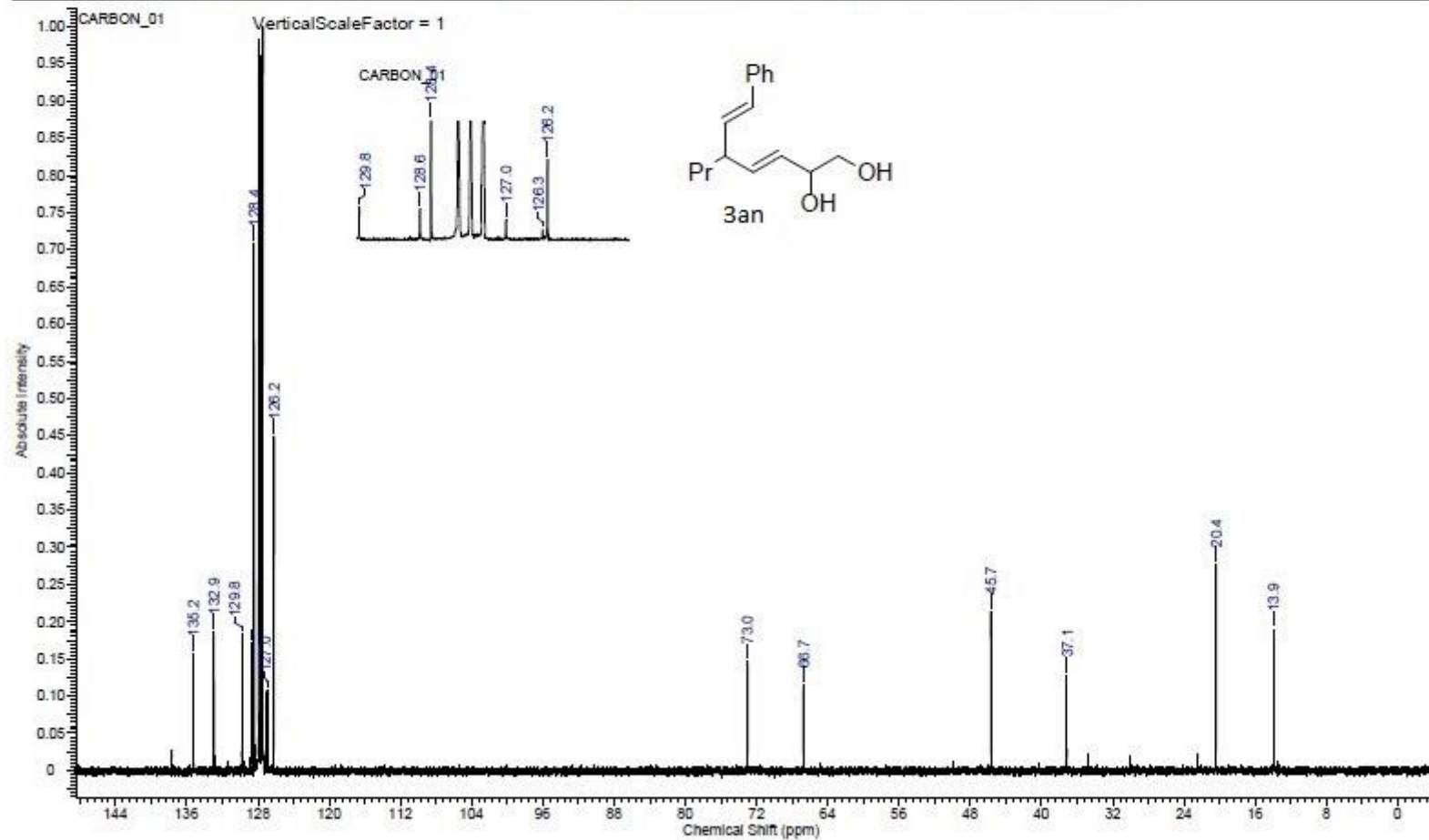
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	400
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



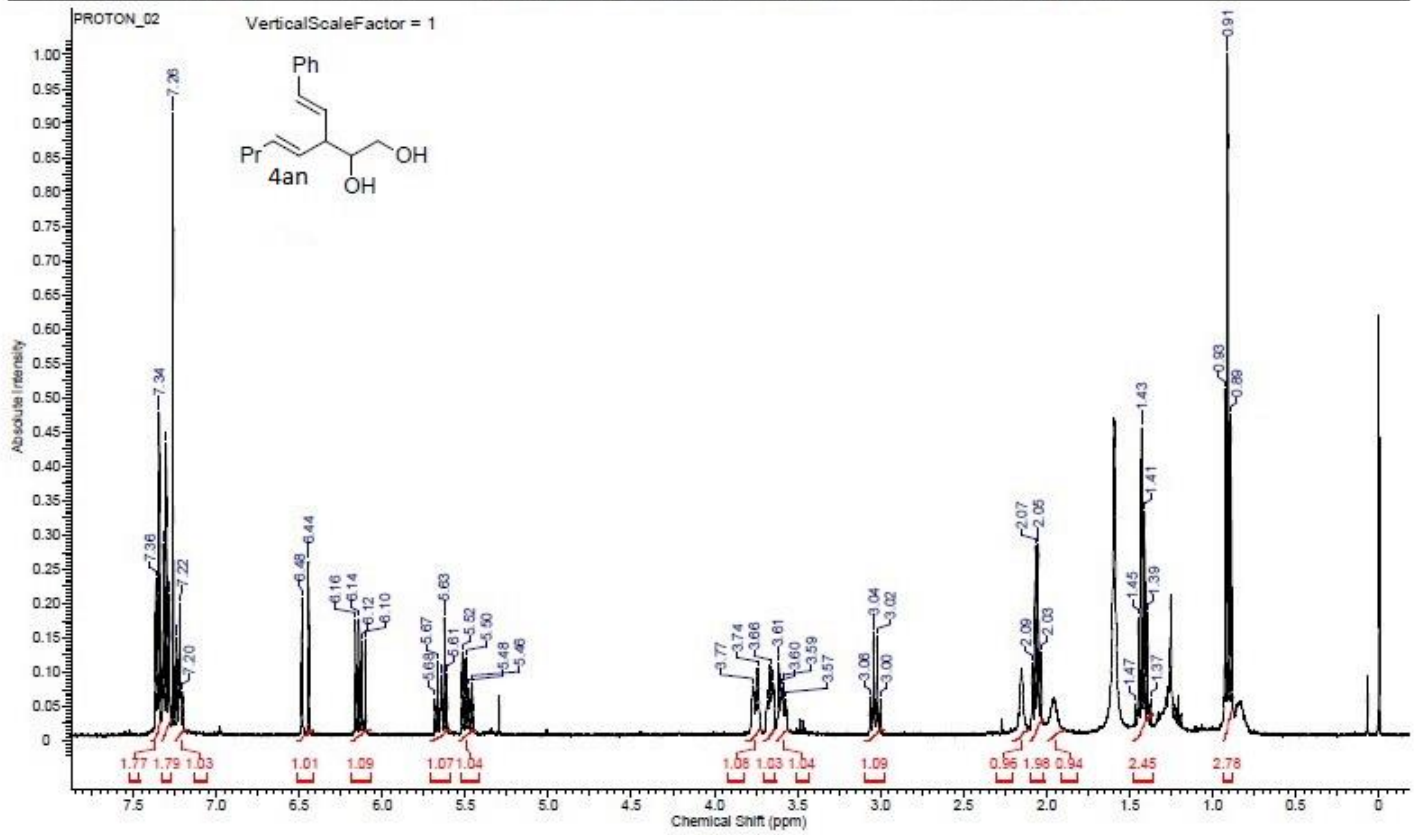
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Date Stamp	Feb 26 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-202-1_3	20160226_01	PROTON_03.FID\FID
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	32
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	50.00
Spectrum Offset (Hz)	2417.4919	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Original Points Count	16384
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



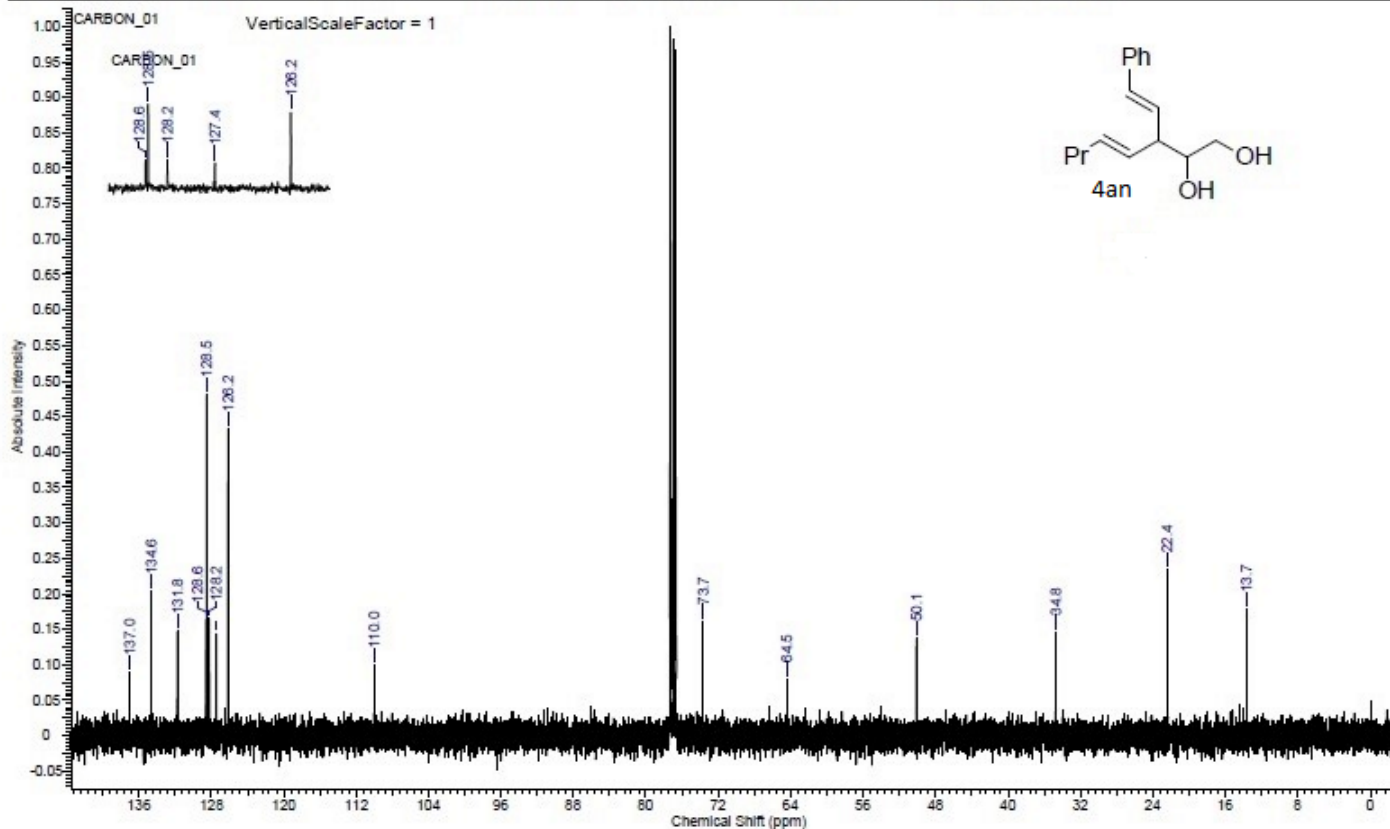
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Date Stamp	Feb 26 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-202-1_3_20160226_01\CARBON_01.FID\FID		
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



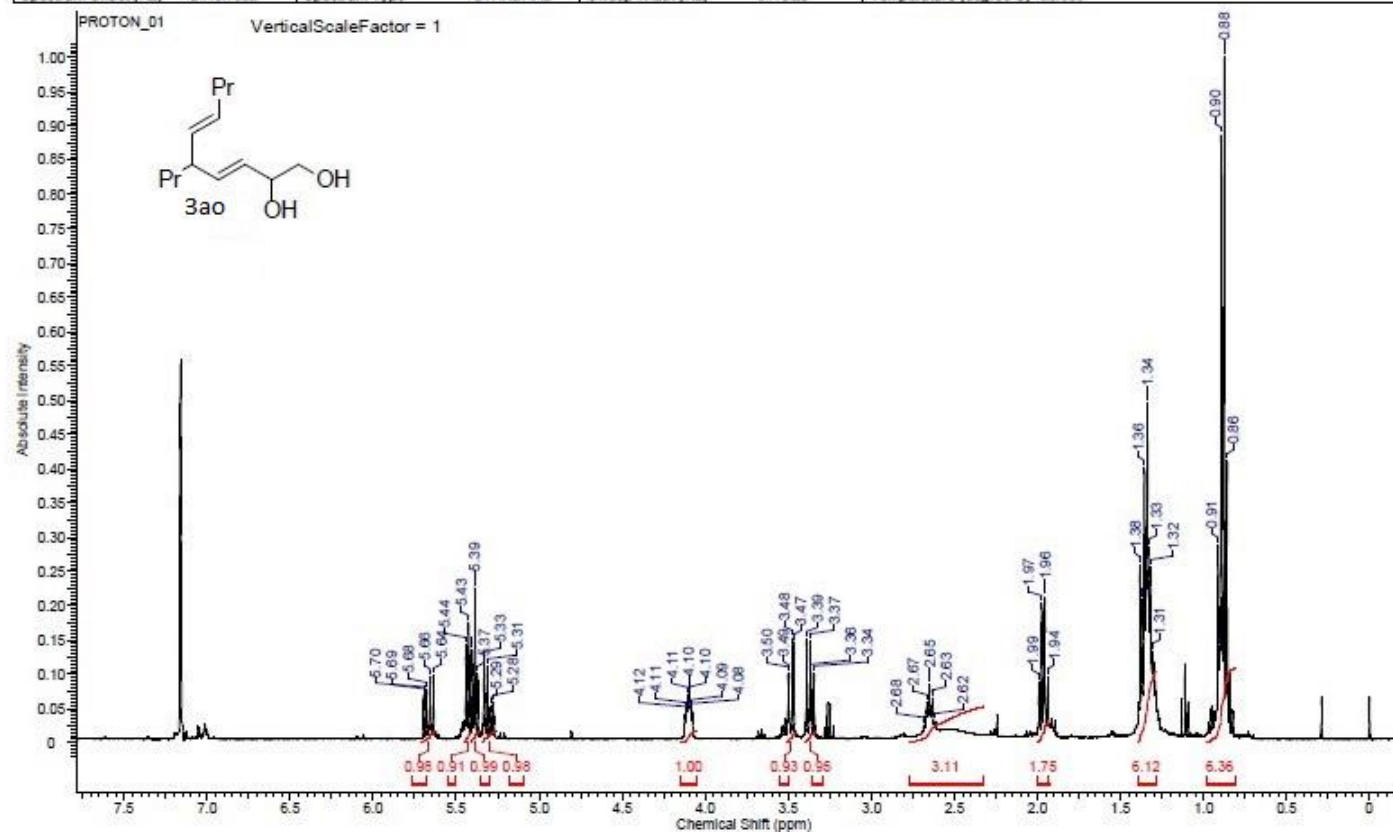
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Date Stamp	Feb 26 2016	File Name	C:\USERS\ERM\GOOGLE DRIVE\ERMAN\ERM-ENEP-202-1	1_20160226_01\PROTON_02.FID\FID	
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



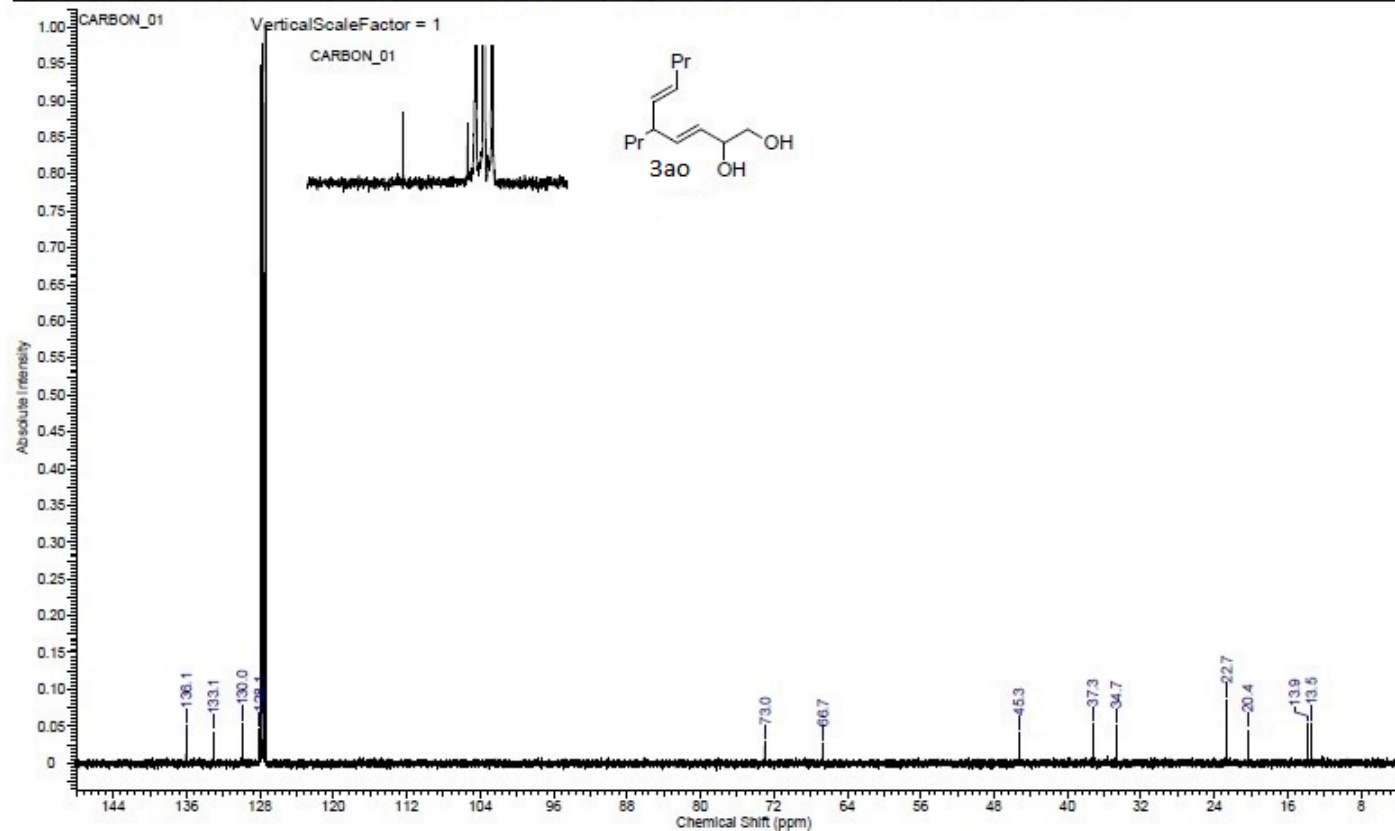
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Date Stamp	Feb 26 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-202-1_1	20160226_01\CARBON_01.FID\FID	
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	400
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



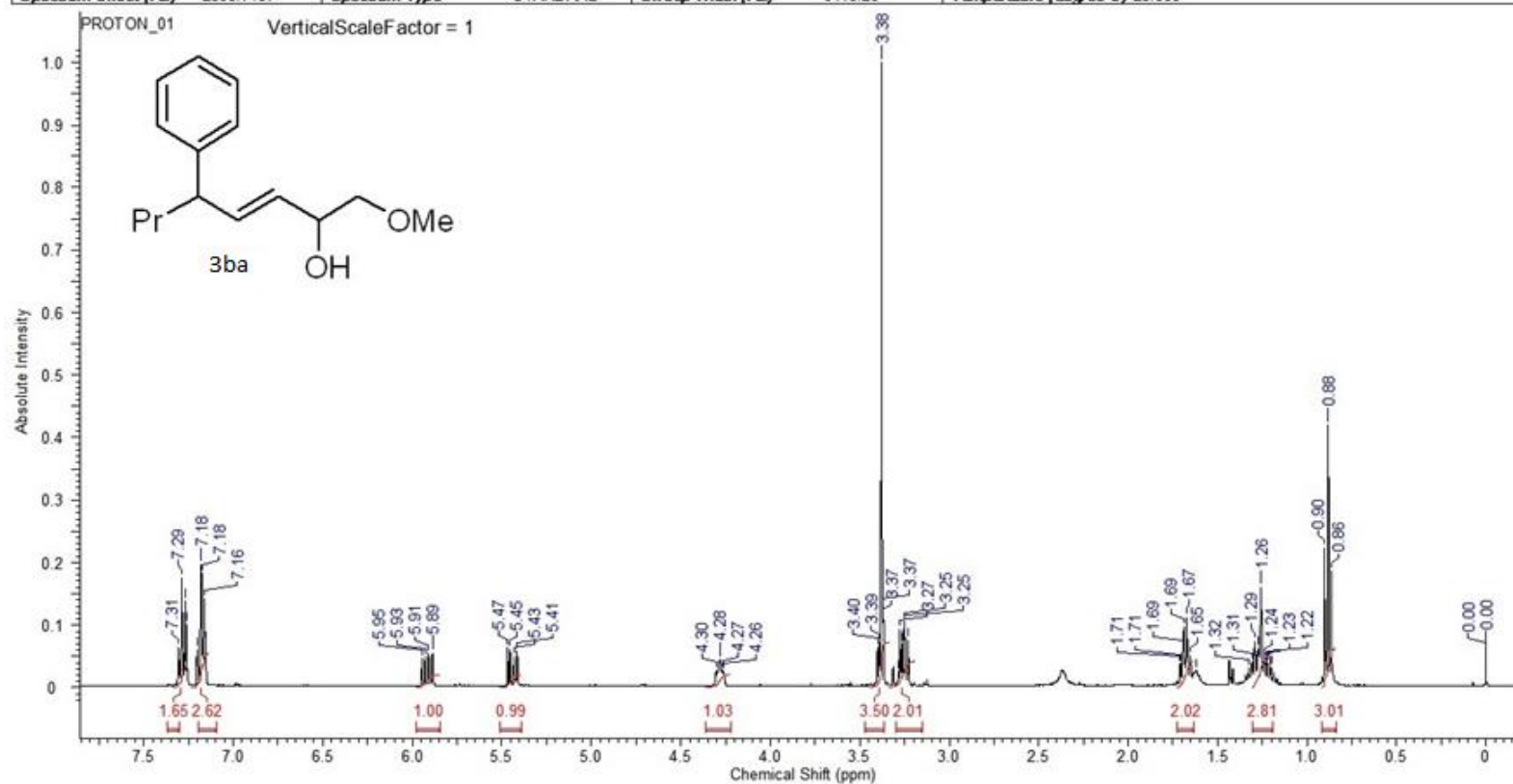
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-204-TKR-1-3		Date	Apr 5 2016	
Date Stamp	Apr 5 2016	File Name	D:\NMR\ENEP-204-TKR-1-3_20160405_01\PROTON_01.FID\FID				
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	58.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	2416.7092	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-204-TKR-1-3	Date	Apr 6 2016
Date Stamp	Apr 6 2016	File Name	C:\USERS\ERMAN\DESKTOP\EFER\ERM-ENEP-204-TKR-1-3 20160405_01\CARBON_01.FID\FID		
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	400
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



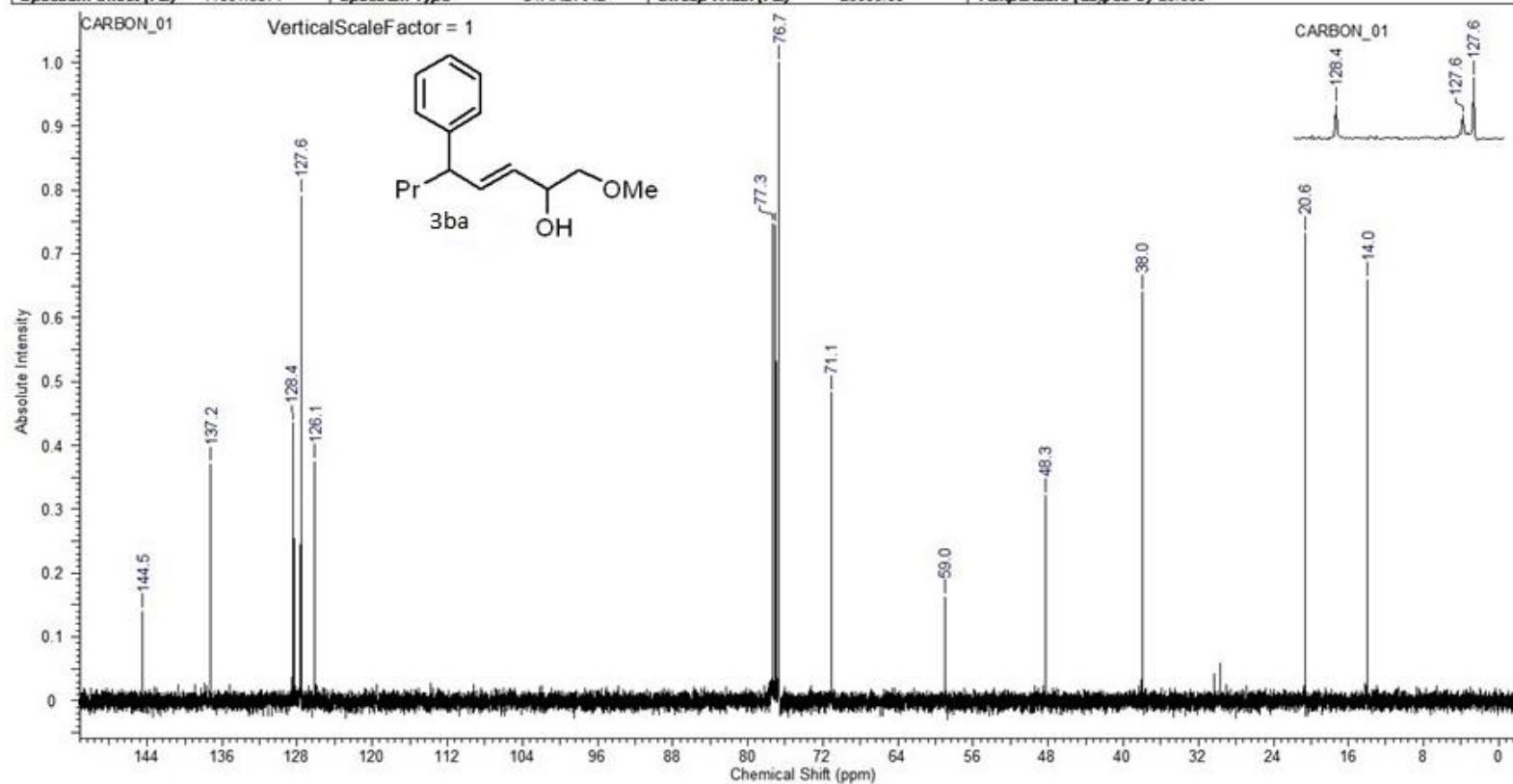
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Date Stamp	Oct 6 2015	File Name	C:\Users\Erman\Desktop\123\ERM-ENEP-114-1_3_20151006_01\PROTON_01.fid\fid				
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	32	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	56.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2398.7107	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



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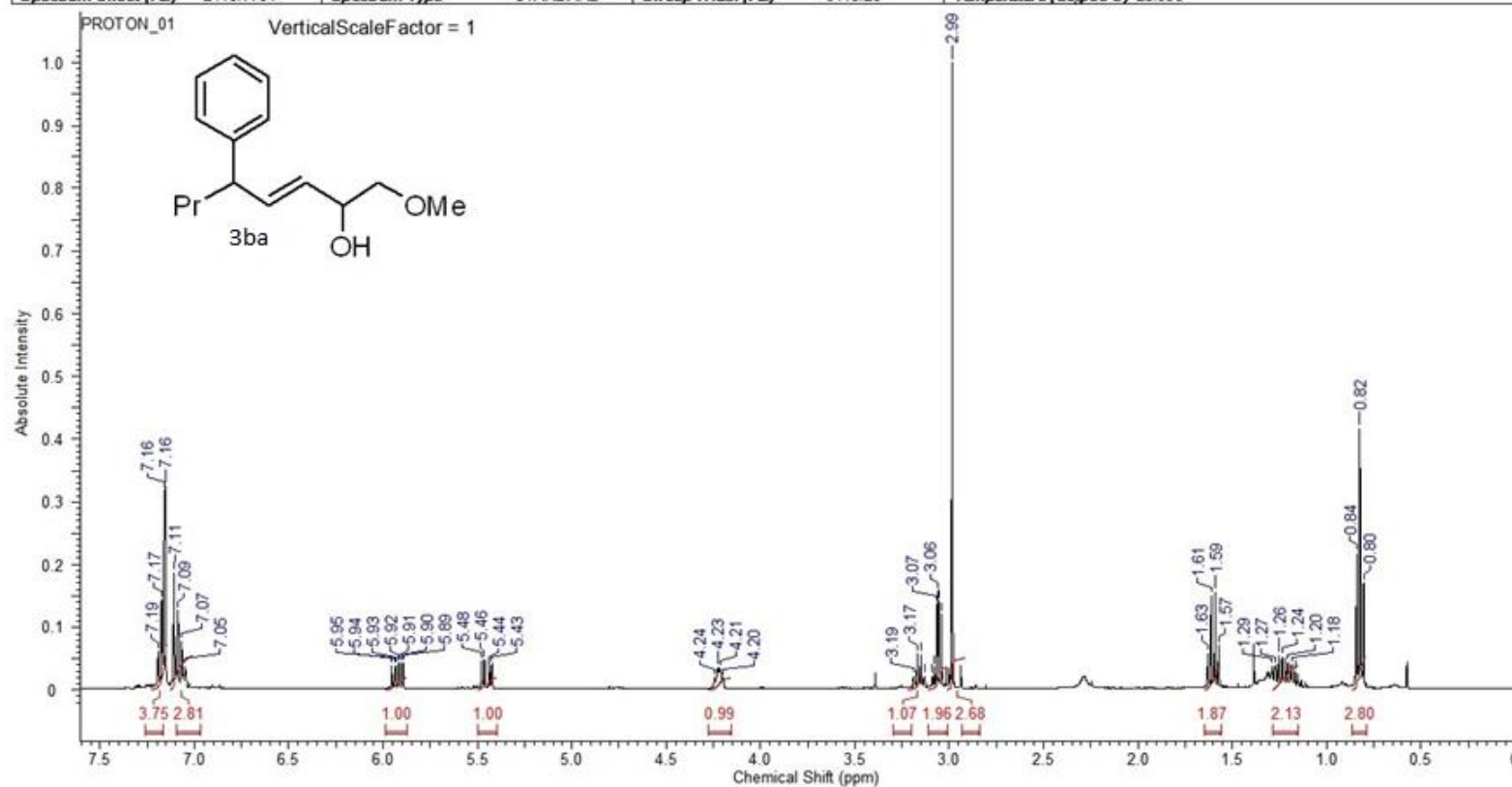
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Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-114-C-CDCL3		Date	Oct 7 2015	
Date Stamp	Oct 7 2015	File Name	C:\Users\Erman\Desktop\123\ERM-ENEP-114-C-CDCL3_20151007_01\CARBON_01.fid				
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	512	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



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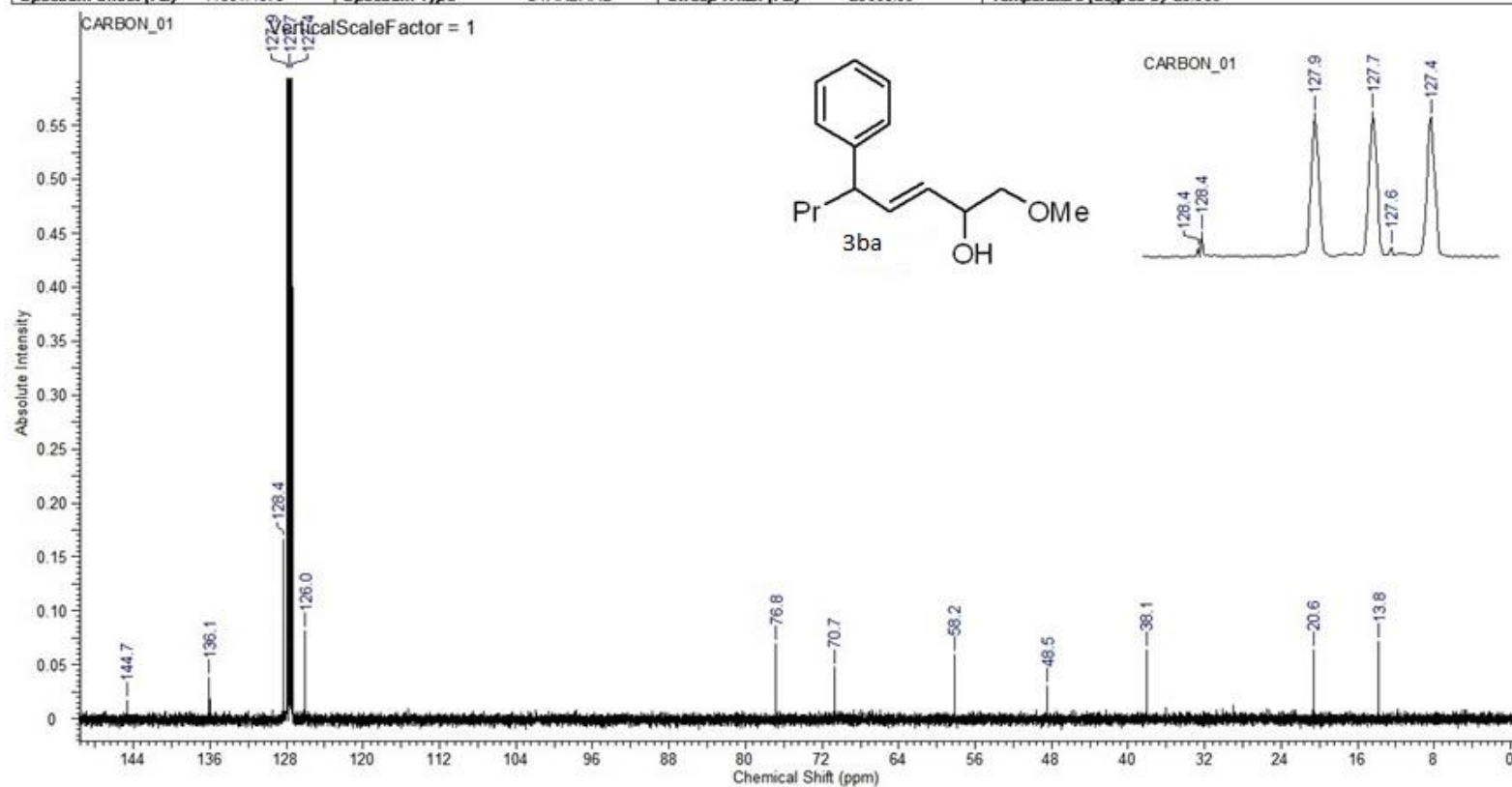
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-81-1_3		Date	Apr 28 2015	
Date Stamp	Apr 28 2015	File Name	C:\Users\Erman\Desktop\ERMAN\ERM-ENEP-81-1_3_20150428_01\PROTON_01.fid				
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	32	Original Points Count	16384
Points Count	16384	Pulse Sequence	sgpul	Receiver Gain	56.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	2416.7764	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



C:\Users\Erman\Desktop\ERMAN\ERM-ENEP-81-1_3_20150428_01\PROTON_01

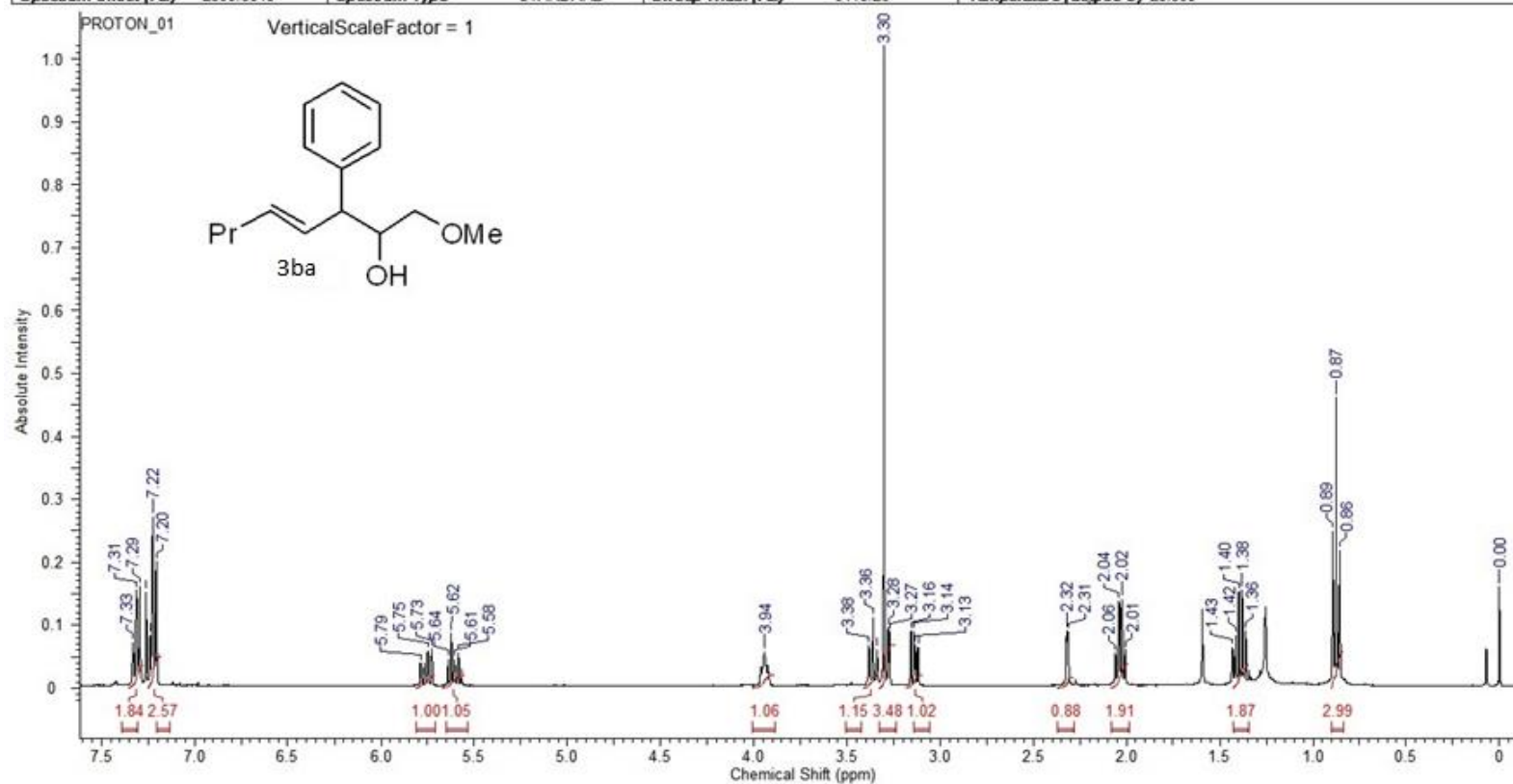
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Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-75-1_3-CABONE		Date	Apr 21 2015	
Date Stamp	Apr 21 2015	File Name	C:\Users\Erman\Desktop\ERMAN\ERM-ENEP-75-1_3-CABONE_20150421_01\CARBON_01.fid				
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



C:\Users\Erman\Desktop\ERMAN\ERM-ENEP-75-1_3-CABONE_20150421_01\CARBON_01

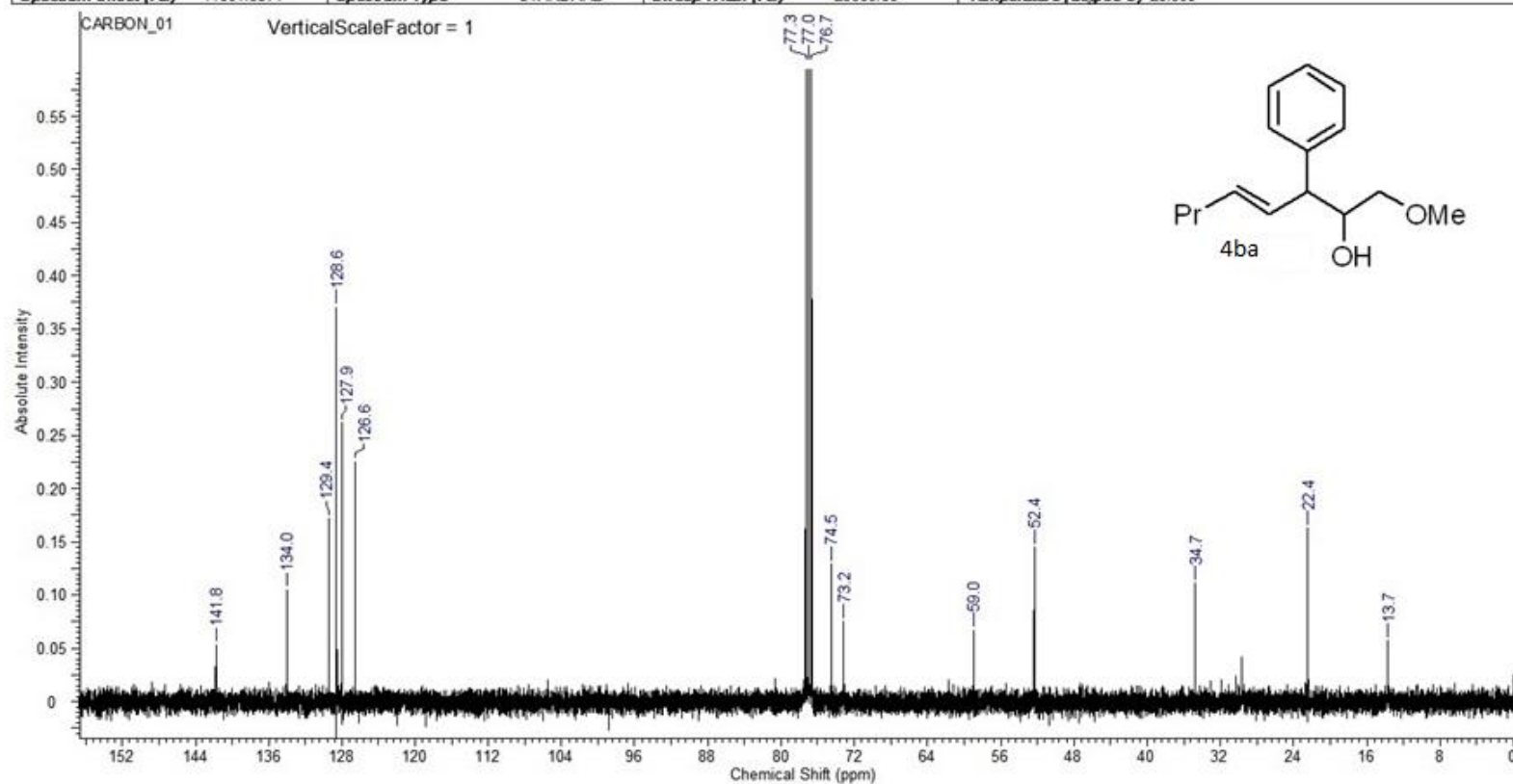
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-54-1_1sub		Date	Mar 21 2015	
Date Stamp	Mar 21 2015	File Name	C:\Users\Erman\Desktop\ERMAN\ERM-ENEP-54-1_1sub_20150321_01\PROTON_01.fidfid				
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	32	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	58.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2399.8845	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



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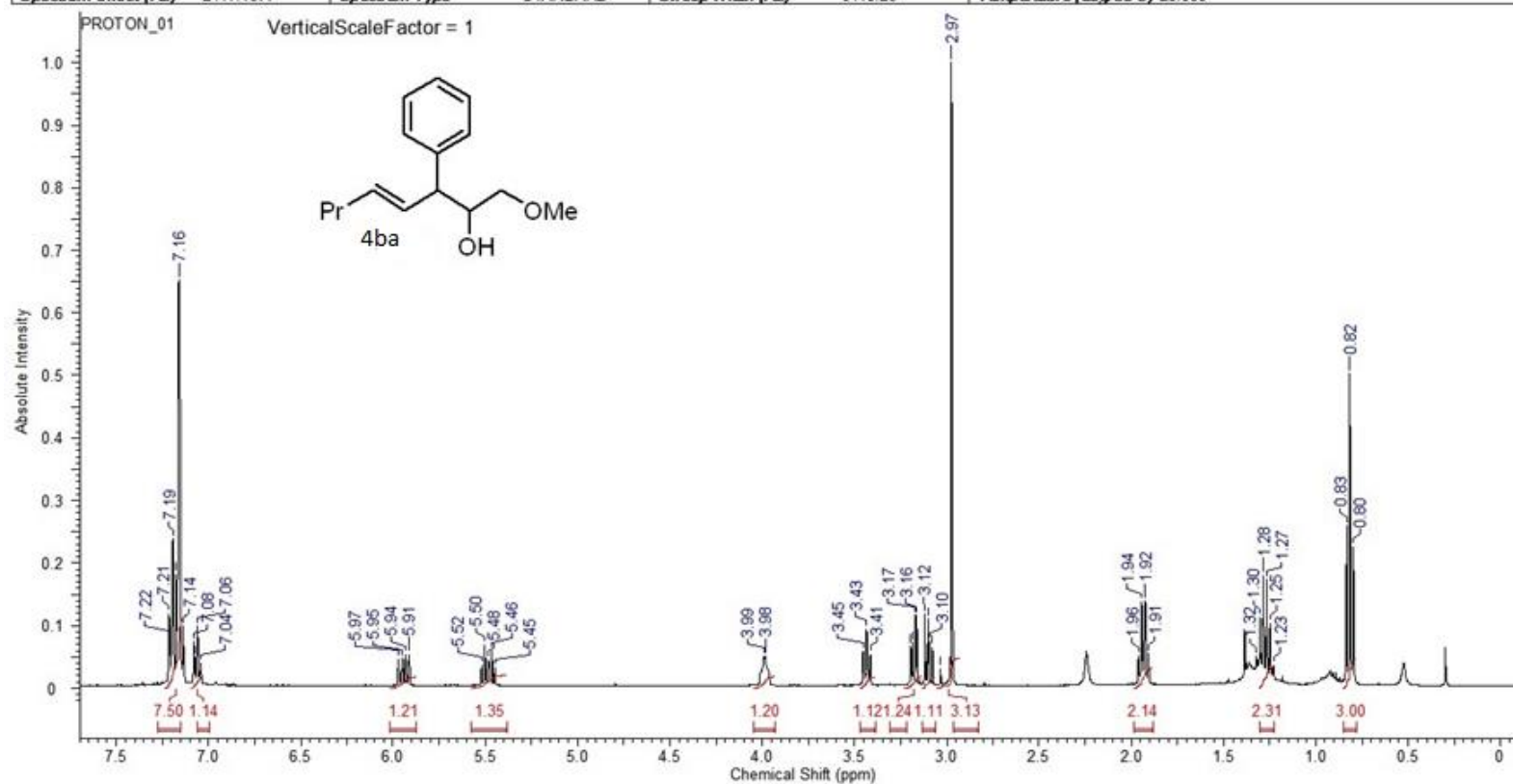
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Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-114-1_1-C		Date	Oct 6 2015	
Date Stamp	Oct 6 2015	File Name	C:\Users\Erman\Desktop\123\ERM-ENEP-114-1_1-C_20151006_01\CARBON_01.fid.fid				
Frequency (MHz)	100.57	Nucleus	13C	Number of Transients	1500	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



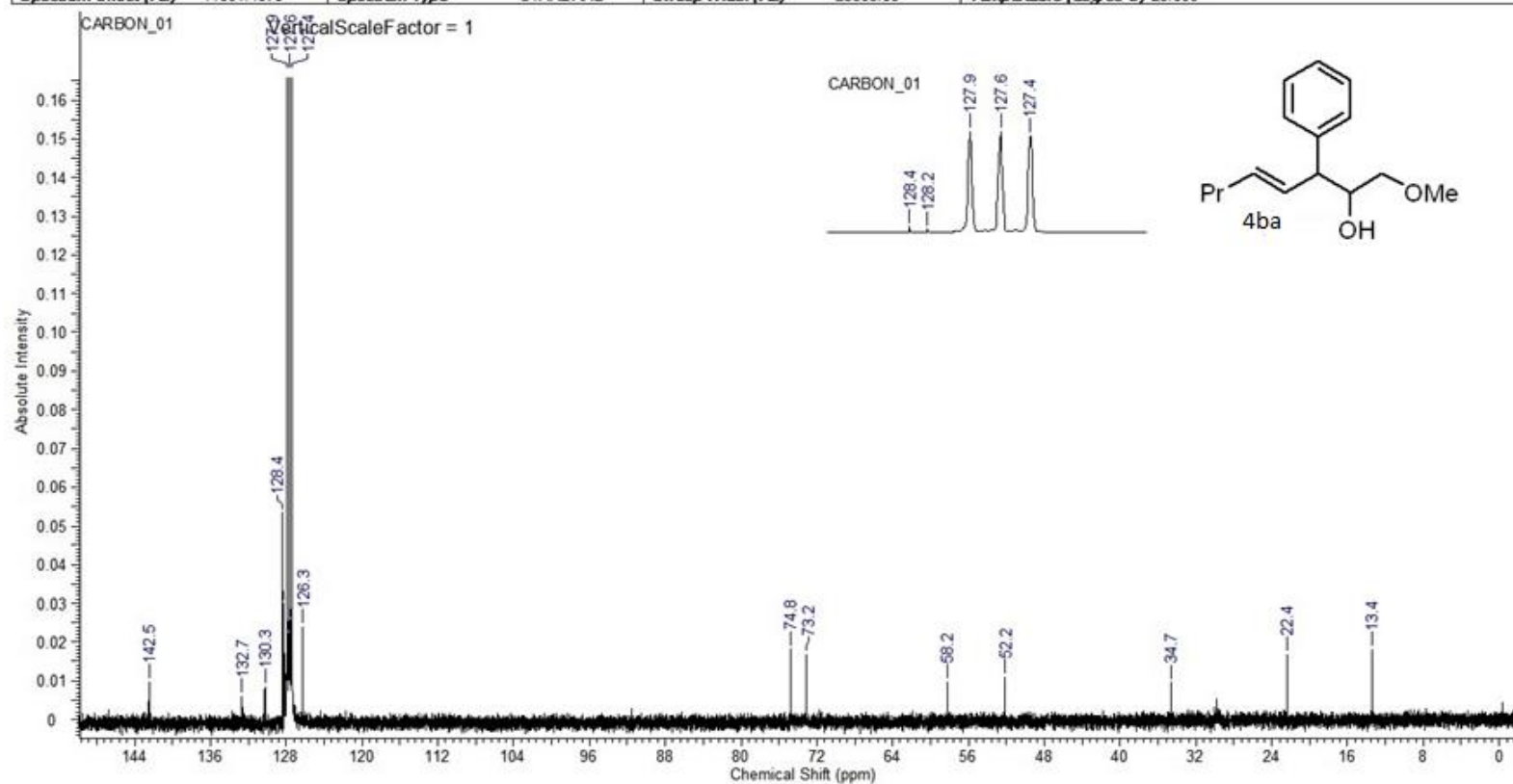
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Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-54-1_1subC6D6	Date	Mar 24 2015
Date Stamp	Mar 24 2015	File Name	C:\Users\Erman\Desktop\ERMAN\ERM-ENEP-54-1_1subC6D6	20150324_01\PROTON_01.fid	
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	32
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	56.00
Spectrum Offset (Hz)	2417.1677	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Original Points Count	16384
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



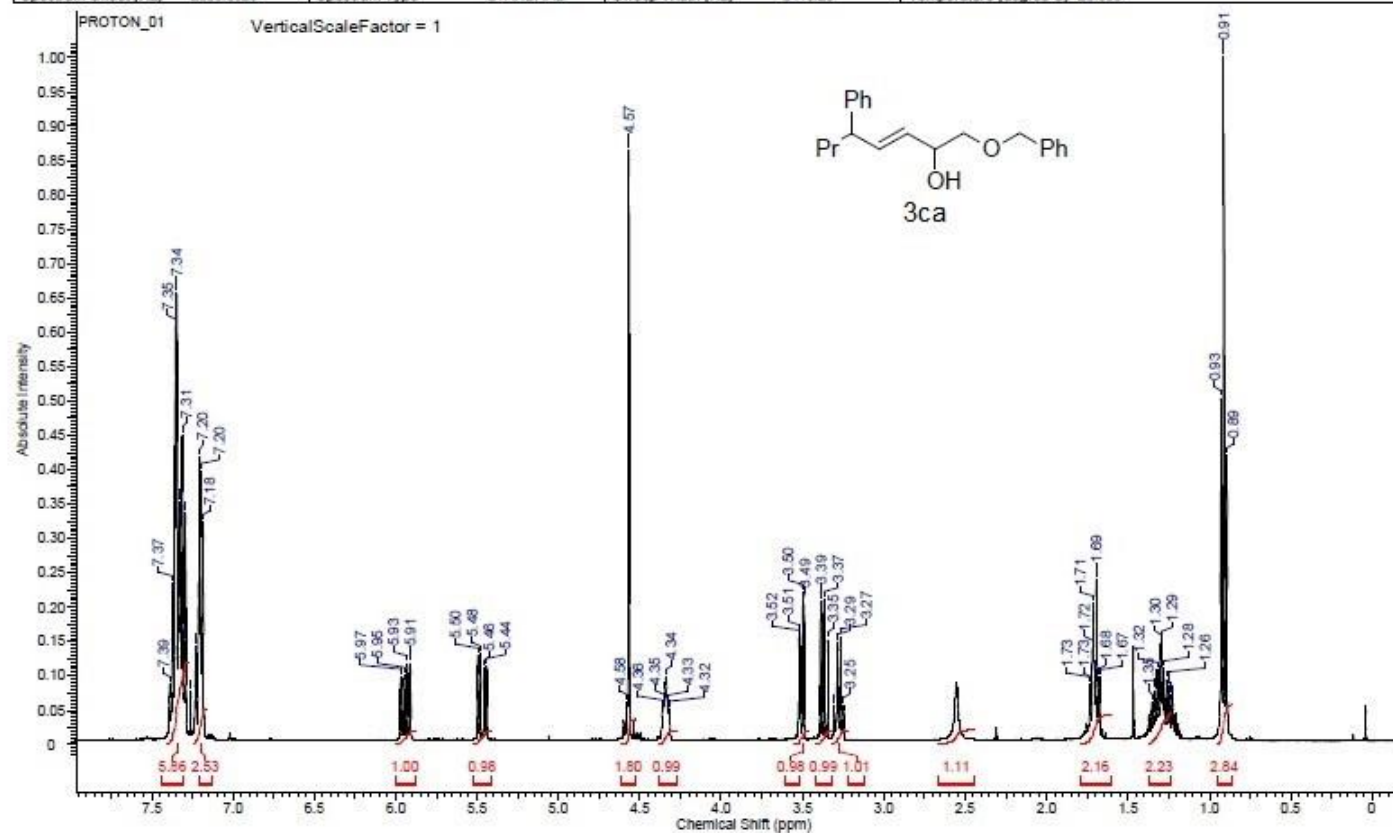
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Date Stamp	Oct 8 2015	File Name	C:\Users\Erman\Desktop\123\ERM-ENEP-114-1_1-BNZ_20151008_03\CARBON_01.fid				
Frequency (MHz)	100.57	Nucleus	13C	Number of Transients	2000	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000

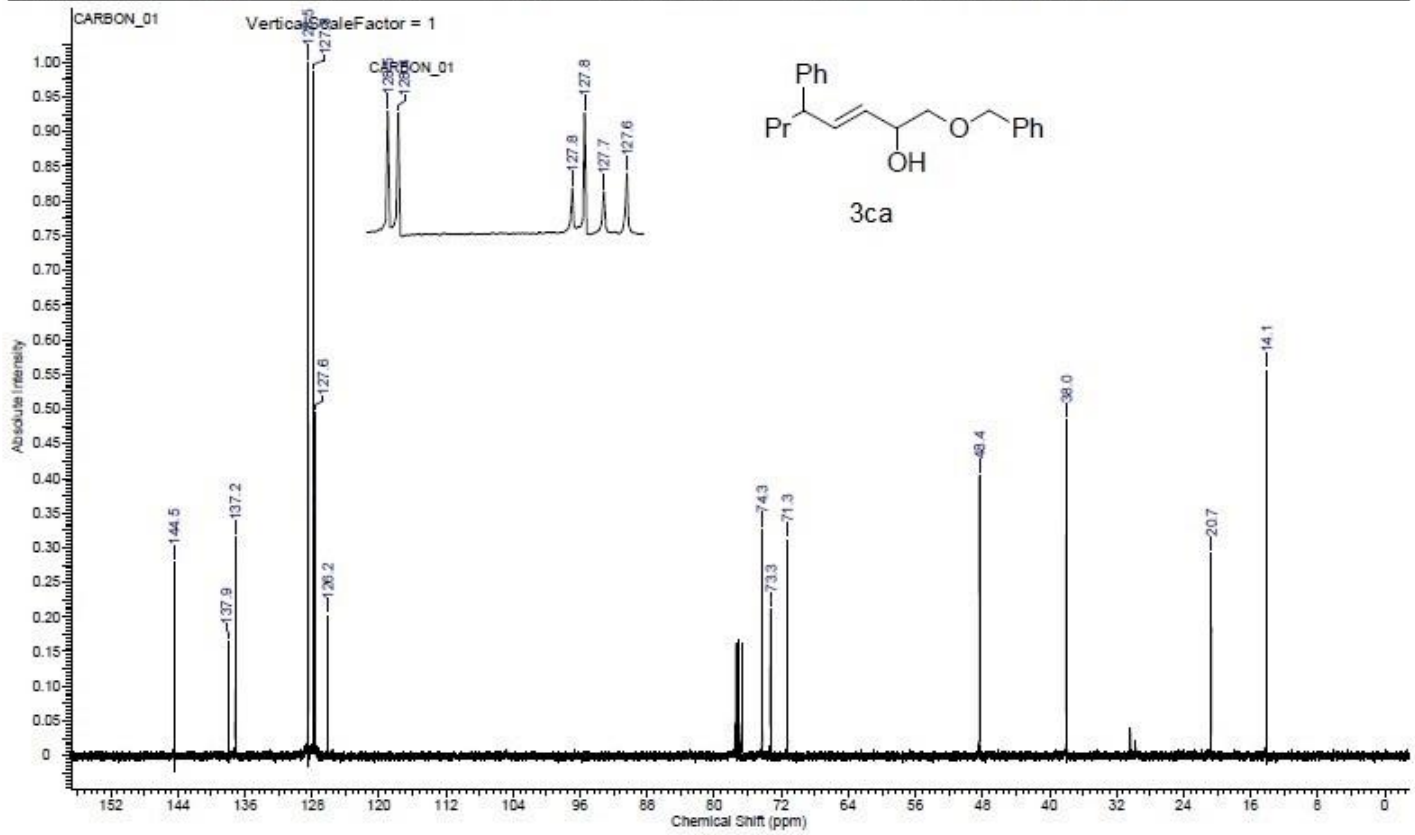


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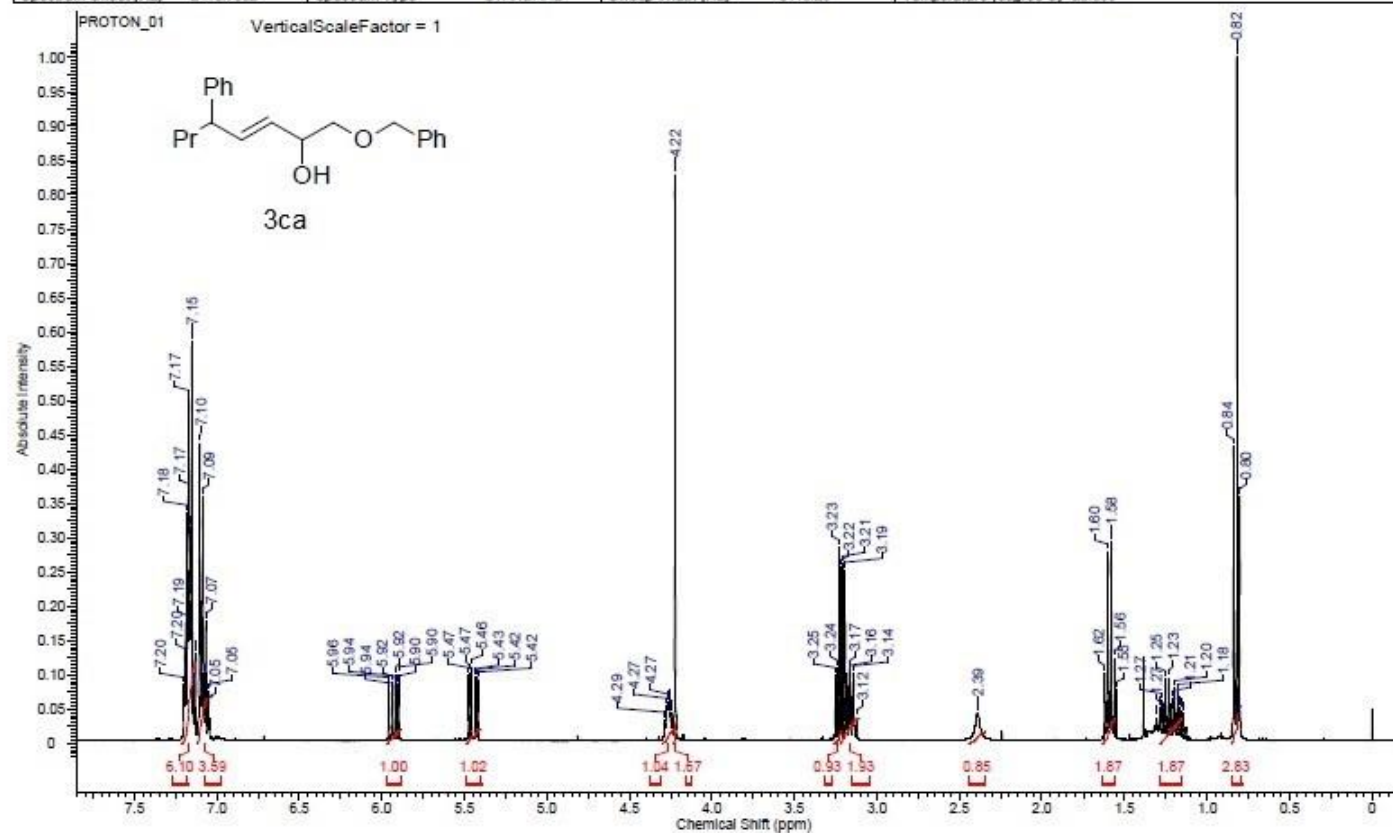
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Date Stamp	Jan 19 2016	File Name	C:\USERS\ERM\GOOGLE DRIVE\ERMAN\ERM-ENEP-185-1_3	PROTON_01.FID\FID	
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	46.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



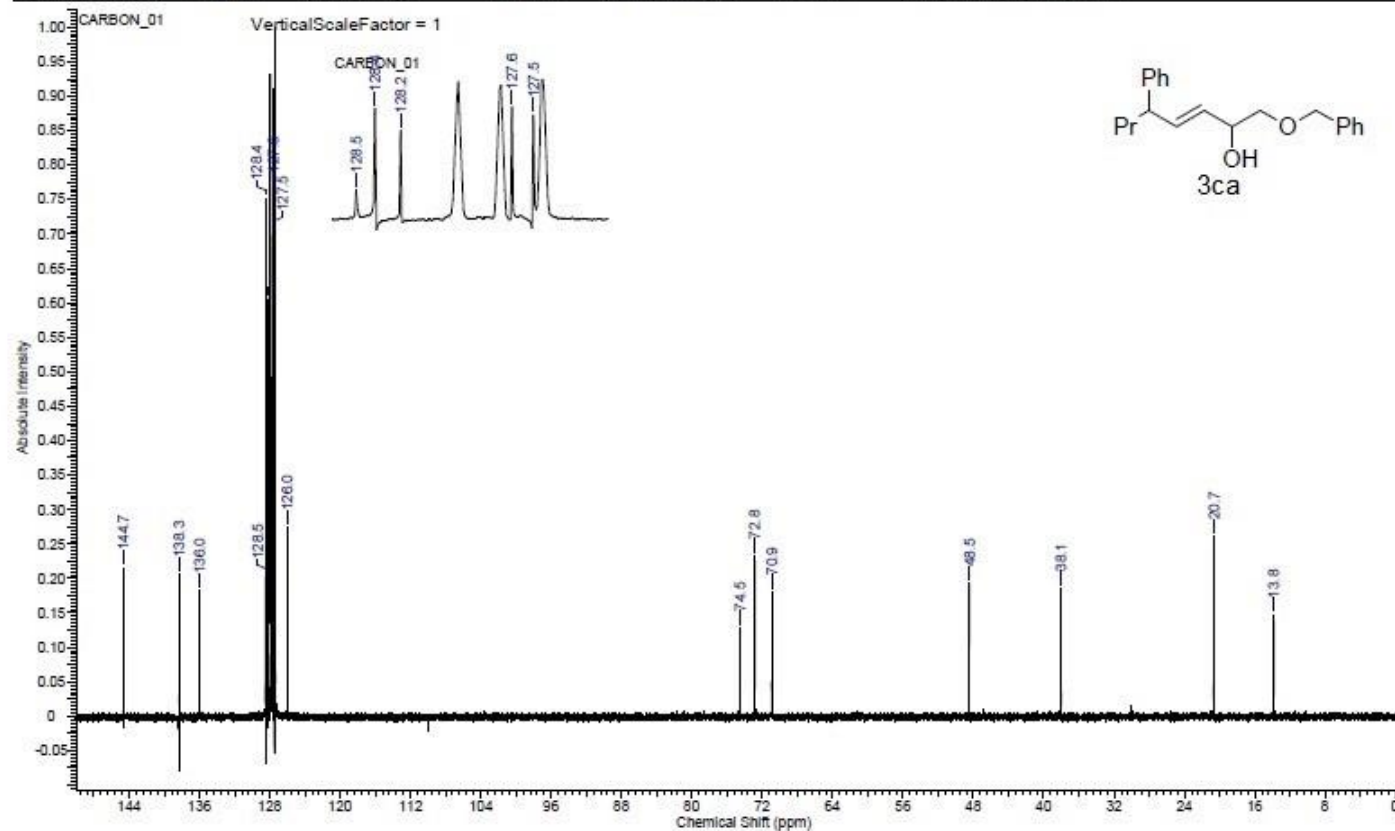
Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-185-1_3		Date	Jan 19 2016	
Date Stamp	Jan 19 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-185-1_3_20160119_01\CARBON_01.FID\FID		Original Points Count	32768	
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256		
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



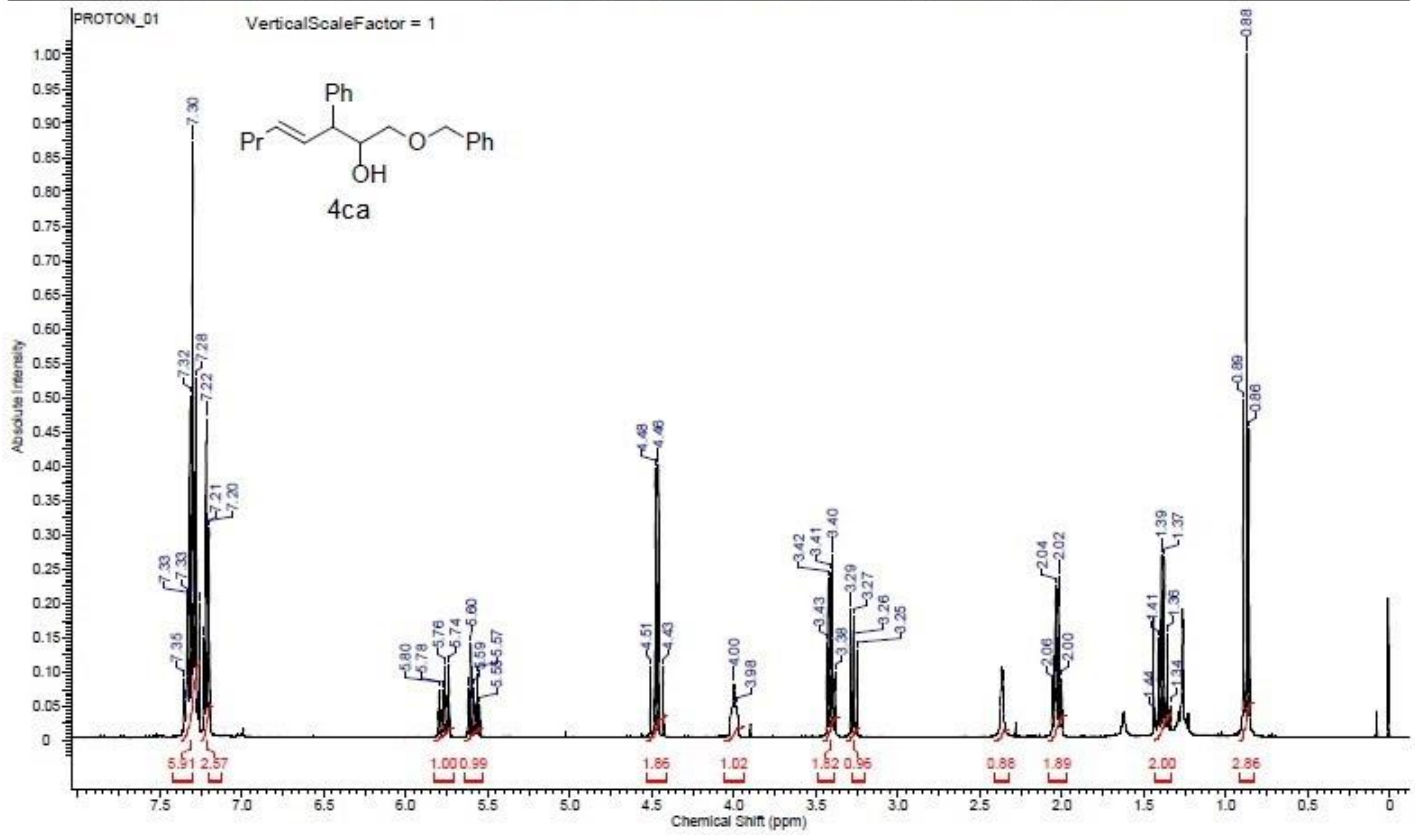
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Date Stamp	Jan 8 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-185-1_3_20160108_01\PROTON_01.FID\FID	Original Points Count	16384
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	32
Points Count	16384	Pulse Sequence	s20u1	Receiver Gain	52.00
Spectrum Offset (Hz)	2416.7082	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



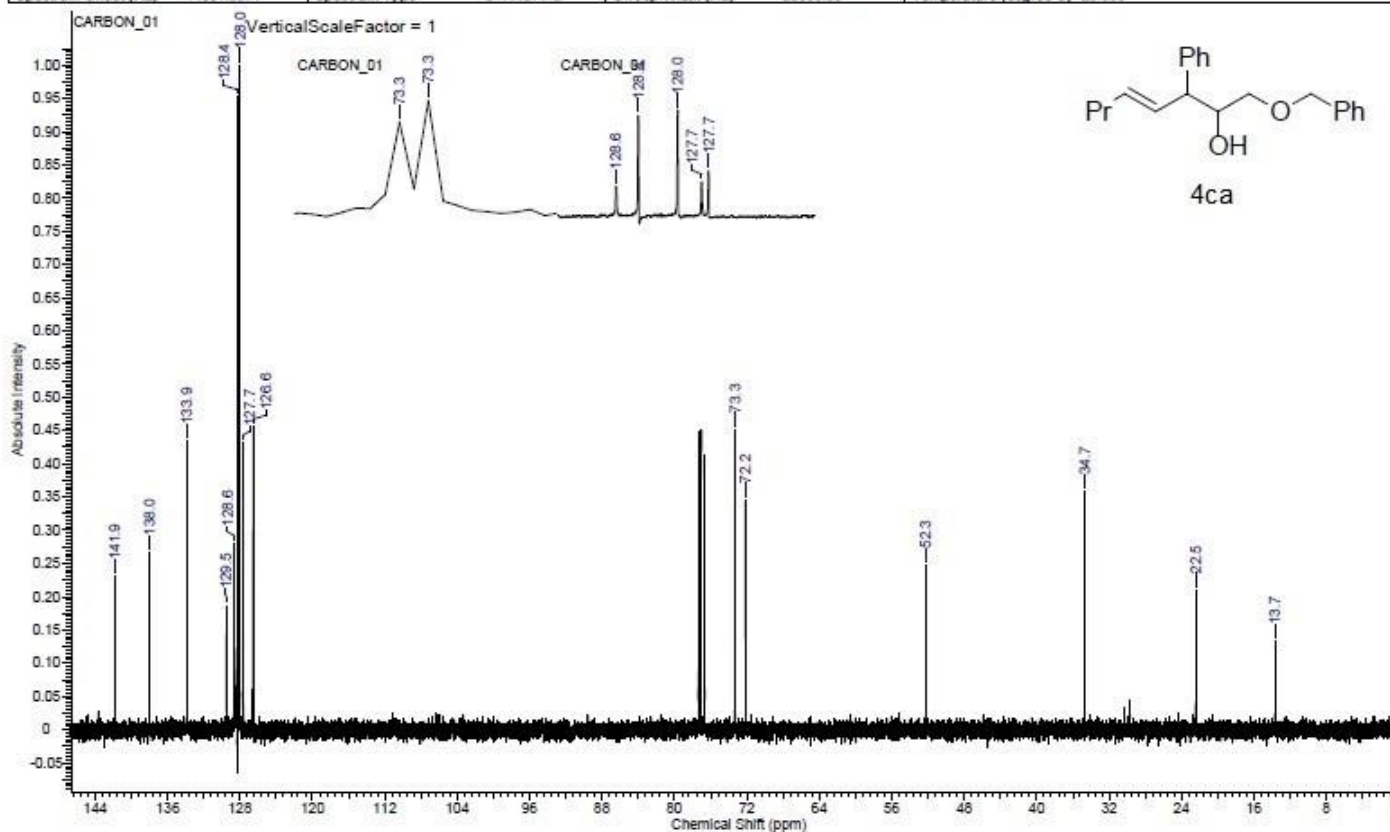
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Date Stamp	Jan 11 2016	File Name	C:\USERS\ERM\GOOGLE DRIVE\ERMAN\ERM-ENEP-185-1_3_20160108_01\CARBON_01.FID\FID		
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2bul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



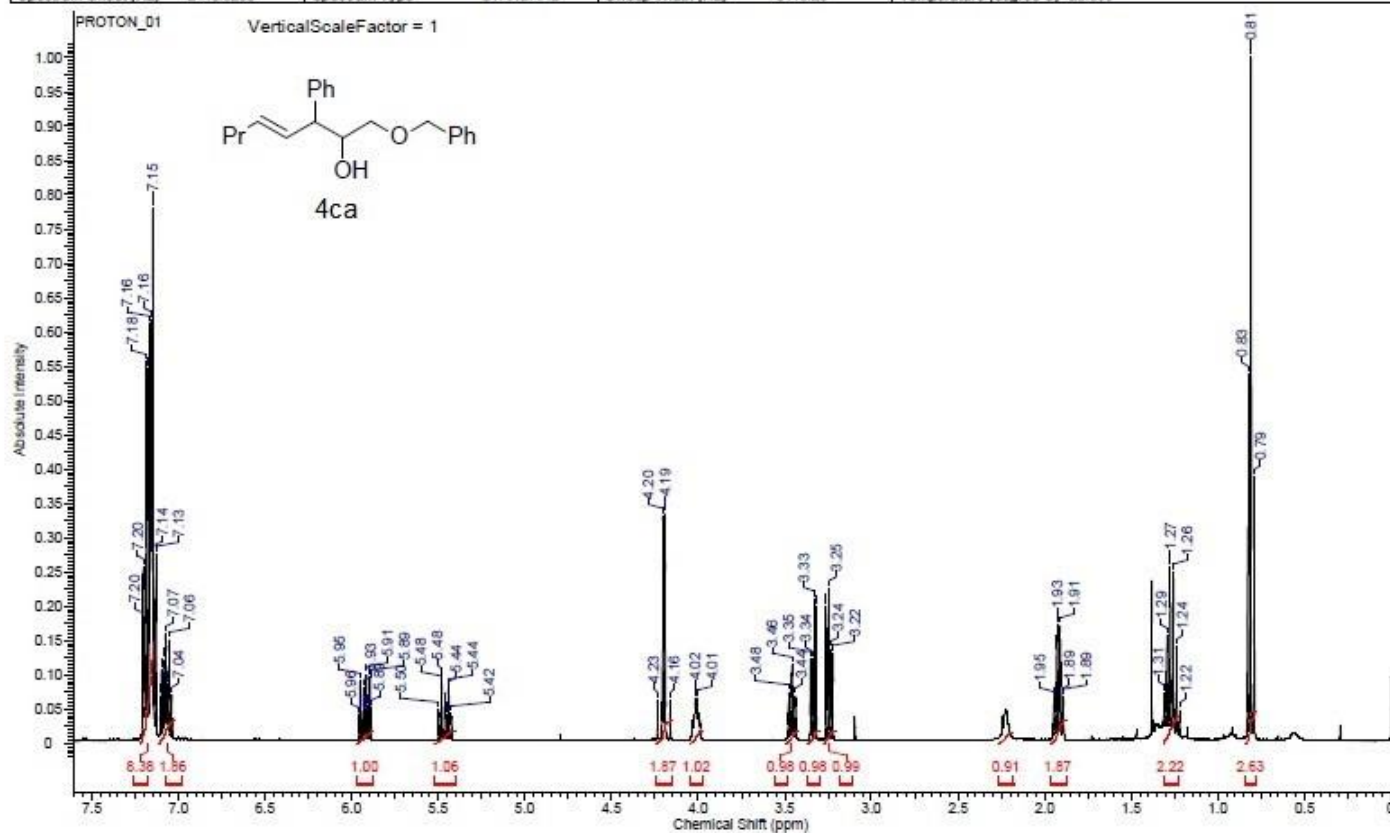
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Date Stamp	Jan 19 2016	File Name	C:\USERS\ERM\GOOGLE DRIVE\ERM\ERM-ENEP-185-1.1	PROTON_01.FID\FID	
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	56.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



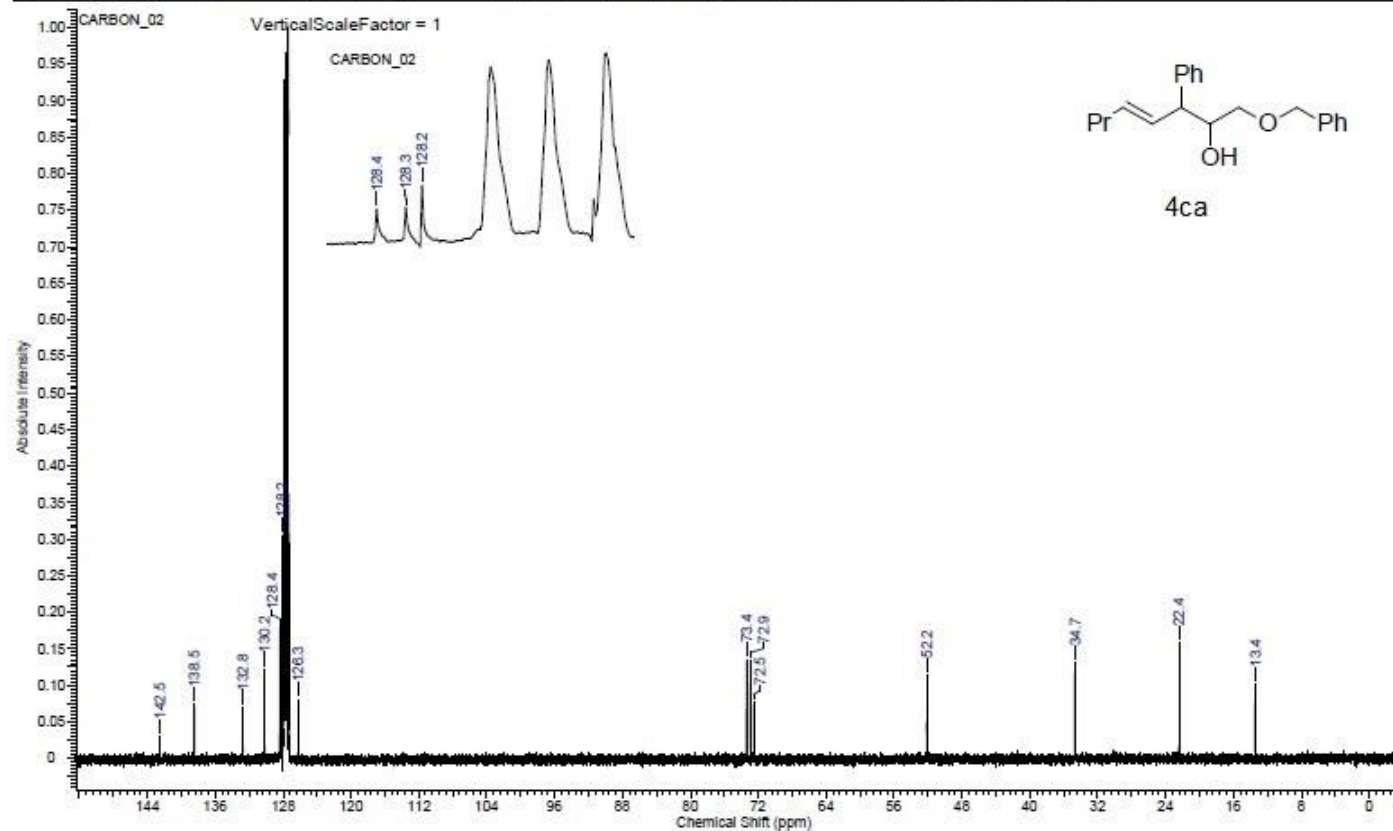
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Date Stamp	Jan 19 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-185-1_1_20160119_01\CARBON_01.FID\FID	Original Points Count	32768
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



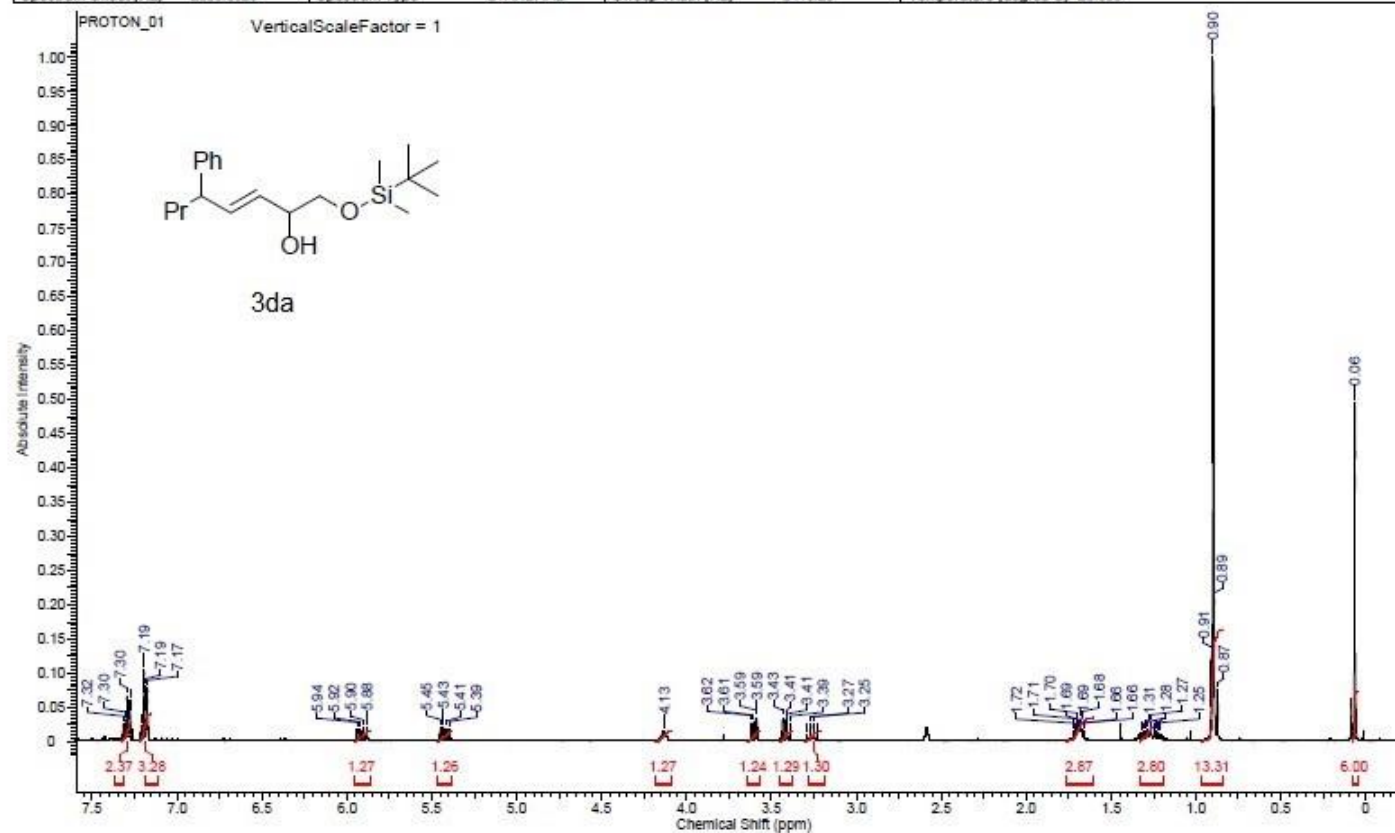
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Date Stamp	Jan 8 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-185-1.1	20160108_01\PROTON_01.FID\FID	
Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	32
Points Count	16384	Pulse Sequence	s20u1	Receiver Gain	58.00
Spectrum Offset (Hz)	2415.9268	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Original Points Count	16384
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



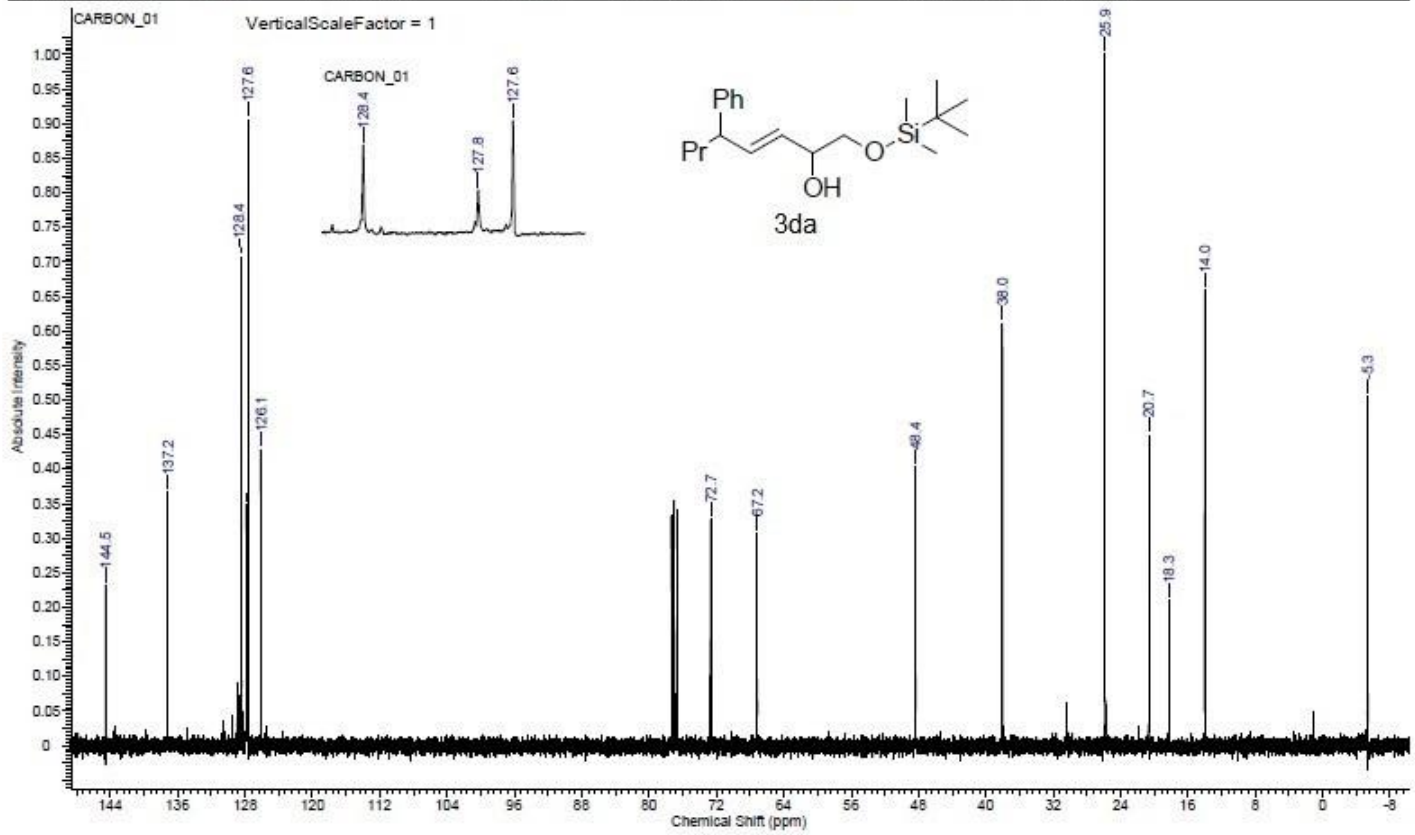
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	512
Points Count	32768	Pulse Sequence	s2bul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



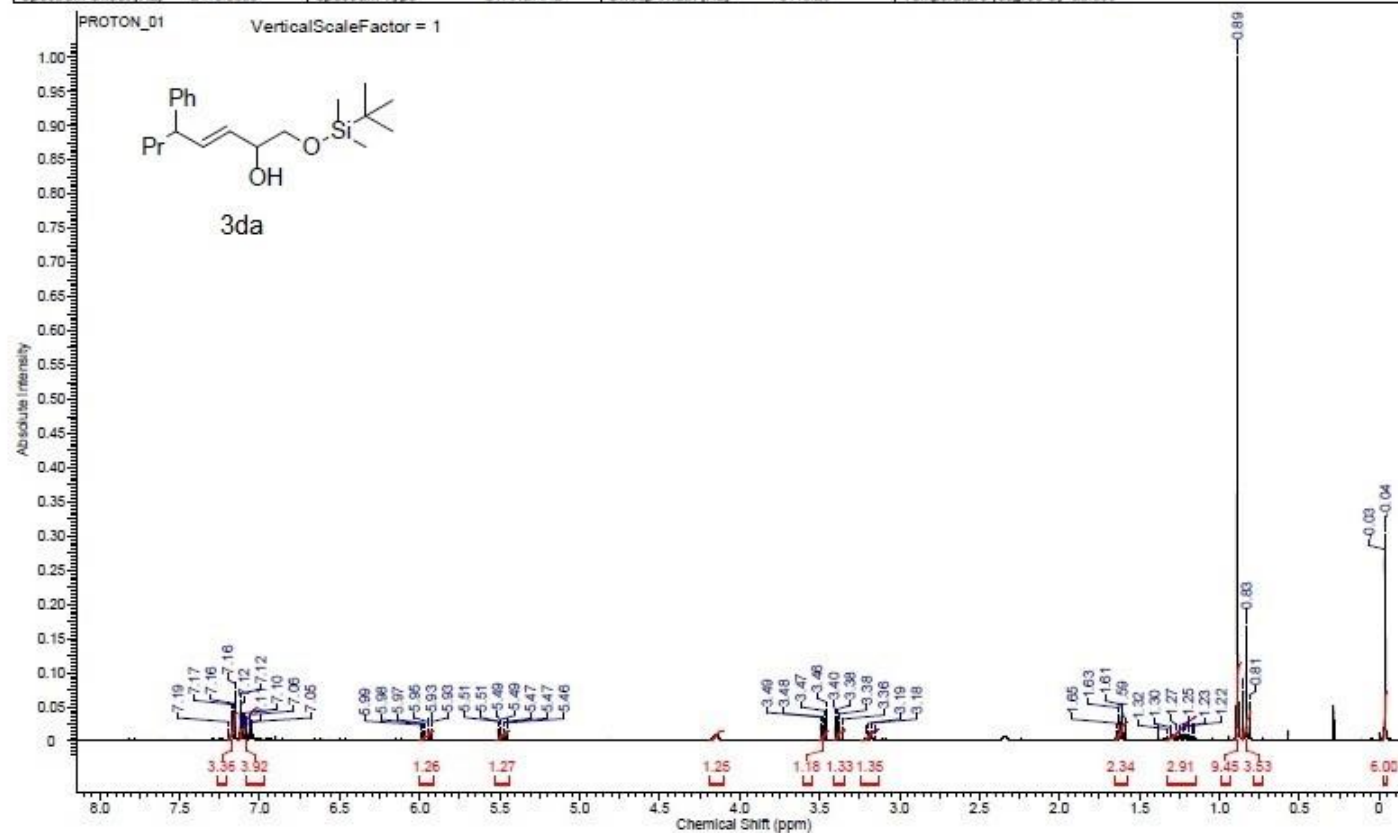
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-184-1_3	Date	Jan 21 2016
Date Stamp	Jan 21 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-184-1_3_20160121_01\PROTON_01.FID\FID	Original Points Count	16384
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	48.00
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



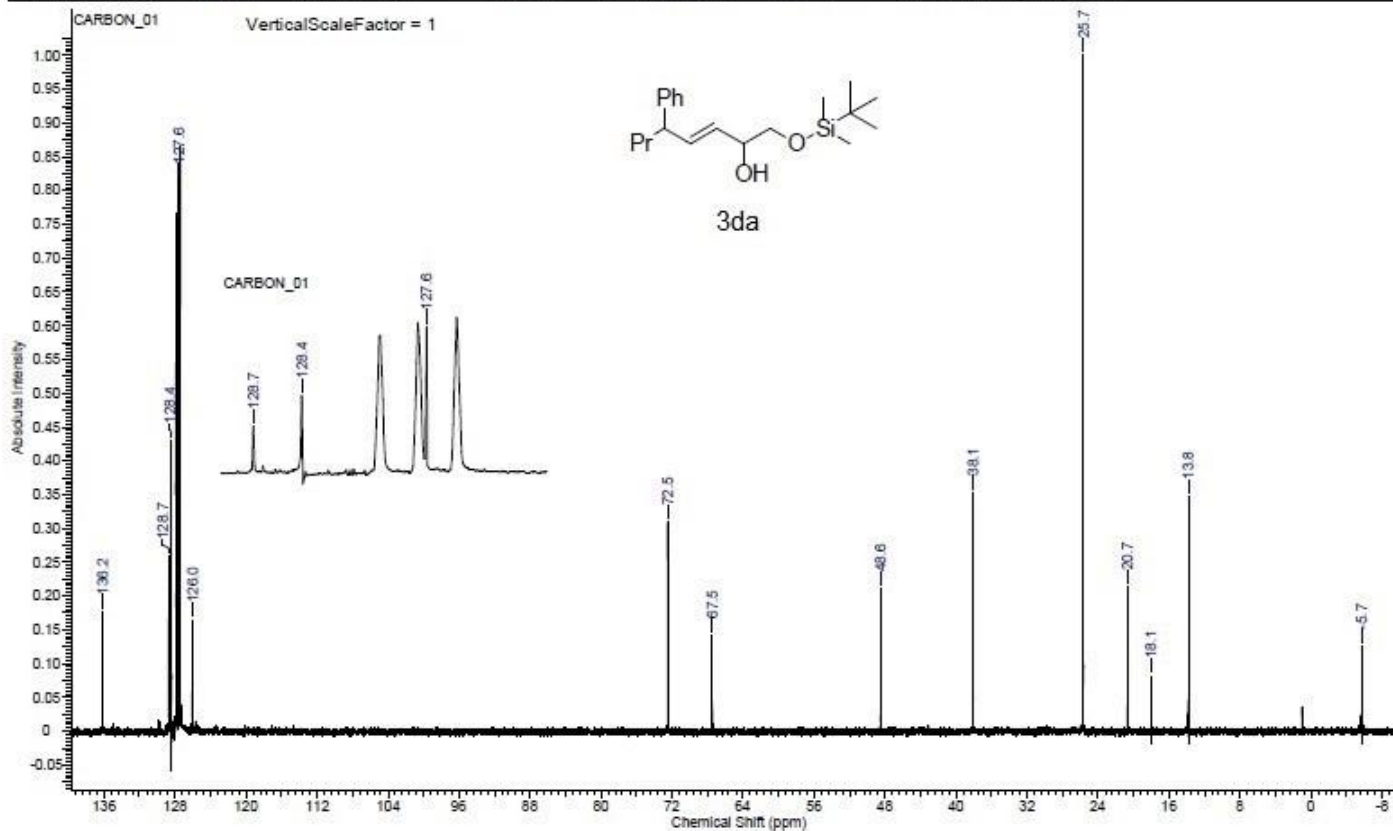
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Date Stamp	Jan 21 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-184-1_3-13C_20160121_01\CARBON_01.FID\FID	Original Points Count	32768
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	160
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



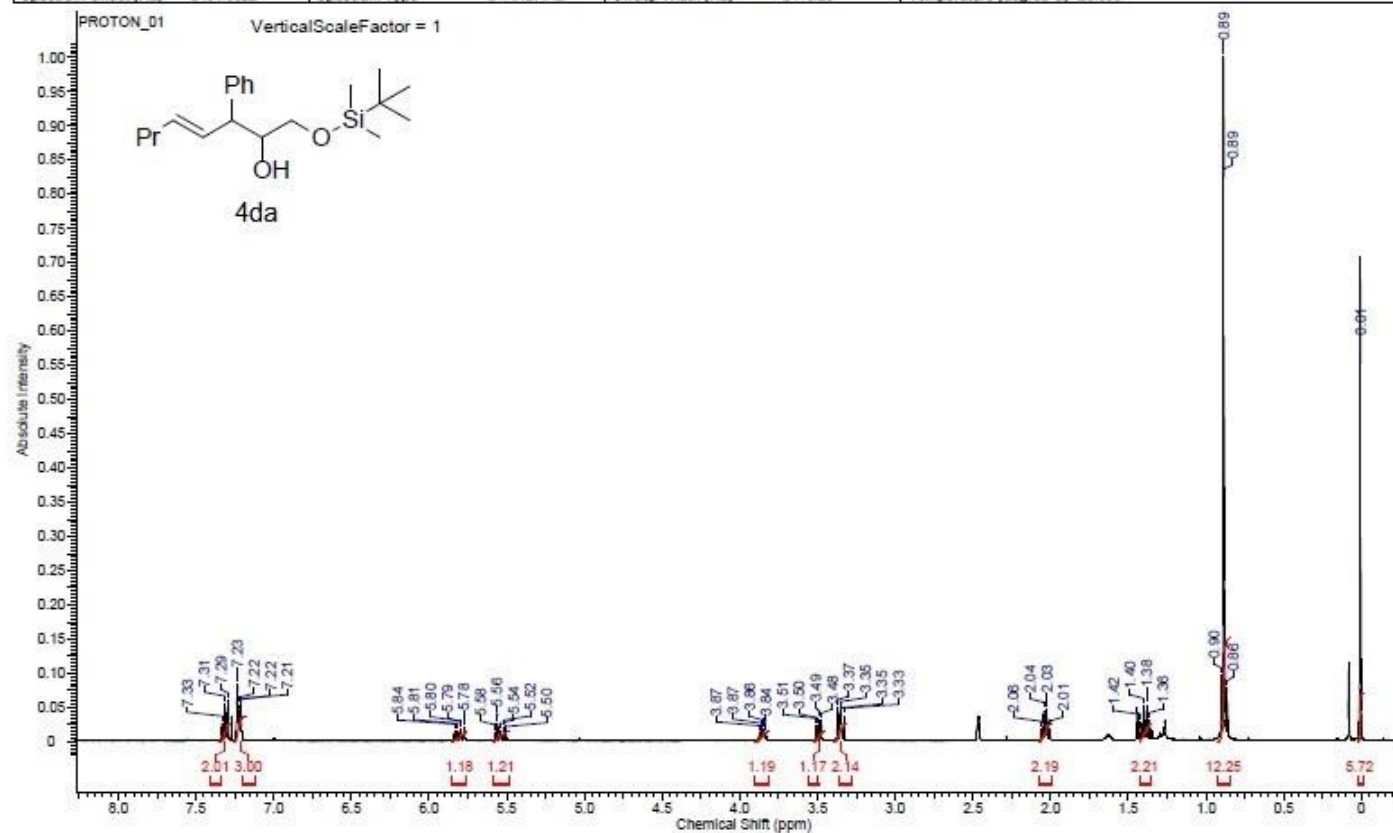
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-184-1_3	Date	Jan 18 2016
Date Stamp	Jan 18 2016	File Name	C:\USERS\ERMAN\GOOGLE DRIVE\ERMAN\ERM-ENEP-184-1_3	20160118_01\PROTON_01.FID\FID	
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s20uI	Receiver Gain	48.00
Spectrum Offset (Hz)	2416.3853	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Original Points Count	16384
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



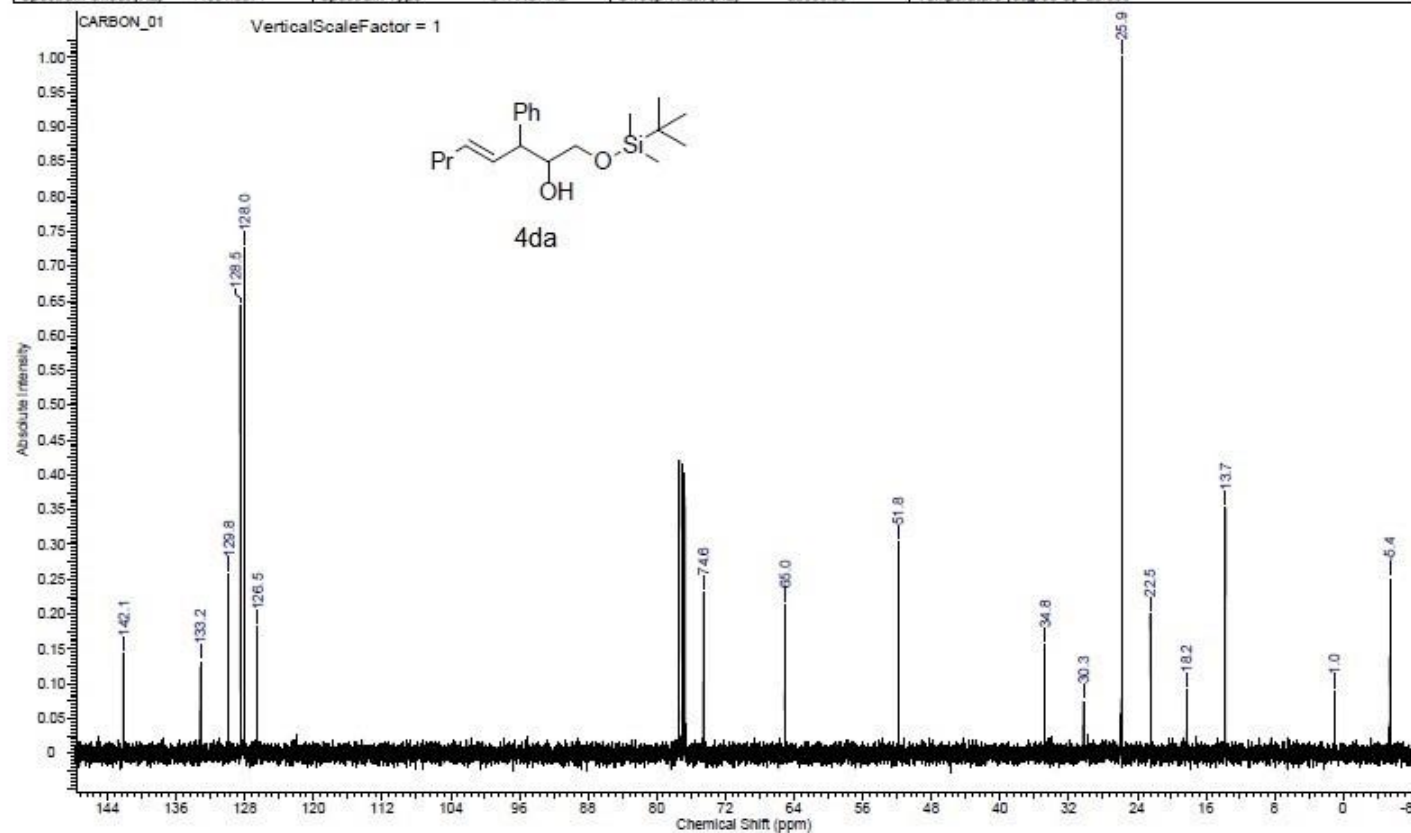
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2bul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



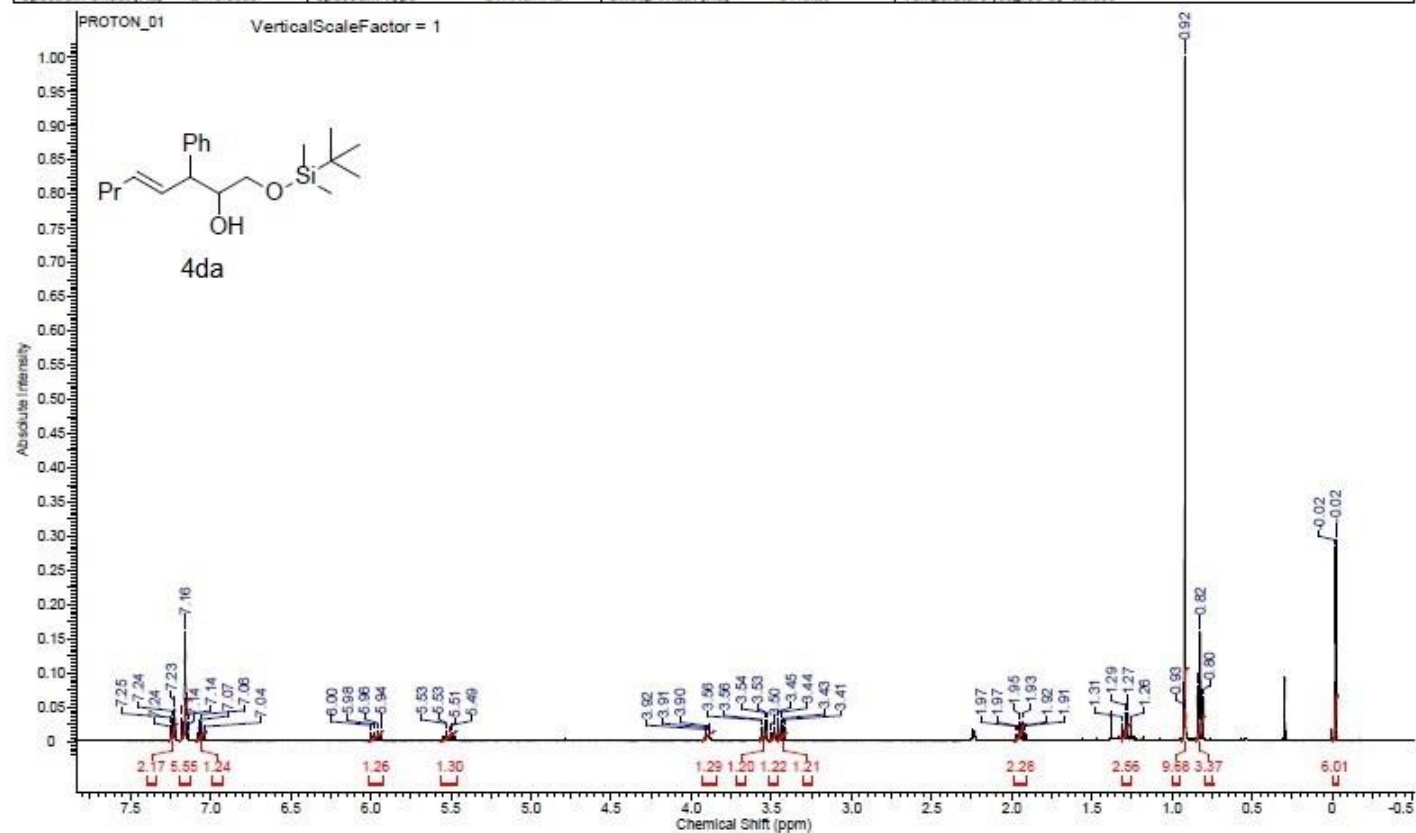
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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	56.00
Spectrum Offset (Hz)	2404.0322	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	CHLOROFORM-d
				Temperature (degree C)	25.000



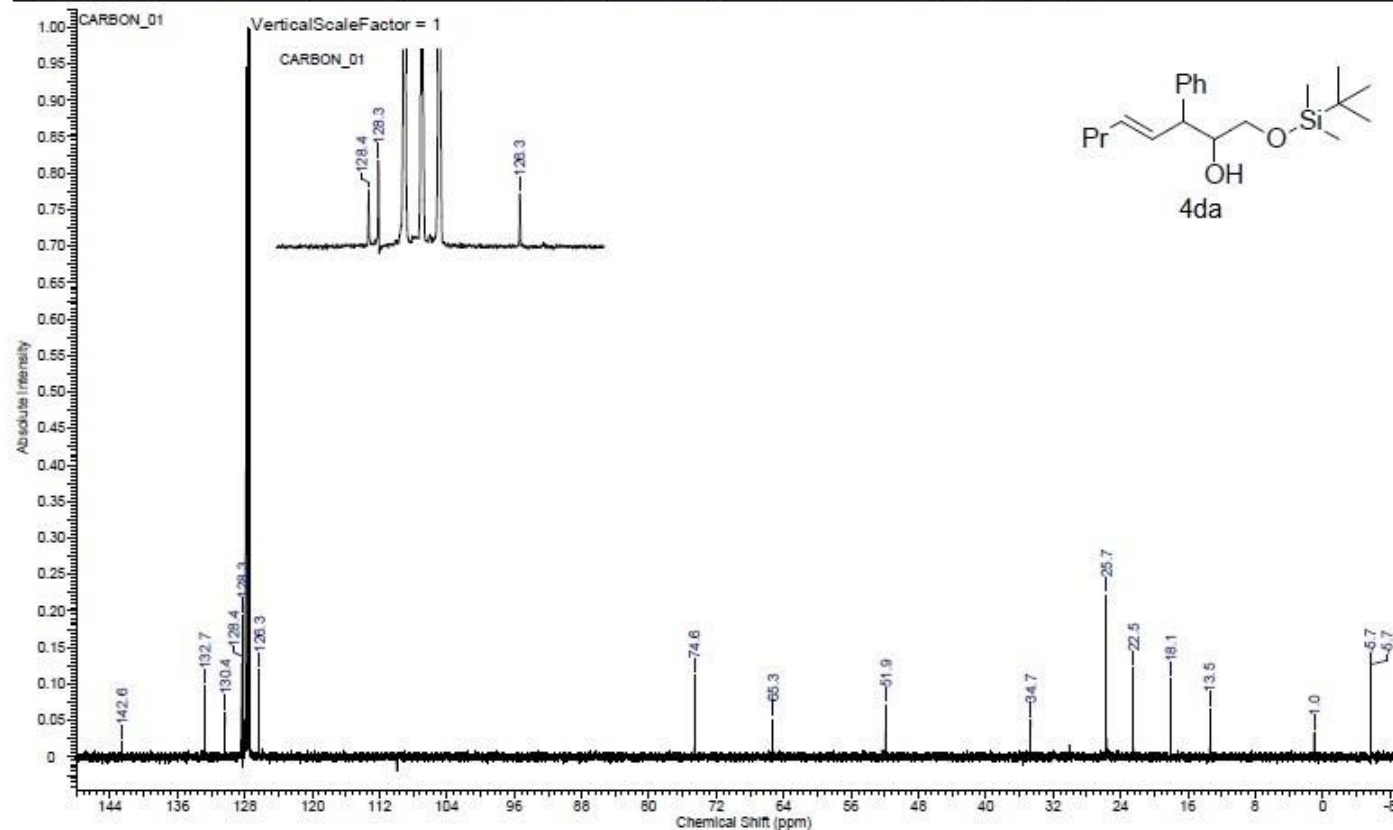
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256		
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



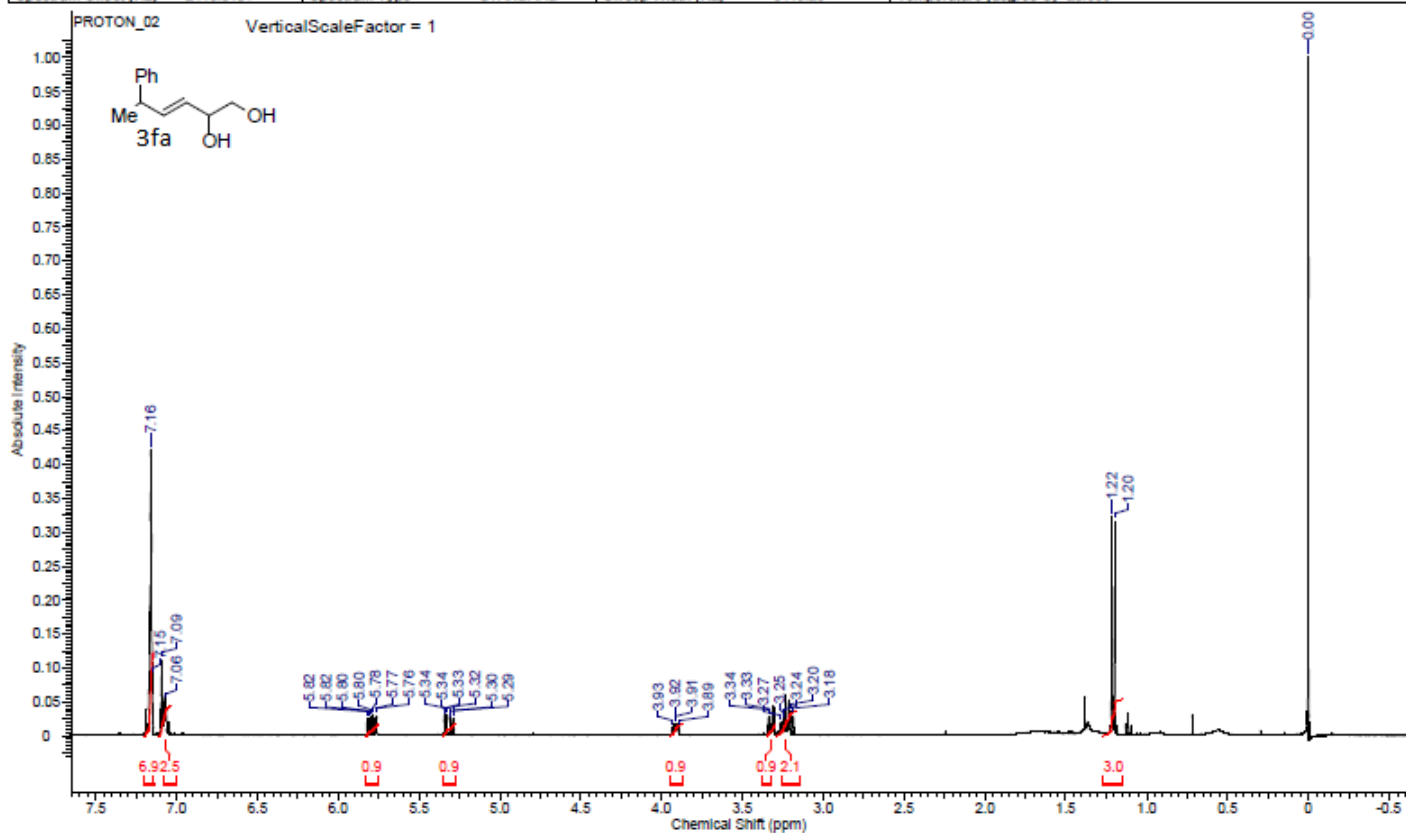
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Frequency (MHz)	399.92	Nucleus	1H	Number of Transients	8
Points Count	16384	Pulse Sequence	s20u1	Receiver Gain	58.00
Spectrum Offset (Hz)	2416.3853	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



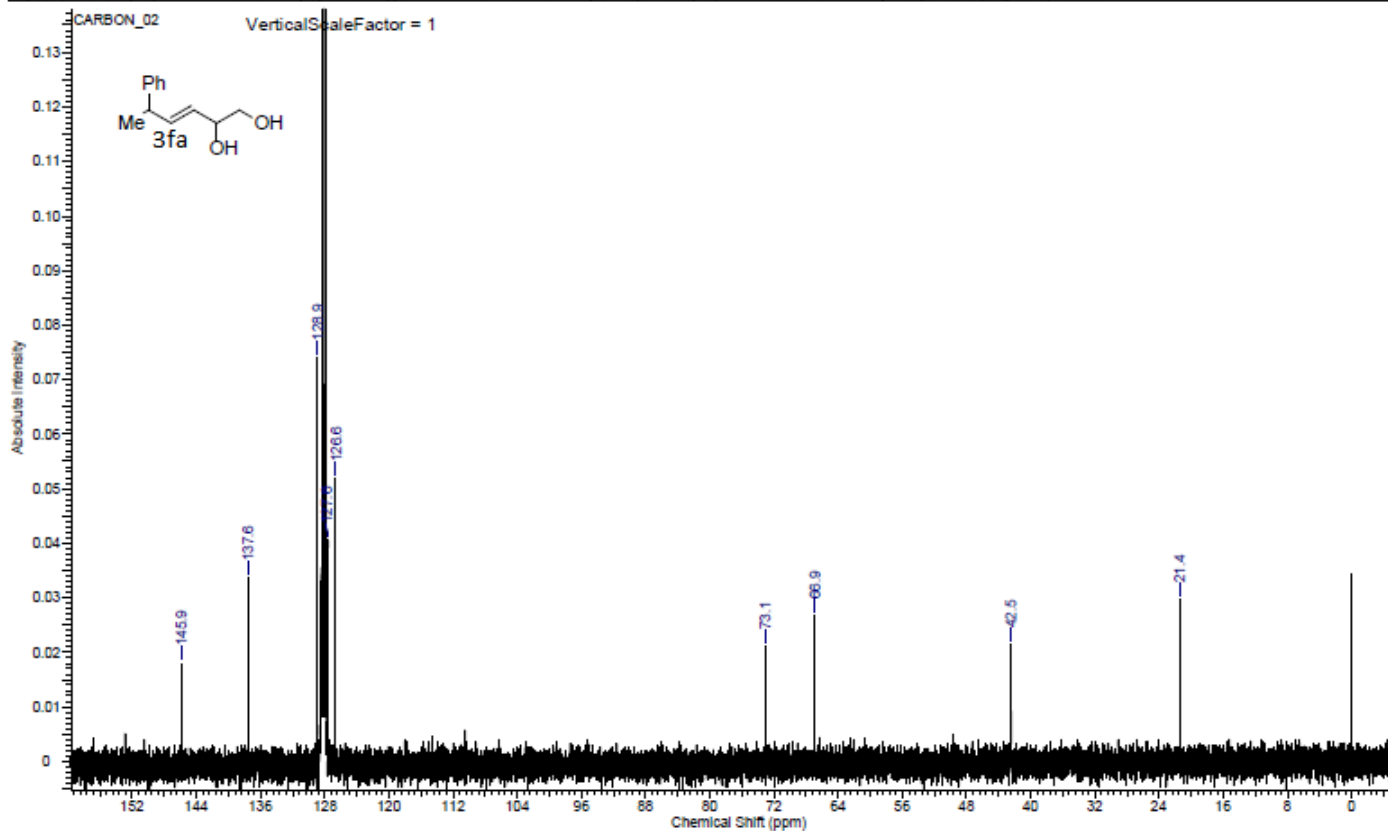
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2bul	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



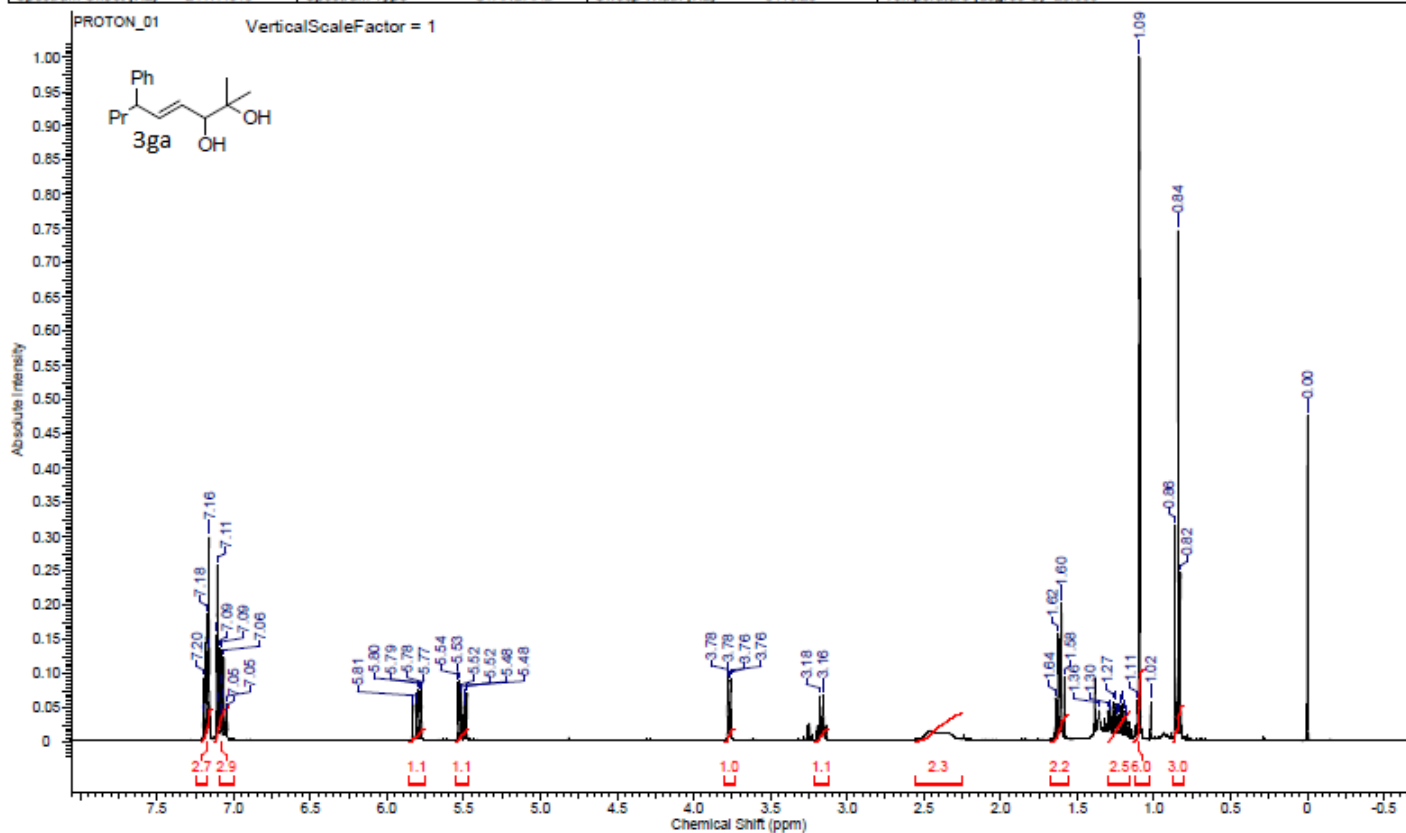
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Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s20u1	Receiver Gain	60.00
Spectrum Offset (Hz)	2416.3181	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



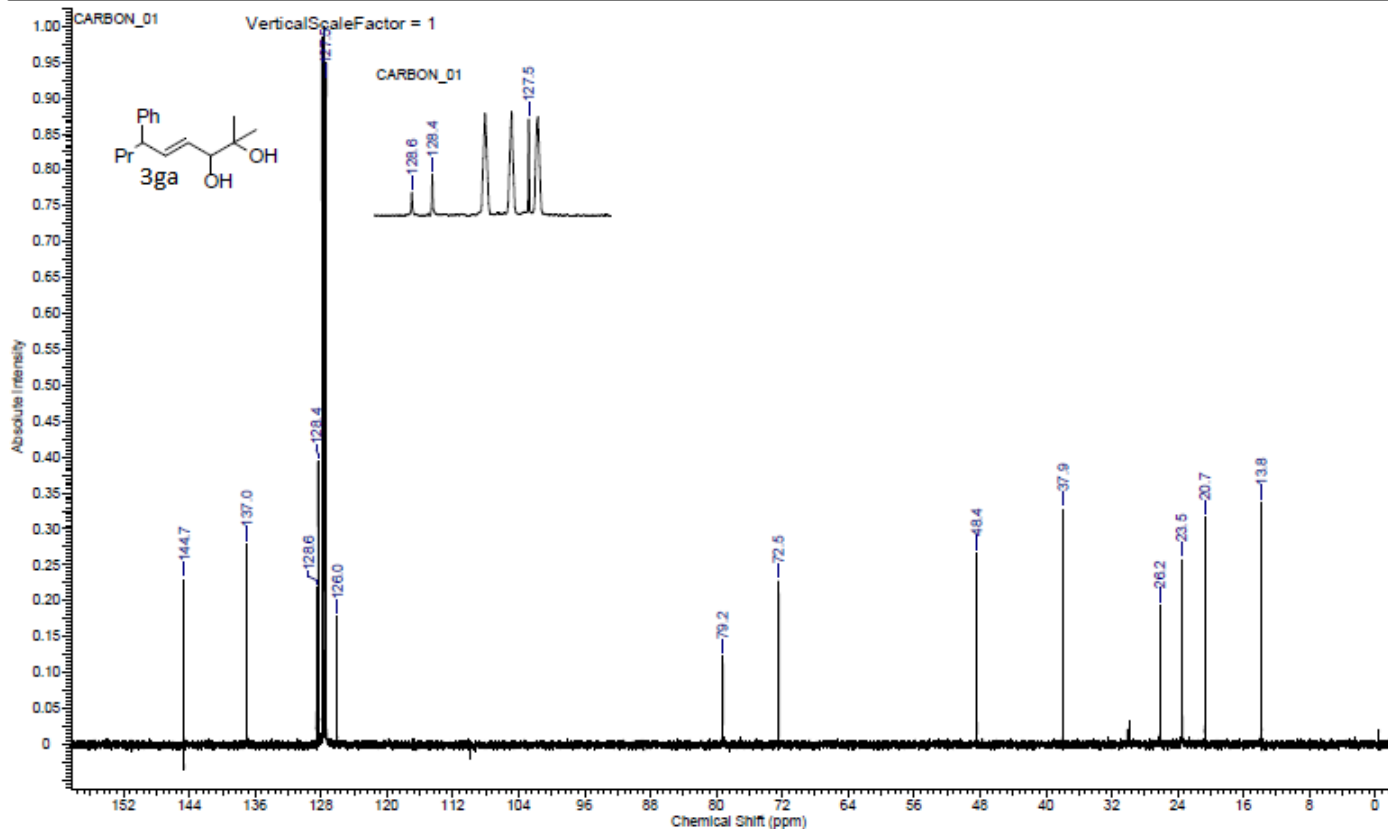
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Date Stamp	Jun 7 2016	File Name	C:\USERS\ERMAN\DESKTOP\WET\ERM-ENEP-215-1_3_20160607_01\CARBON_02\FID\FID				
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	700	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2piJ	Receiver Gain	30.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	11111.2148	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



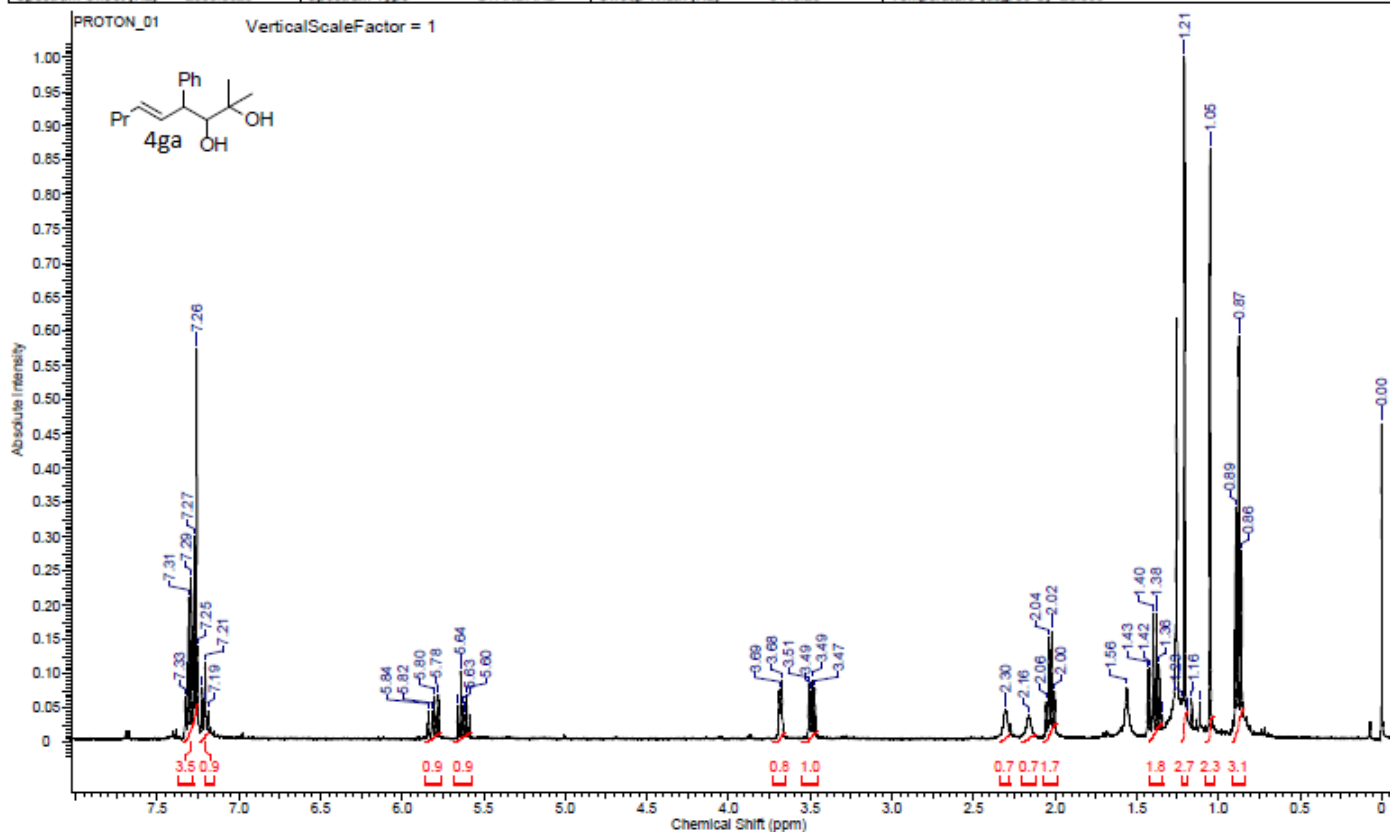
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Date Stamp	Jun 13 2016	File Name	C:\USERS\ERMAN\DESKTOP\WET\ERM-ENEP-217-1_3_20160613_01\PROTON_01.FID\FID				
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	52.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	2417.4919	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



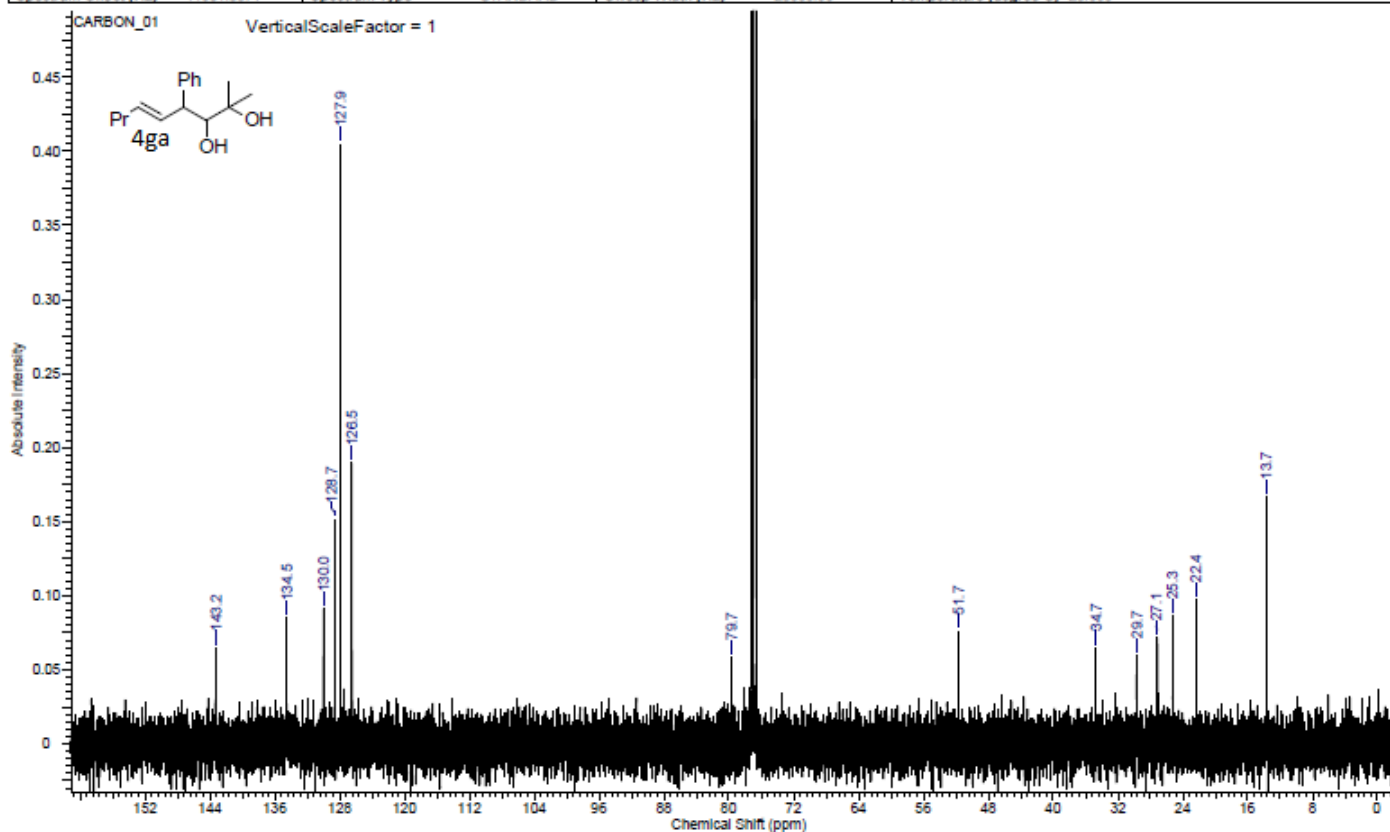
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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2nuJ	Receiver Gain	30.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



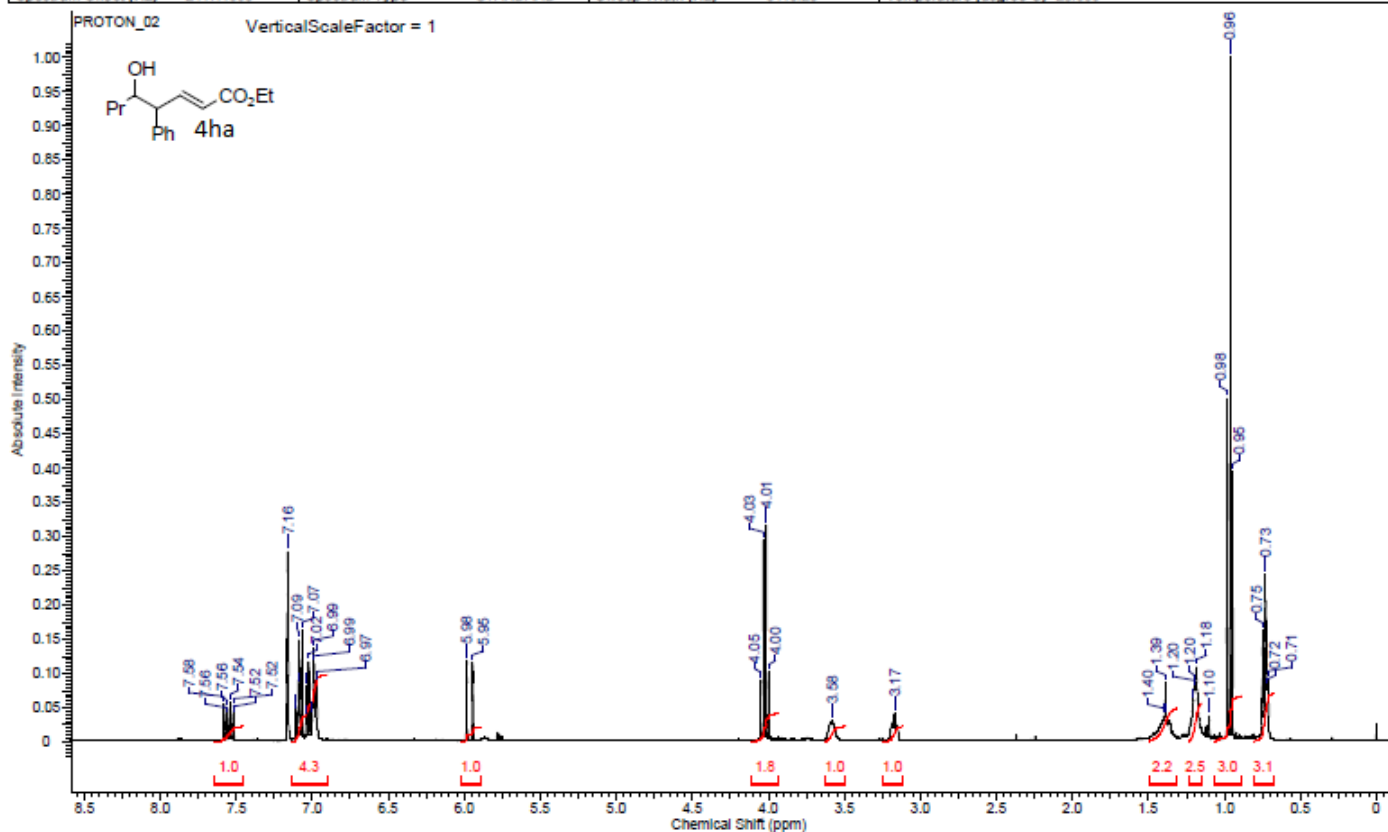
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Date Stamp	Jun 13 2016	File Name	C:\USERS\ERMANDESKTOP\WET\ERM-ENEP-217-1_1_20160613_01\PROTON_01\FID\FID				
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2nu1	Receiver Gain	60.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



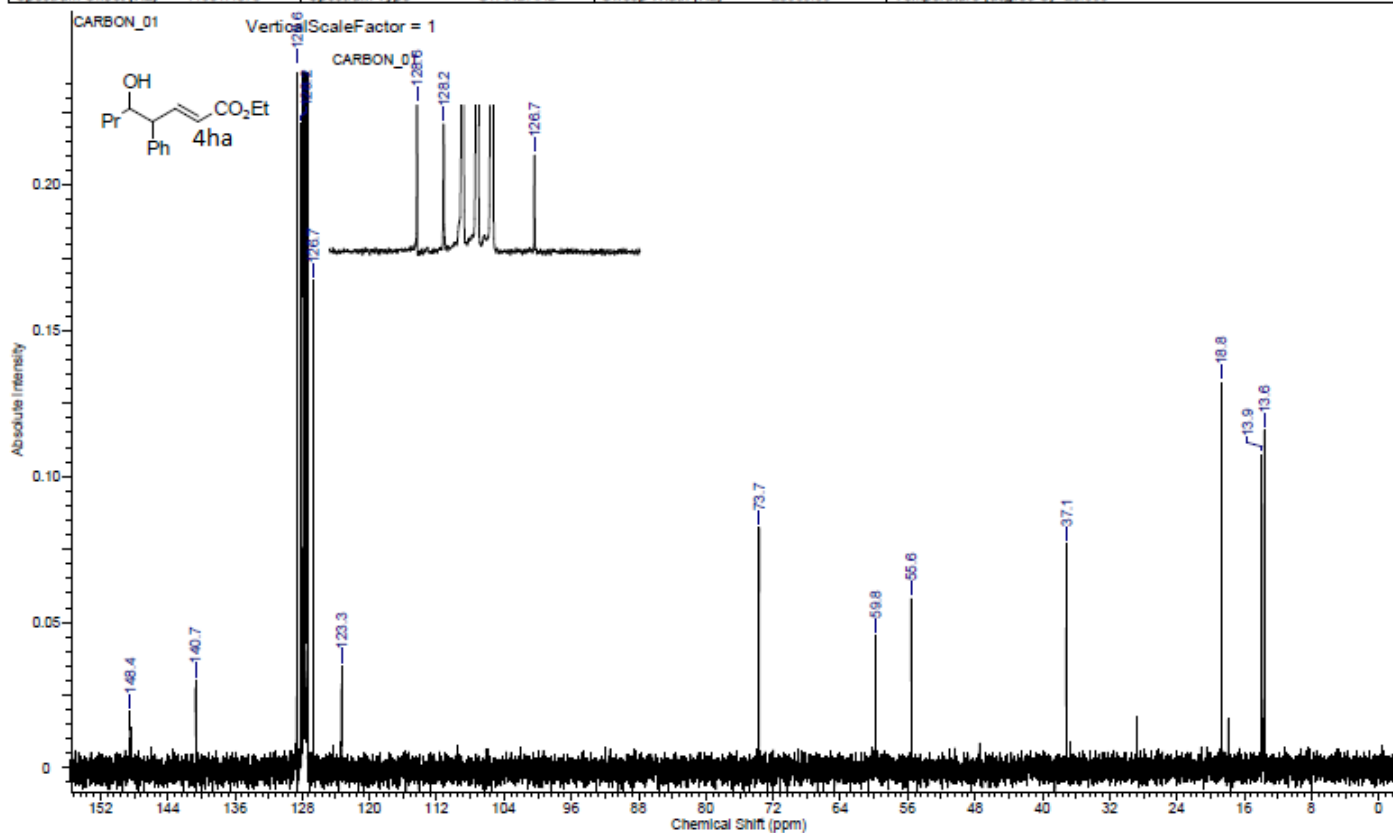
Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-217-1_1	Date	Jun 13 2016
Date Stamp	Jun 13 2016	File Name	C:\USERS\ERMAN\DESKTOP\WET\ERM-ENEP-217-1_1_20160613_01\CARBON_01.FID\FID		
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	512
Points Count	32768	Pulse Sequence	s2pu1	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



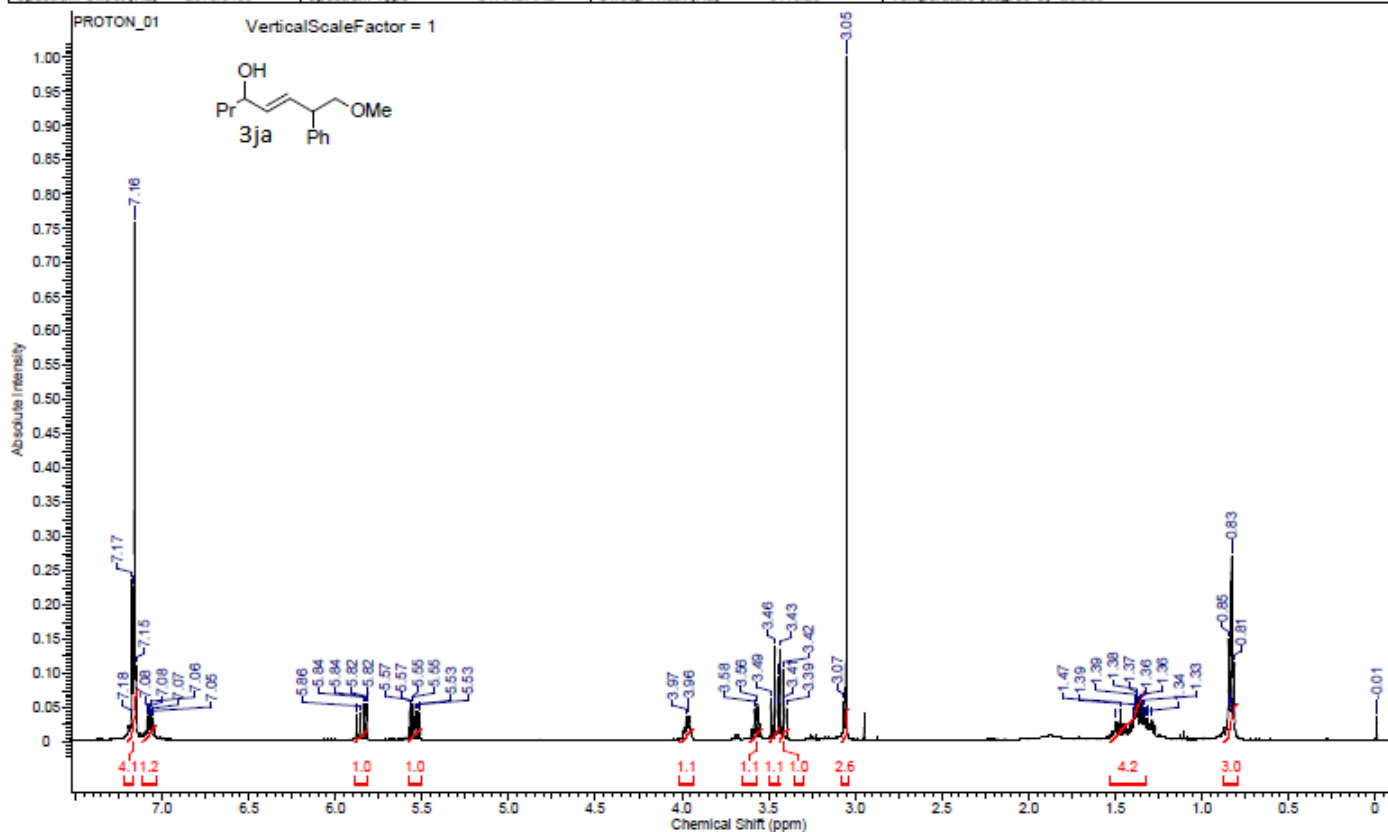
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-214-1_3		Date	May 26 2016	
Date Stamp	May 26 2016	File Name	C:\USERS\ERMANDESKTOPIWET\ERM-ENEP-214-1_3_20160526_01\PROTON_02\FID\FID				
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2pul	Receiver Gain	60.00	Solvent	BENZENE-d6
Spectrum Offset (Hz)	2417.1006	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



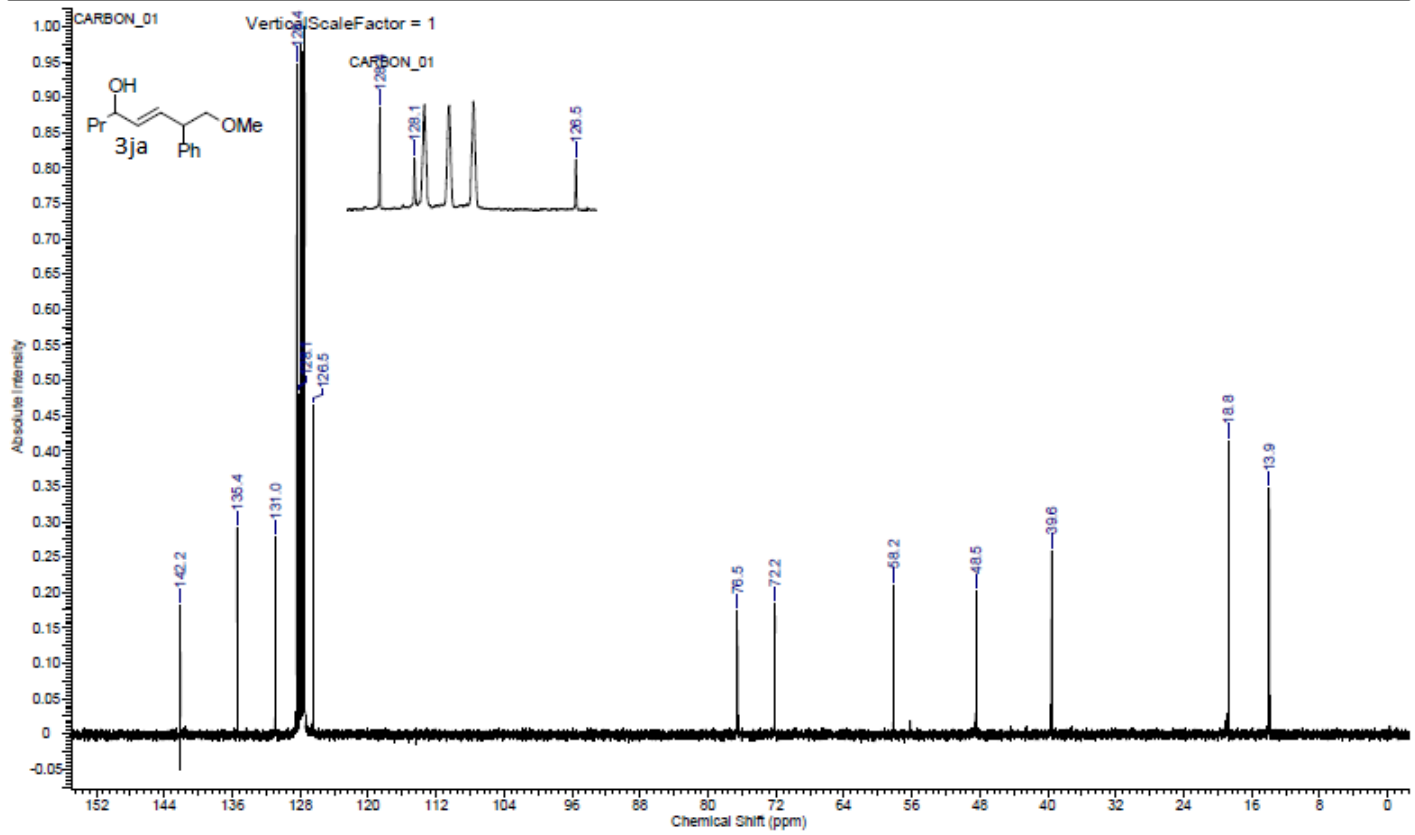
Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-214-1_3	Date	May 26 2016
Date Stamp	May 26 2016	File Name	C:\USERS\ERMAN\DESKTOP\WET\ERM-ENEP-214-1_3_20160526_01\CARBON_01.FID\FID		
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2nu1	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



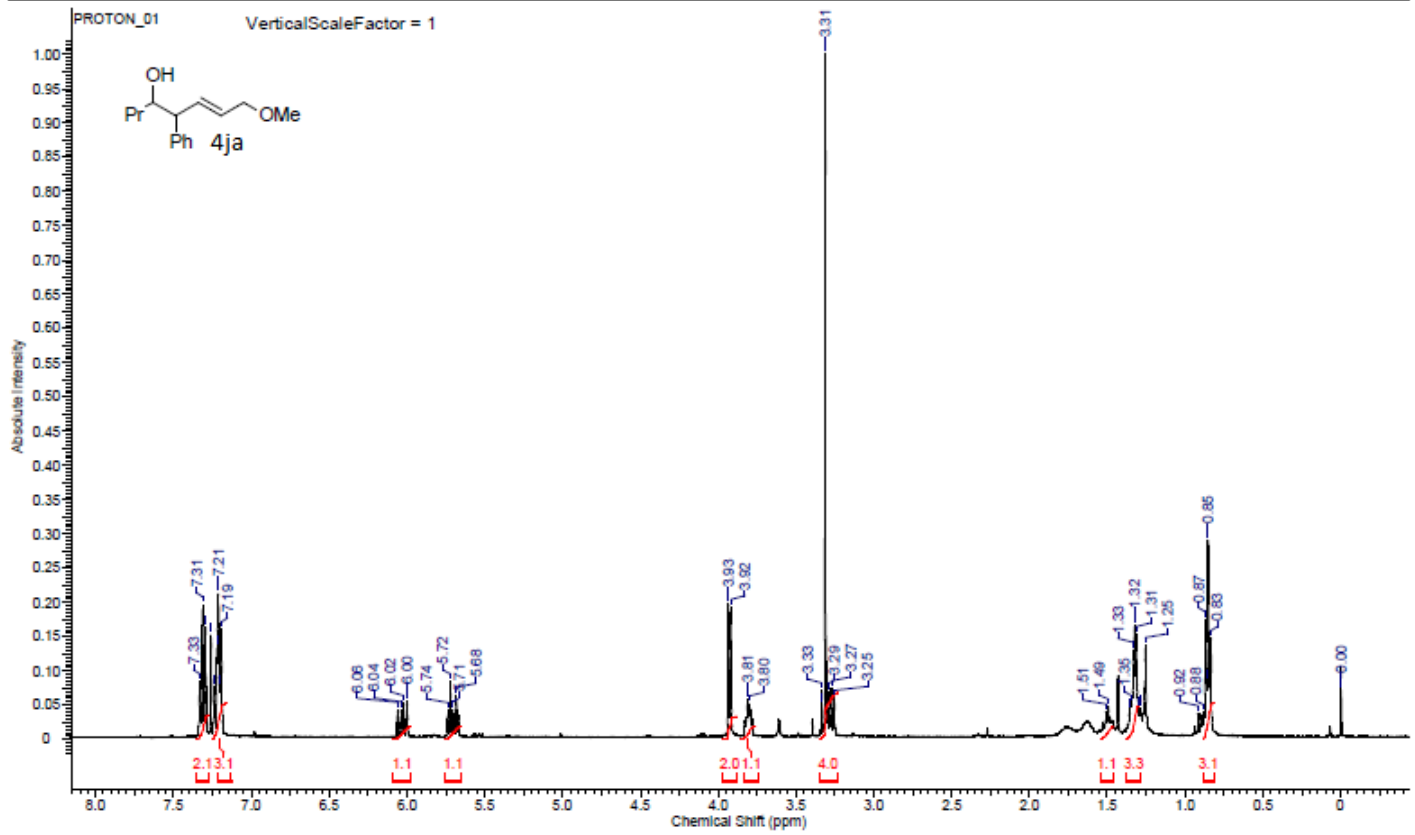
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-219-1_3	Date	Jun 21 2016
Date Stamp	Jun 21 2016	File Name	C:\USERS\ERMAN\DESKTOP\WET\ERM-ENEP-219-1_3_20160621_02\PROTON_01\FID\FID		
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8
Points Count	16384	Pulse Sequence	s2nu1	Receiver Gain	52.00
Spectrum Offset (Hz)	2378.0400	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26
				Solvent	BENZENE-d6
				Temperature (degree C)	25.000



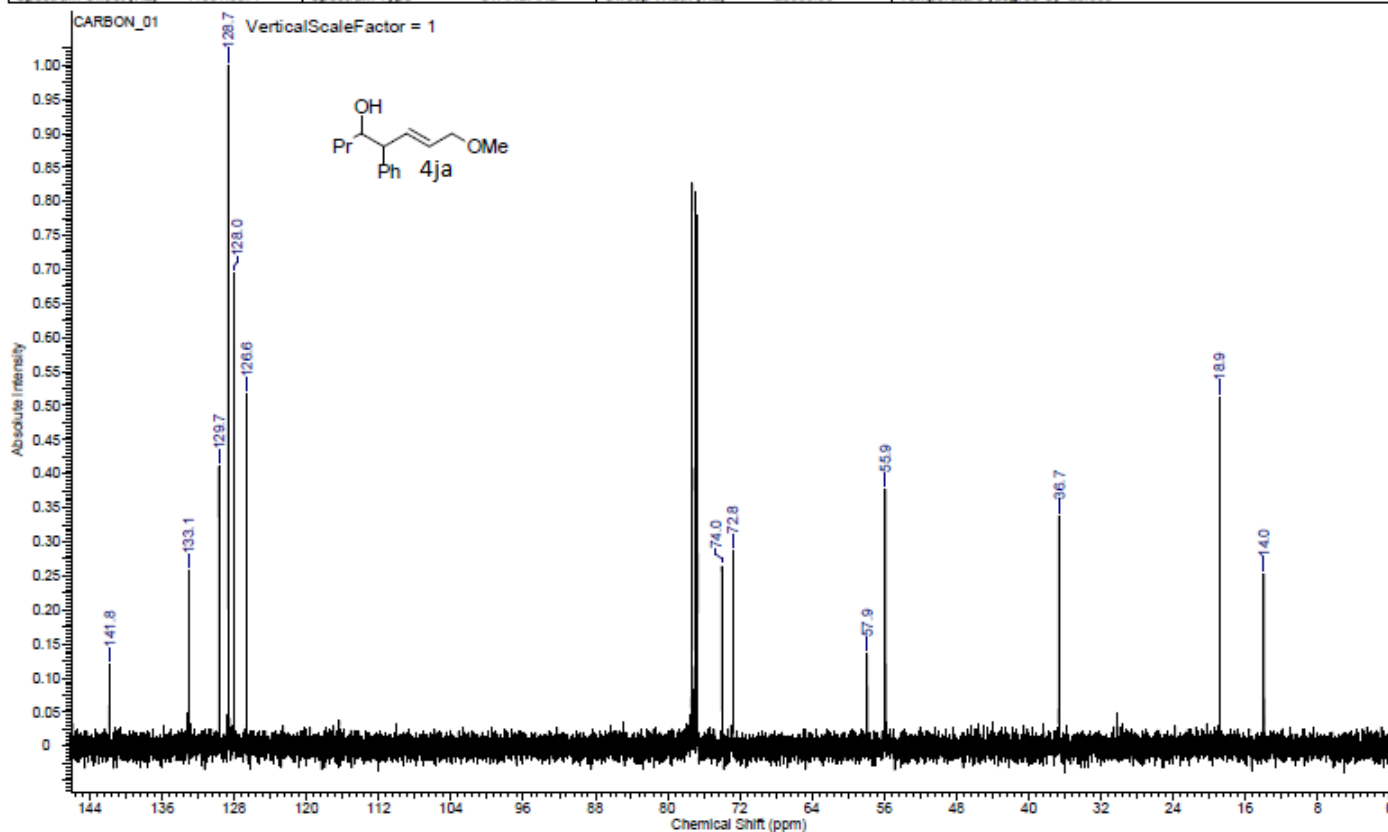
Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-219-1_3	Date	Jun 22 2016
Date Stamp	Jun 22 2016	File Name	C:\USERS\ERMAN\DESKTOP\WET\ERM-ENEP-219-1_3_20160621_02\CARBON_01.FID\FID		
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256
Points Count	32768	Pulse Sequence	s2pu1	Receiver Gain	30.00
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
				Temperature (degree C)	25.000



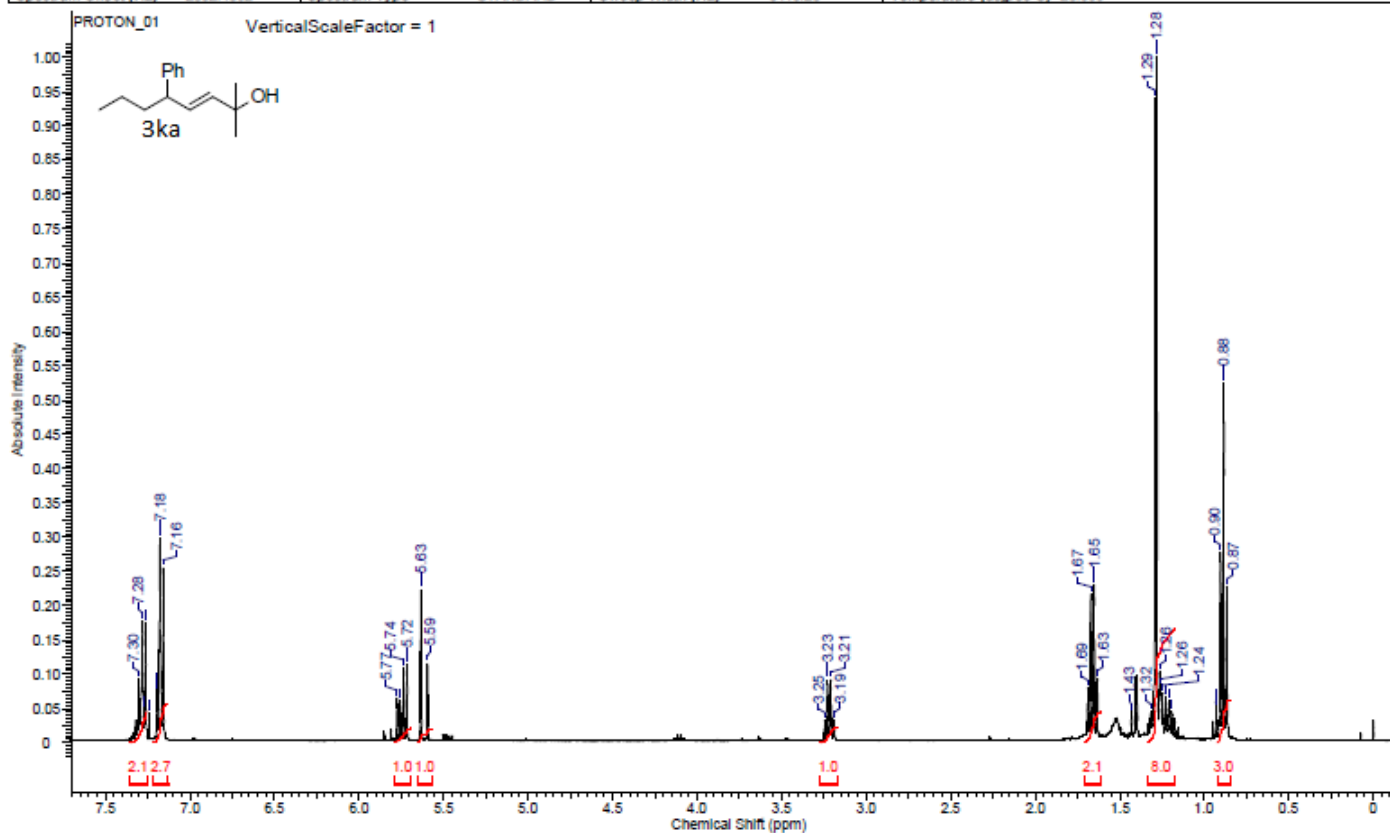
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Date Stamp	Jun 22 2016	File Name	C:\USERS\ERMAN\DESKTOP\WET\ERM-ENEP-219-1_1_20160622_01\PROTON_01\FID\FID				
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2nu1	Receiver Gain	60.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



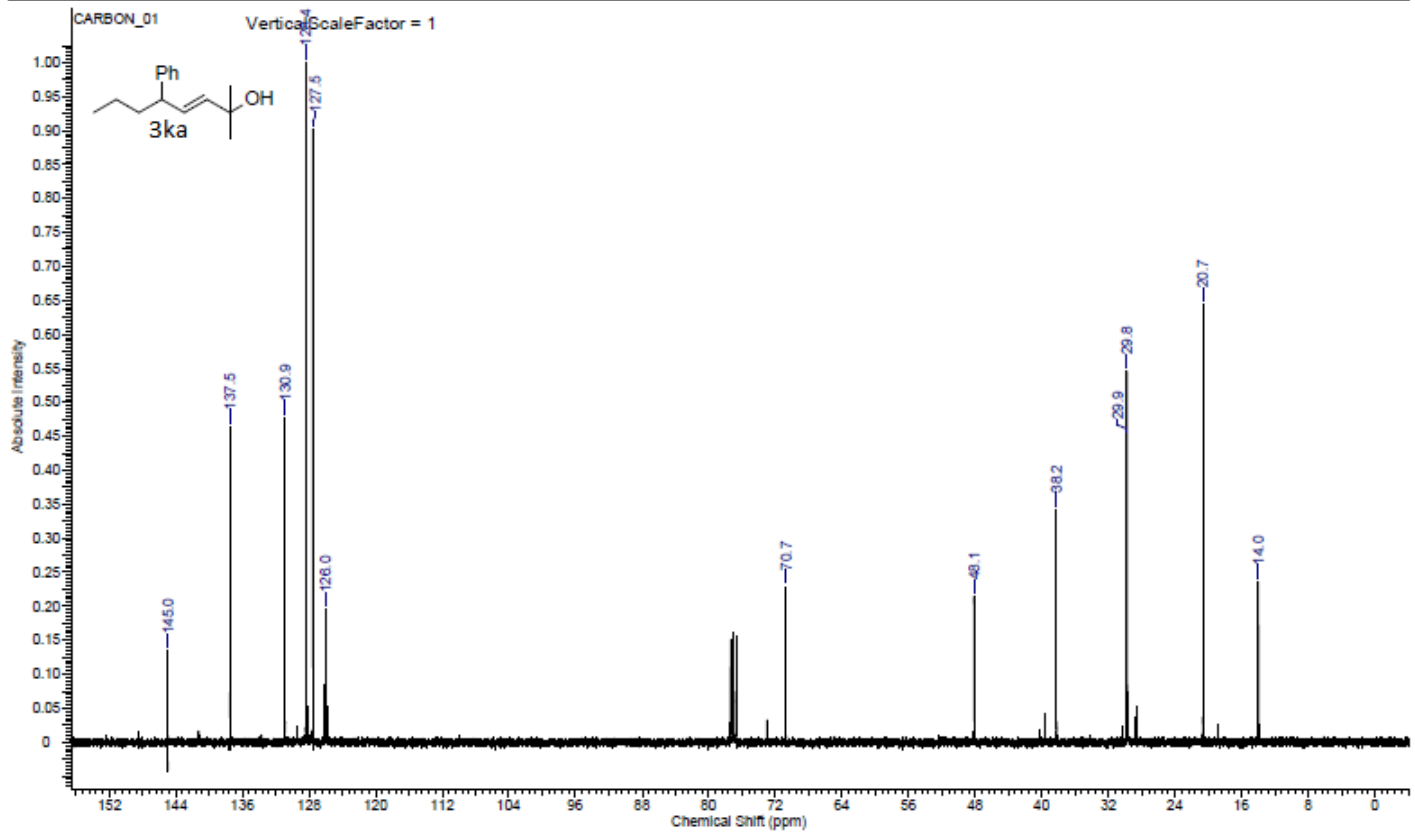
Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-219-1_1		Date	Jun 22 2016	
Date Stamp	Jun 22 2016	File Name	C:\USERS\ERMAN\DESKTOP\WET\ERM-ENEP-219-1_1_20160622_01\CARBON_01.FID\FID				
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	512	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pu1	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



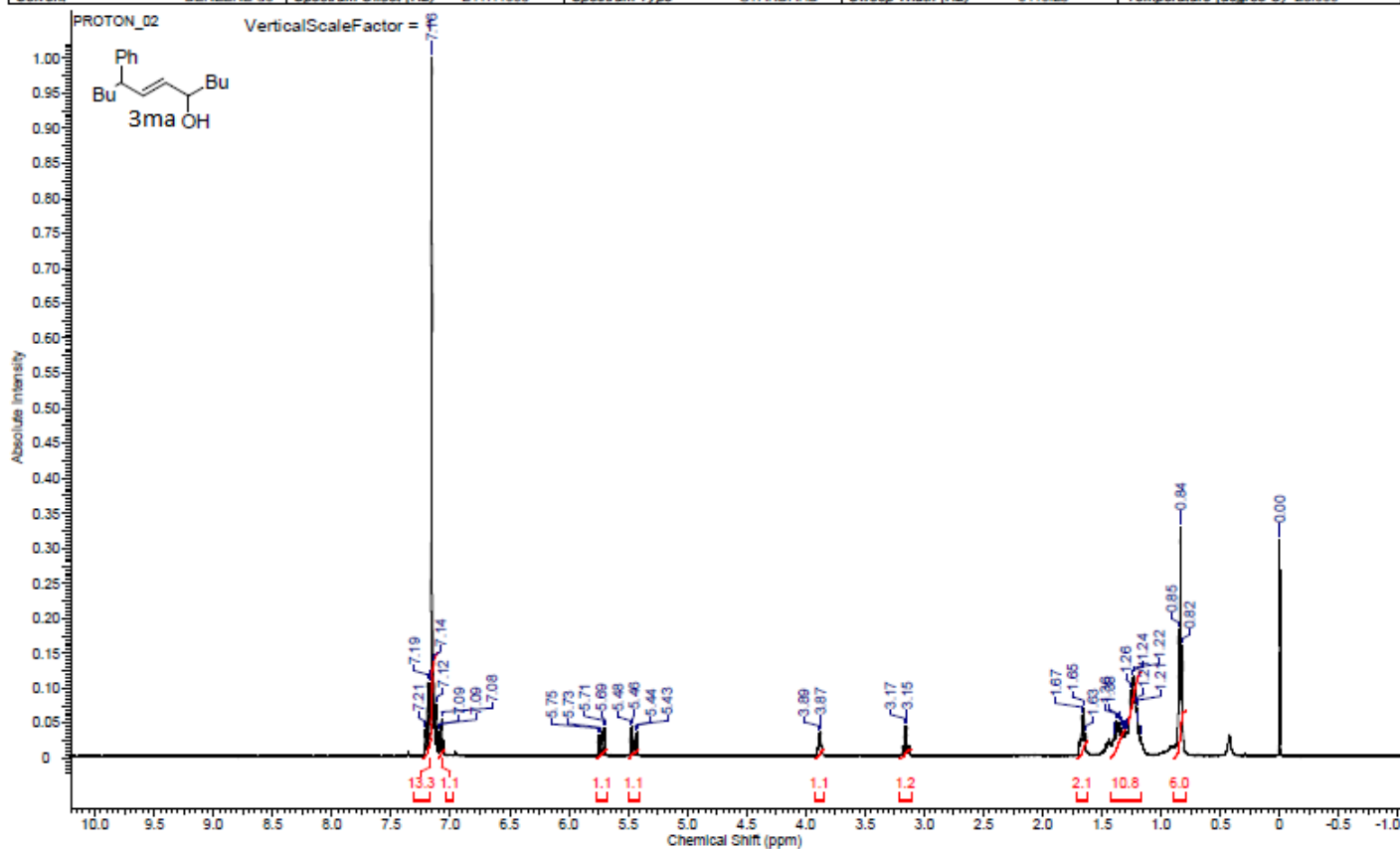
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-218-1_3		Date	Jun 17 2016	
Date Stamp	Jun 17 2016	File Name	C:\USERS\ERMANDESKTOP\WET\ERM-ENEP-218-1_3_20160617_01\PROTON_01\FID\FID				
Frequency (MHz)	399.92	Nucleus	¹ H	Number of Transients	8	Original Points Count	16384
Points Count	16384	Pulse Sequence	s2nu1	Receiver Gain	50.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2392.4502	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



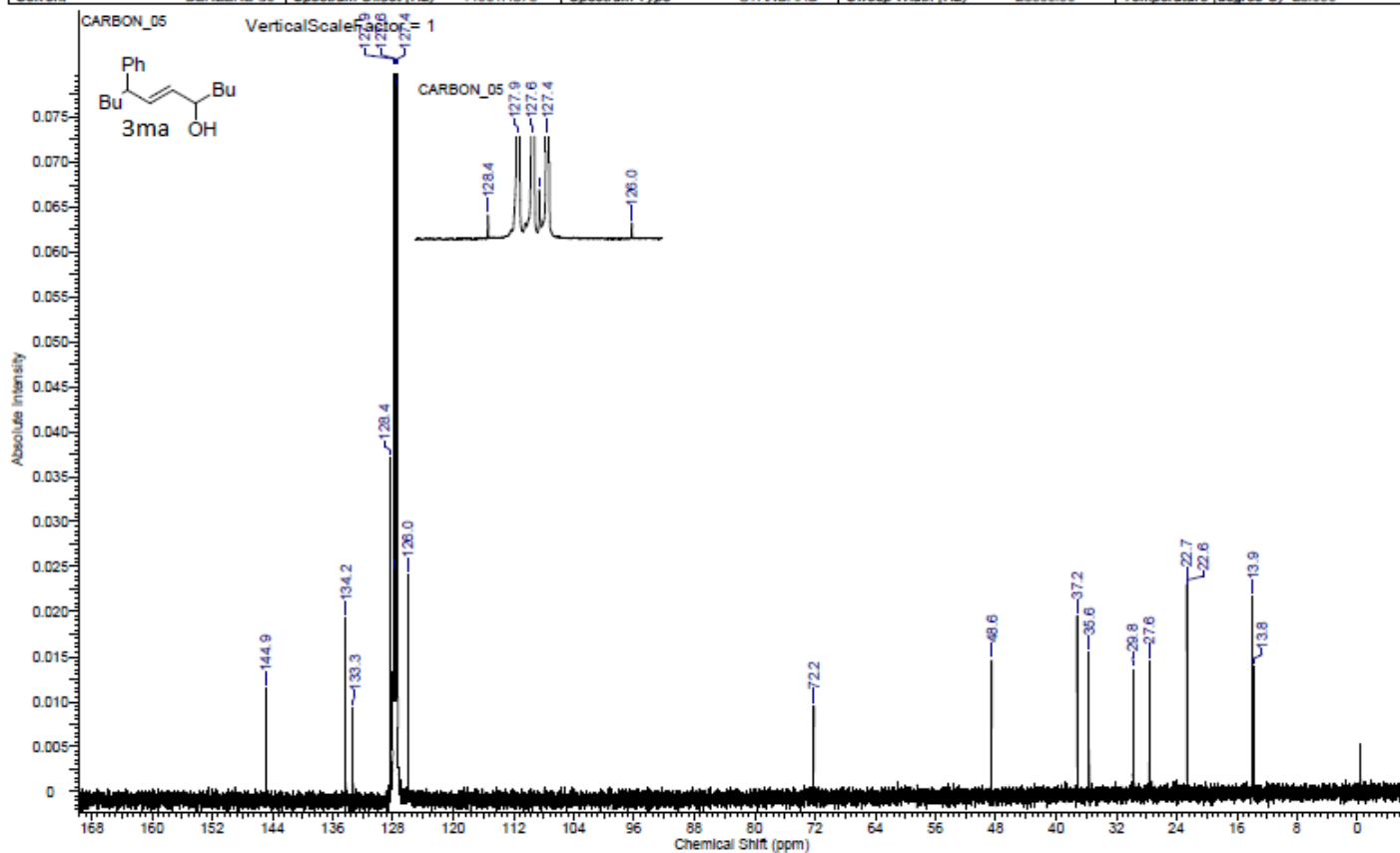
Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-218-1_3		Date	Jun 17 2016	
Date Stamp	Jun 17 2016	File Name	C:\USERS\ERMAN\DESKTOP\WET\ERM-ENEP-218-1_3_20160617_01\CARBON_01.FID\FID				
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	256	Original Points Count	32768
Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00	Temperature (degree C)	25.000



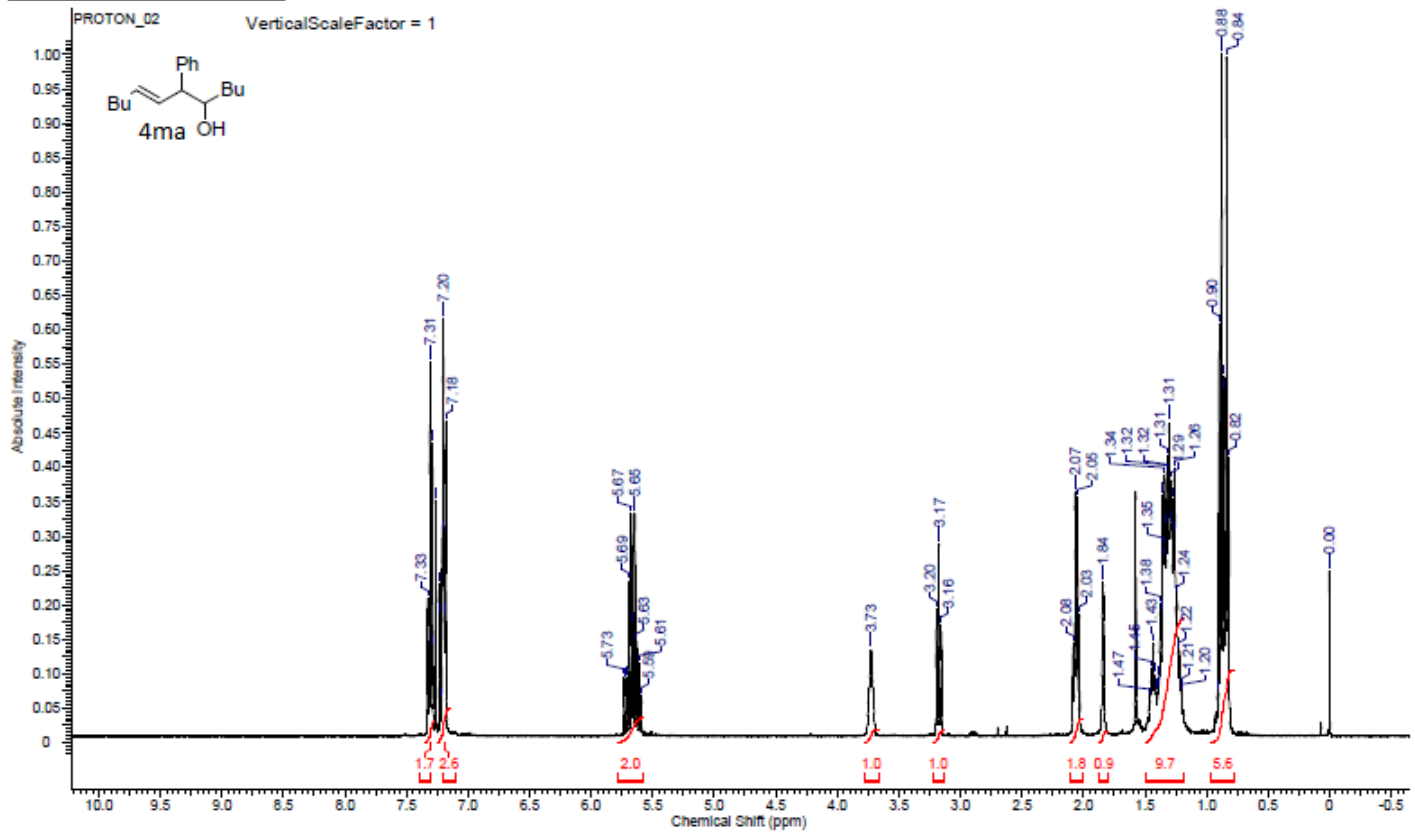
Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-224-1 3-TMZ2		Date	Jul 22 2016	Date Stamp	Jul 22 2016	
File Name	F:\ERM-ENEP-224-1 3-TMZ2 20160722 01\PROTON_02.FID\FID	Frequency (MHz)	399.92		Nucleus	1H			
Number of Transients	8	Original Points Count	16384	Points Count	16384	Pulse Sequence	s2nu1		
Solvent	BENZENE-d6	Spectrum Offset (Hz)	2417.1006	Spectrum Type	STANDARD	Sweep Width (Hz)	6410.26	Temperature (degree C)	25.000



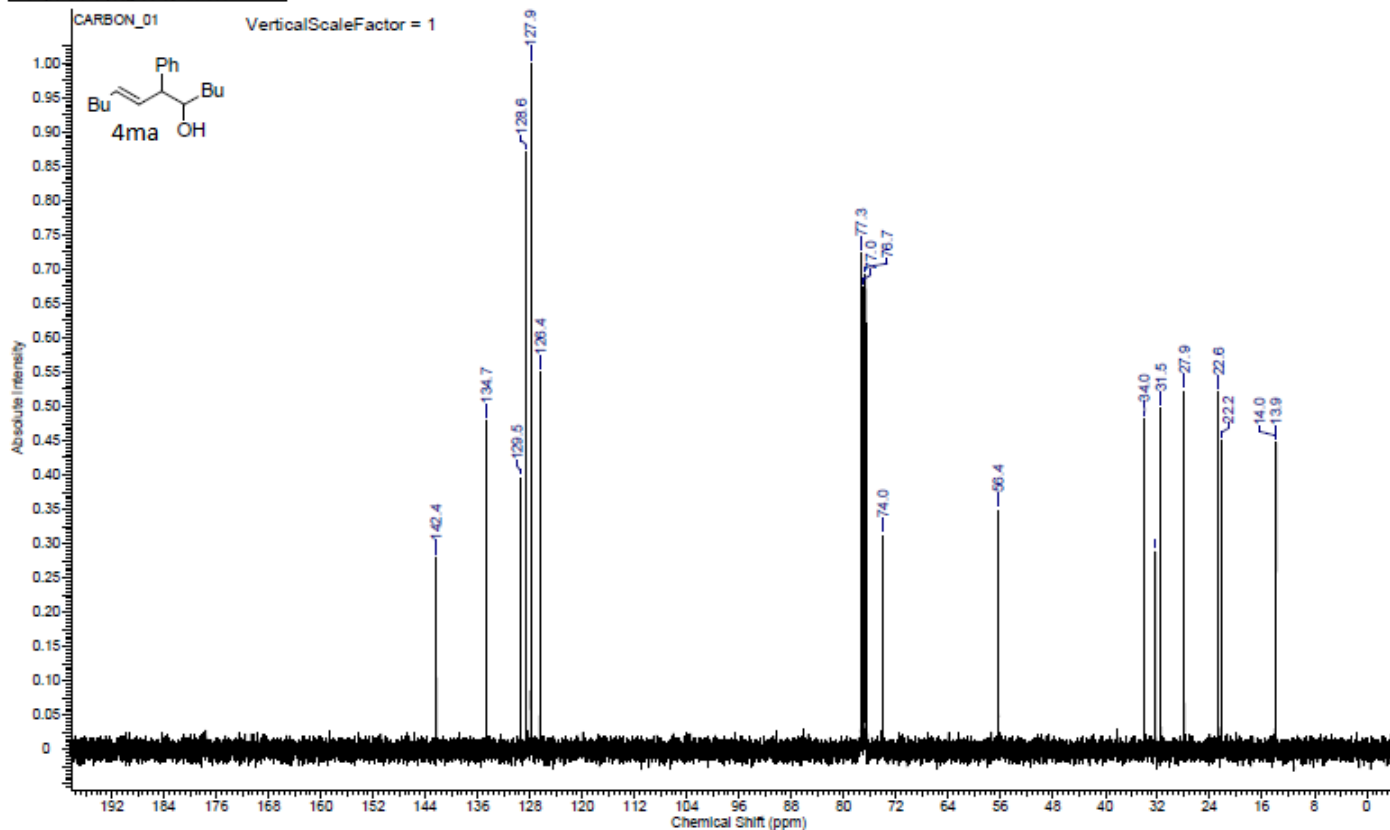
Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-224-1_3	Date	Jul 22 2016	Date Stamp	Jul 22 2016
File Name	F:\ERM-ENEP-224-1_3_20160722_01\CARBON_05.FID\FID			Frequency (MHz)	100.57	Nucleus	¹³ C
Number of Transients	10000	Original Points Count	32768	Points Count	32768	Pulse Sequence	s2pul
Solvent	BENZENE-d6	Spectrum Offset (Hz)	11061.4873	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
						Temperature (degree C)	25.000



Acquisition Time (sec)	2.5559	Comment	ERM-ENEP-224-1.1	Date	Jul 18 2016	Date Stamp	Jul 18 2016
File Name	F:\ERM-ENEP-224-1.1\20160718_01\PROTON_02.FID\FID	Frequency (MHz)	399.92	Nucleus	1H		
Number of Transients	8	Original Points Count	16384	Points Count	16384	Pulse Sequence	s2pul
Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	2399.5020	Spectrum Type	STANDARD	Receiver Gain	60.00
Temperature (degree C)	25.000					Sweep Width (Hz)	6410.26



Acquisition Time (sec)	1.3107	Comment	ERM-ENEP-224-1.1	Date	Jul 18 2016	Date Stamp	Jul 18 2016
File Name	F:\ERM-ENEP-224-1.1.20160718_01\CARBON_01.FID\FID	Points Count	32768	Frequency (MHz)	100.57	Nucleus	¹³ C
Number of Transients	512	Original Points Count	32768	Pulse Sequence	s2pul	Receiver Gain	30.00
Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	11061.5371	Spectrum Type	STANDARD	Sweep Width (Hz)	25000.00
Temperature (degree C)	25.000						



APPENDIX C

MASS SPECTRUMS OF PRODUCTS

